



Review

# A Critical Review on the Role of Food and Nutrition in the Energy Balance

Simona Bo <sup>1,\*</sup>, Maurizio Fadda <sup>2</sup>, Debora Fedele <sup>2</sup>, Marianna Pellegrini <sup>1</sup>, Ezio Ghigo <sup>1</sup> and Nicoletta Pellegrini <sup>3</sup>

- Department of Medical Sciences, University of Turin, 10126 Turin, Italy; mariannapellegrini87@gmail.com (M.P.); ezio.ghigo@unito.it (E.G.)
- Dietetic and Clinical Nutrition Unit, S. Giovanni Battista Hospital, Città della Salute e della Scienza, 10126 Turin, Italy; maurizio.fadda@unito.it (M.F.); d.fedele85@gmail.com (D.F.)
- Department of Agricultural, Food, Environmental and Animal Sciences, University of Udine, 33100 Udine, Italy; nicoletta.pellegrin@uniud.it
- \* Correspondence: simona.bo@unito.it; Tel.: +39-11-633-6036

Received: 14 February 2020; Accepted: 19 April 2020; Published: 22 April 2020



Abstract: The mass media has increasingly frequently suggested to the general population that specific foods or nutritional schemes are able to affect both human metabolism and energy expenditure, thus facilitating weight loss. This critical review is aimed at assessing available evidence on the roles of nutrients, food and dietary regimens in energy intake and energy expenditure. We queried the National Library of Medicine, the Cochrane Library, Excerpta Medica dataBASEand the Cumulative Index to Nursing and Allied Health Literature database, and a search strategy was performed by using database-specific subject headings and keywords. We found that available scientific evidence on these topics is scarce, and that the limited number of available studies often have poor methodological quality. Only a few foods show beneficial effects on metabolism and energy expenditure, as the human energy balance is complex and multifactorial. Finally, microbiota may interfere with the intake, use and expenditure of energy in the human body. Conclusive evidence is still lacking, and, at present, it is not possible to identify a food or a diet with a significant impact on human energy expenditure.

Keywords: energy expenditure; energy balance; obesity; fat burners

#### 1. Introduction

A large amount of misleading news has circulated on social media, blogs, TV and magazines about human nutrition. A specific food or nutrient is often presented as a cure for one or more pathologies, ranging from diabetes mellitus to cancer or Alzheimer's disease [1]. A great amount of information without scientific reliability relative to the treatment of overweightness/obesity is available, a topic in which myths and presumptions are very common [2]. Comprehension of the individual energy balance is particularly complex, owing to physiological compensation to changes in energy intake and/or expenditure [3]. Social media, the Internet, TV and magazines frequently propose direct-to-consumer "information" about food, dietary schemes or supplements which increase the energy expenditure and/or burn fats or, otherwise, reduce the energy expenditure and lead to fat accumulation. However, most of these advertisements contain mis- or dis-information. Some examples include: "drink a lot and consume fat-burning foods" (e.g., pineapple, ginger, onion, avocado, asparagus, celery, chili, broccoli, green tea, garlic, etc.) and "avoid the foods that make you fat" (e.g., pasta, bread and foods containing gluten, oil, dairy products, etc.), in order to lose weight [4]. All these suggestions are generally incorrect: there are no foods with negative calories and focusing on one or a few foods or nutrients does not work, as a multifaceted and individualized program with careful follow-up over

Nutrients **2020**, 12, 1161 2 of 27

time is required to lose weight [5]. This kind of mis-/dis-information is particularly concerning, owing to its influence on the general population, and such wrong beliefs have been found to be hard to correct, especially in people with lower cognitive ability [6].

The aim of the present paper is to critically review the available evidence about the roles of nutrients, food, and dietary regimens on energy intake and energy expenditure, taking into consideration all the conditions potentially impacting on the final energy balance, including the gut microbiota. In particular, we analyzed the following topics:

- (i) The energy balance in humans;
- (ii) Energy intake from food;
- (iii) Energy expenditure due to food intake;
  - the role of nutrients;
  - the role of foods;
  - the role of diet plans;
- (iv) The impact of the gut microbiota on the human energy balance.

#### 2. Methods

The following databases were queried: PubMed (National Library of Medicine), the Cochrane Library, Excerpta Medica dataBASE (EMBASE) and the Cumulative Index to Nursing and Allied Health Literature (CINAHL). The search strategy was performed by using database-specific subject headings and keywords (i.e., energy, energy expenditure, energy balance, energy intake, caloric intake, diet-induced thermogenesis, thermogenesis, plus the specific nutrients/foods/diets or gut microbiota). No restrictions were placed. Hand-searching the references of the studies and reviews of the field was performed to augment the search strategy. To search for toxicity information (of single foods), the following terms were used: toxicity, adverse events, adverse effects, side effects, reactivity and interactions.

Few papers were available about many topics; therefore, all the research articles were considered with the following scale of priority: systematic reviews and meta-analyses, randomized controlled trials (RCTs), human observational studies, case series, animal studies and in vitro studies.

#### 3. The Energy Balance in Humans

In humans, energy intake (EI) and energy expenditure (EE) are in a complex balance, resulting from the difference between EI and EE, aimed at maintaining a relatively constant level of energy stores over time in accordance with the principle of energy conservation [3]. When EI is reduced, a corresponding reduction in EE occurs (and vice versa), in order to minimize perturbations to energy homeostasis [7]. Energy intake is derived from dietary macronutrients (proteins, carbohydrates and lipids) and alcohol. The absorption of calories depends both on food and individual characteristics [3]. A high degree of overfeeding was associated with a greater fractional decrease in stool energy loss in lean but not in obese individuals, thus indicating that the degree of overnutrition relative to individual weight-maintaining energy needs may play a role in the determination of the efficiency of nutrient absorption [8]. Daily total energy expenditure (TEE) can be split into different components: resting energy expenditure (REE), which is the energy required to support body's basic metabolic activities; activity-induced energy expenditure (AEE), the energy cost of physical activity and exercise; diet-induced thermogenesis (DIT), the energy spent to process the ingested food (about 10%-15% of TEE); and the energy necessary for body thermoregulation. REE can be 3%–10% higher than basal energy expenditure (BEE), which is the energy required to maintain vital body functions [9–11]. REE is mostly determined by body size and composition and is positively correlated with body weight and fat-free mass. AEE is the most variable component of TEE depending on an individual's lifestyle [12,13]. Food intake affects all the

Nutrients **2020**, 12, 1161 3 of 27

components of TEE—but predominantly DIT—with different effects according to the macronutrient composition of a meal and daily variation within the same individual [3,14].

Energy homoeostasis is fundamental for survival and, hence, highly specialized adaptive mechanisms counteract energy imbalances, making energy balance a complex process. Adaptive thermogenesis (AT) and facultative thermogenesis (i.e., the heat production in response to environmental variations) both protect an organism from exposure to cold and regulate the energy balance after dietary changes, and are influenced by the activity of the sympathetic nervous system (SNS), leptin and many hormones (mainly 3,5,3'-tri-iodothyronine) [15,16]. A major site of AT is the brown adipose tissue (BAT), where non-shivering thermogenesis occurs with the uncoupling of mitochondrial substrate oxidation from adenosine triphosphate (ATP) production and the release of fatty acid oxidation energy as heat [17,18]. BAT is activated not only by cold exposure, but also by certain food ingredients, thus contributing to DIT [19,20]. The same signals activating BAT also induce the expression of uncoupling protein 1 (UCP1) in white adipose tissue (WAT) cells (the "beige" cells), a phenomenon known as browning [21].

Either energy restriction or overfeeding induce adaptive changes in the energy balance, with AT, respectively directed towards energy sparing or vice versa. Reducing habitual energy intake by about 10% reduces TEE by 10%–15%, mainly due to reduced REE; furthermore, AT can explain about 50% of the less-than-expected weight loss in patients with obesity [16,22].

## 4. Energy Intake from Food

The actual energy content of some foods may differ from the energy, which is theoretically calculated, due to differences in macronutrient digestibility and food structure [23,24]. One of the best examples of this discrepancy is represented by nuts. Herein, we shortly describe the energy of this paradigmatic food.

Tree nuts are energy-dense foods, due to their high content of lipids (ranging between 40–75 g per 100 g) [25]. However, the inclusion of nuts as part of a healthy diet does not affect body weight, as reported by observational and experimental studies, even though nuts may benefit weight-loss diets [24]. Several mechanisms have been proposed to explain this discrepancy, including appetite control, increased DIT (as discussed below), and discrepancies in available metabolizable energy (ME, i.e., the amount of the food available energy to the human organism) [24]. To calculate the food ME, each energy-contributing food component is multiplied by its Atwater factor [23]. However, recent evidence has demonstrated that the Atwater factors do not provide accurate ME values for several nuts in healthy volunteers [26-29]. Indeed, based on the measurements on urine and feces, ME values were found to be 25%, 20%, 16% and 5% less than those calculated for almonds, walnuts, cashews and pistachios, respectively. The reason for this discrepancy is partly due to the structure of nuts, which limits the accessibility of digestive enzymes. In oilseeds, such as nuts, lipids are stored in oil bodies which are covered by a thin layer of phospholipids and proteins and encapsulated in cell walls [30], whose components (e.g., cellulose, hemicellulose, peptic substances and lignin) are mainly indigestible by human digestive enzymes [23]. After nut mastication, large particles representing clusters of intact cells remain, which provide protection against disintegration and a physical barrier for enzyme hydrolysis and microbiota metabolism [31]. These clusters of cells, with intracellular lipids encapsulated within the cell walls, were still intact after having passed through the human intestine, thus reducing the intake of energy. Furthermore, it has been demonstrated that when almonds were chewed 10 times, a higher number of larger particles was obtained than when they were chewed 25 or 40 times [32]. These large particles retain more energy and lipids (which are then lost in the stool) than smaller ones (43.7%, 32.7% and 30.8% of the lipid load was lost in the stool after 10, 25 and 40 chews, respectively). On the other hand, other processes (such as roasting) make almonds more brittle and crunchy, with the subsequent production of smaller particles after mastication [33] and the induction of swelling of the cell walls with increased porosity and destruction of oil bodies, favoring the access of digestive enzymes [30]. These changes slightly increased the measured ME

Nutrients **2020**, 12, 1161 4 of 27

of roasted almonds, compared to whole almonds, even though their ME was still lower than that predicted with the Atwater factor (-25% and -19% for whole natural almonds and whole roasted almonds, respectively) [34]. In almond butter, where the cellular structure is fully destroyed, there is a full release of energy, with no discrepancy between the measured ME and the predicted energy content. Similarly, fecal fat content was significantly higher when 70 g of whole peanuts were consumed in healthy adults, compared to other forms of peanuts (i.e., oil, butter and flour) [35].

The effect of structure on the actual energy content of foods has been shown mainly for nuts, but the same effect may be extended to other seeds, legumes and some cereals. This lower actual energy content may have an impact on the overall energy intake when a diet is rich in unprocessed foods where the food structure is retained [23].

## 5. Energy Expenditure Due to Food Intake

## 5.1. The Role of Nutrients

Food intake stimulates energy expenditure; this is a well-known phenomenon, called DIT or the thermic effect of food. DIT accounts for ~10%–15% of TEE, which is a meaningful amount of the human body daily energy expenditure [36] and which can be measured by indirect calorimetry through the assessment of oxygen consumption and carbon dioxide production [37]. However, this method of measurement based on respiratory exchange has been recently blamed for overestimating DIT, as it is based on the assumption that all metabolic processes of the organism consume oxygen and produce carbon dioxide, which is not always true [38]. Both insulin resistance and, to a lesser extent, abdominal adiposity, have an impact on DIT by reducing the thermic effect of a meal [39]. In fact, insulin, by increasing glucose oxidation and inhibiting lipid oxidation, regulates the cellular substrate flow and utilization, which is therefore impaired in the presence of abnormal insulin sensitivity [40].

At present, there is great interest in the possibility of modulating DIT in order to increase the body's energy expenditure and promote weight loss. First of all, DIT has been proven to be influenced by meal timing, with DIT being higher in the morning and reduced in the evening [41]. Increased nocturnal insulin-resistance and heightened ghrelin levels, slower evening gastric emptying with increased carbohydrate absorption, and increased morning sympathetic activity have been proposed as possible explanations [42]. In addition to meal timing, DIT is influenced by the caloric content of a meal and increases in a direct proportion to the energy intake [14,43]. Finally, the macronutrient composition of food seems to meaningfully affect post-prandial energy expenditure, even if the data in the literature are controversial. Commonly, proteins have been considered to induce an increased energy expenditure which, combined with a higher satiating effect, could determine a higher weight loss [39,44]. On the other hand, carbohydrates and lipids determine a lower DIT than proteins (protein > carbohydrates > lipids) [45,46]. Meals with protein percentages ranging from 11%–30% of the total calories proportionally increase DIT until the value of 30%, where a plateau is reached and a subsequent increase in the protein intake does not increase further the thermic effect of the meal [47]. The protein source should be taken into account, as well: casein, soy or whey proteins are metabolized differently, which may explain the variability in the speed and extent of DIT increase. In particular, whey proteins lead to higher DIT than caseins, while contrasting results have been obtained in the comparison of whey and soya proteins [48]. Regarding the quality of other nutrients, medium-chain lipids seem to heighten DIT more than long-chain triglycerides [49,50] and unsaturated fats more than saturated, probably due to up-regulation of proliferator-activated receptor (PPAR)- $\alpha$  expression [51]. Finally, unrefined, fiber-rich carbohydrates determine an increased energy expenditure, especially if contained in low-processed foods [39].

Overall, very few data are available about this topic and the clinical significance of any single nutrient or single meal is unclear in a weight-loss strategy.

Nutrients **2020**, 12, 1161 5 of 27

#### 5.2. The Role of Foods

An increasing number of foods are supposed to increase human energy expenditure [52]. The list is long and is gradually getting longer (Supplementary Table S1). Herein, we examined those for which scientific studies were available (Figure 1).

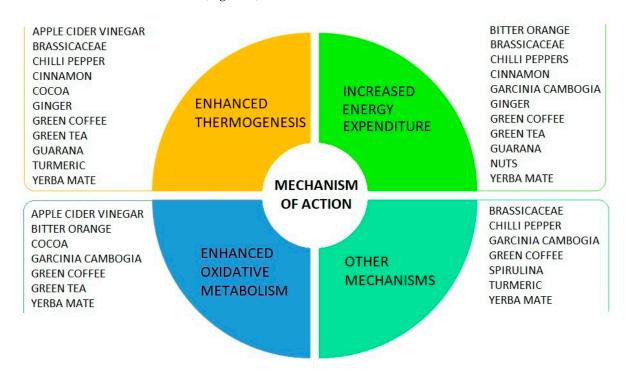


Figure 1. Supposed mechanisms of action of food impacting on energy balance.

#### 5.2.1. Green Coffee

#### Available Evidence

Almost the whole world's coffee consumption derives from the beans of two coffee plants—Coffea canephora and Coffea arabica—which contain many bioactive compounds, such as caffeine (1,3,7-trimethylxanthine) and chlorogenic acid [53]. Green (unroasted) coffee and roasted coffee contain the same amount of caffeine (1.2%-2.2%) but a different percentage of chlorogenic acids (6.5%-10% vs 2.7%-3.1%, respectively) [54]. Caffeine increases thermogenesis and energy expenditure by several mechanisms [55,56]. In humans, the thermic effect lasts about 150 min after a single-dose caffeine ingestion [56] and one RCT reported a stronger metabolic impact among habitual low consumers of caffeine, thus suggesting the possibility of a long-term insensitivity to the effects of caffeine after high and prolonged exposure [57]. In human trials, an increase in energy expenditure has been reported, varying from 6% (after 50 mg caffeine intake) [58] to 7% (after 200 mg caffeine consumption) [57]. Chlorogenic acid has been reported to have beneficial effects against obesity and other dysmetabolic disorders, as well as playing a favorable role in energetic metabolism in both human and animal studies [59,60]. In a pilot study, the consumption of 1 cup of green coffee (containing 6 mg caffeine per kg of lean body mass, about 215–280 mg) determined an increase of REE by 6.4% at 30 min and 2.2% at 180 min, with a positive correlation between the chlorogenic acid assumed and the REE values at 30 min [61].

## Molecular Mechanisms of Action

In cultured adipocytes, caffeine has been shown to enhance BAT function and thermogenesis by up-regulating UCP1 and BAT-selective regulatory genes including PPAR- $\gamma$ , PPAR- $\gamma$  coactivator (PGC)-1 $\alpha$  and PR domain containing 16 (PRDM16) [55]. PGC-1 $\alpha$  also induces mitochondrial biogenesis

Nutrients **2020**, 12, 1161 6 of 27

and stimulates fatty-acid oxidation and oxygen consumption through the co-activation of PPAR- $\gamma$  [55]. Caffeine induces PGC1 $\alpha$  and UCP1 indirectly as well, by antagonizing the transient receptor potential vanilloid (TRPV)-4, a negative regulator of PGC-1 $\alpha$  in the TRPV receptor family and a modulator of beige/brown adipocyte thermogenesis [55]. Adipose tissue browning is additionally stimulated through the expression of other specific genes (*CD137*, *LHX8*, *P2RX5*, *CITED1* and *COX8b*) [55]. Further mechanisms have been implicated in the caffeine-induced thermogenesis, such as the antagonism of adenosine-mediated inhibition of the secretion of epinephrine and norepinephrine and the inhibition of phosphodiesterase, which increases intracellular levels of cyclic adenosine monophosphate (cAMP). Catecholamines stimulate  $\beta$ -adrenergic receptors and cAMP activates the protein kinase A, which enhances UCP1 activity through increased free fatty acid release [55,58,62]. Chlorogenic acid principally up-regulates the AMP activated protein kinase (AMPK) with increased fatty acid oxidation and ATP production [63].

# Toxicity and Reactivity

According to the Food and Drug Administration (FDA), European Food Safety Authority (EFSA) and Health Canada, the consumption of up to 400 mg caffeine can be considered safe in healthy adults, without overt, adverse cardiovascular, behavioral, reproductive, bone and developmental effects [64–66]. No reactivity with drugs, supplements or food has been reported for green coffee.

#### 5.2.2. Green Tea (Camellia sinensis)

#### Available Evidence

The leaves of the plant Camellia sinensis give the three most popular types of tea, green (unfermented), black (fully fermented) and oolong (semifermented) [67]. The main components of green tea are polyphenols, in particular flavon-3-ols, also known as catechins, mostly epigallocatechin-3-gallate (EGCG) [68]. Caffeine is naturally contained in green tea as well, in a variable amount, according to the brewing period and the tea and water rate [68]. The fermentation process lowers the content of polyphenols and increases caffeine in tea; green tea contains two times more catechins, but 2–3 times less caffeine than black tea [68,69]. Indeed, most of the studies available on the effects of tea on the energy balance are related to green tea. Many in vitro and animal studies have documented enhanced thermogenesis, heightened energy expenditure and fat oxidation after green tea consumption [68,70,71]. Other beneficial effects are a reduction in fat mass due to the interruption of lipid emulsification, the inhibition of gastrointestinal digestive enzymes activity, improvements in the gut microbiota and reduction in adipocyte differentiation and food intake [69,71,72]. Both human observational studies and trials have confirmed increased energy expenditure after the acute administration of green tea, but the long-term effects are not currently proven [69,73–76]. It is noteworthy that in most of the trials, the volunteers were provided with high doses of green tea catechins, equivalent to 3-4 cups of brewed green tea a day [72], which are usually consumed only by a few population groups.

#### Molecular Mechanisms of Action

Catechins and caffeine (see the previous paragraph) affect energy expenditure differently. Catechins inhibit the catechol-O-methyl transferase enzyme (COMT) in almost all the tissues, which, in turn, inhibits the degradation of norepinephrine and produces protracted  $\beta$ -adrenergic stimulation. Hence, the SNS activity is increased along with energy expenditure and fat oxidation [68,72,73]. In vivo and in vitro, EGCG has been shown to affect energy expenditure by the activation of AMPK, which promotes fatty acid oxidation and ATP production [69,72]. EGCG has also been reported to inhibit mitochondrial oxidative phosphorylation, up-regulate the gene expressions of UCPs in BAT and decrease ATP levels, which activates AMPK [69,72,74]. In vitro, fermented green tea induced the up-regulation of fatty acid oxidation-related genes and increased energy expenditure by inducing serotonin secretion [70].

Nutrients **2020**, 12, 1161 7 of 27

## Toxicity and Reactivity

Green tea is safe across a wide range of intakes and preparations, however concentrated solid extracts are less tolerated due to the high content of EGCG [77]. Gastrointestinal symptoms, such as nausea/vomiting, diarrhea, flatulence, abdominal bloating and dyspepsia have been reported after the intake of high doses of beverages or extracts (corresponding to 5–6 L of beverage/day) [76]. The intake of 10–29 mg/kg/day of green tea-based dietary supplements has resulted in liver toxicity due to oxidative stress and cytotoxic damage [67]. The caffeine content in green tea is low but, depending on self-sensitiveness to methylxanthines and doses, symptoms such as nervousness, restlessness, tremors, palpitations, sleep disorders, vomiting, diarrhea, headaches, epigastric pain and tachycardia have also been reported [67]. In adults an intake of ~300 mg EGCG/day in solid bolus dose and ~700 mg for tea preparations are considered safe [77]. The vitamin K contained in green tea leaves can antagonize the effect of anticoagulants [67].

#### 5.2.3. Cocoa and Dark Chocolate

#### Available Evidence

Cocoa, the main constituent of dark chocolate derives from the *Theobroma cacao* tree. Dark chocolate is considered a functional food, due to its content of fatty acids, vitamins, minerals, fiber, several methylxanthine alkaloids (4% of the dry weight), mainly caffeine and theobromine [78,79] and polyphenols (12%–18% of dry weight); in particular, flavan-3-ols, (+)-catechin and (–)-epicatechin and B-type procyanidins [80–82]. Cocoa has been reported to reduce fatty acid synthesis and transport systems, enhance  $\beta$ -cell function, down-regulate insulin receptor kinase activity, improve peripheral insulin sensitivity, inhibit digestive enzymes and increase thermogenesis in liver and WAT both in animals and humans [83–86]. A meta-analysis of human RCTs reported that cocoa/dark chocolate supplementation do not affect anthropometric measures in adults; however, a subgroup analysis indicated that  $\geq$ 30 g dark chocolate per day for at least 4 weeks had favorable effects on weight and body mass index (BMI) [84]. Studies in mice have reported that procyanidins of cocoa liquor (the pure cocoa mass derived from cocoa beans), in addition to thermogenic effects, have a role in the prevention of postprandial hyperglycemia by increasing glucagon-like peptide-1 activity, phosphorylation of the AMPK $\alpha$  and glucose transporter type-4 translocation in skeletal muscle and BAT [87].

#### Molecular Mechanisms of Action

Cultured WAT cells of cocoa-fed rats have shown the upregulation of the gene expression of UCP2, a homolog of UCP1 implicated in non-shivering thermogenesis [83]. Procyanidins affect energy expenditure by inducing the gene and protein expression of UCPs (UCP1, UCP2 and UCP3), AMPK $\alpha$  and PGC-1 $\alpha$  in adipose, liver and muscle tissues [88]. Once activated, AMPK leads to the inhibition of energy-consuming biosynthetic pathways, such as fatty acid and sterol synthesis and the activation of ATP-producing catabolic pathways, such as fatty acid oxidation [89]. PGC-1 $\alpha$  increases mitochondrial biogenesis and the expression of UCPs, promoting fatty acid oxidation as well [88]. Furthermore, the methylxanthines contained in dark chocolate act as adenosine receptor blockers in vivo [90], affecting energy expenditure by stimulating basal and noradrenaline-stimulated lipolysis in rat fat cells [91]. Xanthine derivatives induce secretion of catecholamines, which bind to adipose cells and increase thermogenesis by increasing the expression of thermogenic genes and releasing free-fatty acids which, in turn, enhances UCPs [92]. Studies in humans are needed to confirm these mechanisms.

# Toxicity and Reactivity

Depending on the percentage of dry cocoa, chocolate may contain trace heavy metals–principally cadmium and lead–resulting from the contamination of the soil or during manufacturing processes [93]. European Legislation has set the levels to 1 mg/kg for cadmium and 0.3 mg/kg for lead as the maximum tolerable amount in cocoa powder [94].

Nutrients **2020**, 12, 1161 8 of 27

#### 5.2.4. Yerba Mate (*Ilex paraguariensis*)

#### Available Evidence

The infusion (mate) derived from the dried leaves of Yerba mate is widely consumed throughout South America as well as in many other countries. Its numerous beneficial effects are likely due to the content of several bioactive compounds, such as polyphenols, alkaloids, soaps, triterpenoids, flavonoids and chlorogenic acid [95,96]. In one placebo-controlled study, an infusion containing 1.5 g mate dry extract increased REE by almost 5% and resulted in a 5% reduction of the respiratory quotient in non-obese women and men, probably through increased lipid oxidation capacity [97]. In humans, pharmacological doses of Yerba mate extracts acutely induced a significant increase in the exercise energy expenditure due to the preferential use of fatty acids as an energy substrate [98]. Chronically, these extracts determined an increase in REE, thermogenesis in WAT and a reduction in the WAT synthesis of fatty acids in mice, leading to weight and fat loss and lower circulating leptin levels [99].

#### Molecular Mechanisms of Action

Chlorogenic acid, as already reported for green coffee, increases fatty acid oxidation by upregulation of AMPK [63] and inhibits adipogenesis by down-regulation of the expression of specific genes, such as *Creb-1* and *C/EBPa* [100]. The increased thermogenic effects after supplementation with Yerba mate extracts seem to be due to increased mitochondrial genesis and expression of UCPs, resulting in greater efficiency in the mitochondrial respiratory chain and heat dissipation in mice fed with high-fat diet [101].

## Toxicity and Reactivity

The metabolic effects of Yerba mate have been obtained by using supplements with high doses of the active compounds, thus not well representing the effect of the natural food for which human data are still lacking. Toxicological investigations in rats have reported a good tolerability of single (up to 2 mg/kg dose) and chronic administration of Yerba mate extracts, which seem to be safe for consumption at dosages up to 300 mg/kg/day in pregnant rats [102,103].

# 5.2.5. Bitter Orange (Citrus aurantium)

# Available Evidence

Citrus aurantium, better known as bitter orange, is an evergreen plant whose fruits have been used for many centuries both as a food in Southern Europe and as a supplement in traditional medicine in China and South America [104,105]. These fruits contain alkaloids -particularly synephrine and octopamine—and other compounds, such as flavonoids -in particular hesperidin, naringin, limonene and tangaretin—with potential beneficial effects on metabolism and health [106,107]. A few human studies have demonstrated both an acute thermogenic effect with a statistically significant increase in REE, DIT and blood catecholamines levels, as well as weight loss and appetite suppression after the ingestion of bitter orange extracts [104,107–110]. However, long-term data are lacking, as well as data about the effects of the consumption of the fruit by itself, as the available studies have employed dry and purified extracts from the orange peel, containing a high dose (~26 mg) of p-synephrine.

## Molecular Mechanisms of Action

Synephrine and octopamine, are contemporary  $\alpha$ - and  $\beta$ -adrenergic agonists which display sympathomimetic effects by contributing to oxidative metabolism, lipolysis promotion and  $\beta$ 3- and  $\alpha$ -adrenergic receptor stimulation [106]. The anti-adipogenic effects of p-synephrine in 3T3-L1 preadipocytes are due to the regulation of the Akt signaling pathway and the suppression of adipogenesis-related proteins [111]. After treatment with *Citrus aurantium*, primary cultured brown

Nutrients **2020**, 12, 1161 9 of 27

adipocytes displayed increased differentiation associated with the elevation of thermogenic factors including UCP1 and PPAR coactivator  $1\alpha$ , by AMPK activation [112].

#### Toxicity and Reactivity

Case reports, as well as animal and human studies, have provided evidence for cardiovascular effects due to the ingestion of high synephrine doses contained in supplements, especially in combination with caffeine [106,109,113]. The dietary exposure occurring through ingestion of the citrus fruits is much lower, with the median total daily intake of synephrine being up to 6.7 mg/day, and the safety issues are less evident [109,113].

# 5.2.6. Ginger

#### Available Evidence

Ginger (*Zingiber officinale*) is a plant from the Zingiberaceae family, native to South Eastern Asia, which is widely used for food, flavoring and as a medicine in China and India historically [114]. A few small cross-over human trials have studied the effects of ginger on energy expenditure [115,116], with contrasting results, differently from animal [117–122] or in vitro studies [123], which showed improved energy expenditure, lower weight gain, increased browning of WAT and promotion of mitochondrial biogenesis. The contrasting human data, showing either an increased DIT [87] or no thermogenic effects [116], do not allow us to obtain definitive conclusions.

## Molecular Mechanisms of Action

Ginger enhances thermogenesis, increased mitochondrial biogenesis, enhanced BAT function and activated WAT browning in animals through the activation of the sirtuin-1 (SIRT1)/AMPK/PGC-1 $\alpha$  pathways [118,121,122]. The mRNA expression of Sterol regulatory element-binding protein 1 (SREBP-1c) in the liver and leptin in adipose tissues were downregulated, while those of adiponectin, hepatic carnitine palmitoyltransferase1 (CPT-1), acyl-coA oxidase (ACO), Glucose transporter 2 (GLUT-2) and pyruvate kinase (PK) were upregulated after ginger treatment in rats, thus supporting an effect of this compound at the transcriptional level of energy metabolizing proteins [119]. An increase in cellular fatty acid catabolism via the activation of the PPAR $\delta$  pathway has been shown in mice treated with ginger extracts [120].

## Toxicity and Reactivity

Apart from characteristic burning sensation felt upon the consumption of ginger [115], no adverse effects or toxicity has been reported in the human studies. A recent systematic review has also shown its safety in pregnancy [124].

## 5.2.7. Curcuma Longa

#### Available Evidence

Turmeric (*Curcuma longa*) is an herbaceous plant of the ginger family (*Zingiberaceae*) that has been used both as a flavoring and a stimulating agent [125]. Curcumin, also known as diferuloylmethane, is a natural flavonoid component of turmeric, whose antioxidant, anti-inflammatory, antibacterial, anticancer, insulin-sensitizing and hypoglycemic properties have been demonstrated in many studies [126–128]. One animal [129] and one in vitro [130] study showed that curcumin promotes the browning of WAT, while one observational human study [131] has reported that the supplementation of an extract of *Curcuma* reduced the urinary excretion of niacin metabolites and medium- and short-chain acylcarnitines; thus suggesting the potential induction of mitochondrial  $\beta$ -oxidation of fatty acids for energy production. Therefore, the evidence relative to a thermogenic role of curcumin is still scarce.

Nutrients **2020**, 12, 1161 10 of 27

#### Molecular Mechanisms of Action

The following mechanisms have reported for curcumin: increase of thermogenic gene expression, enhanced mitochondrial biogenesis, promotion of the expression of  $\beta$ 3-adrenoreceptors with increased levels of plasma norepinephrine [129], increased levels of hormone-sensitive lipase and p-acyl-CoA carboxylase with enhanced lipolysis, increased expression of UCP1 by AMPK activation [130] and upregulation of the cAMP/protein kinase A (PKA)/cAMP response element-binding protein (CREB) pathway, which plays an important role in energy expenditure and thermogenesis [132].

## Toxicity and Reactivity

Turmeric has been reported to contain many toxic, mutagenic, carcinogenic and hepatotoxic components [133]. Overall, human studies have reported mild adverse effects after curcumin supplementation, with gastrointestinal upsets being most common [134]. The long-term consumption of high doses of curcumin may be dangerous and case reports of acute liver injury have been described [133]. Owing to its inhibitory effect on cytochromes P450, turmeric can potentially interact with many drugs, such as anticoagulants, antibiotics, cardiovascular drugs, anticancer drugs and antidepressants and interactions with clopidogrel, warfarin and etoricoxib have been reported [134,135].

#### 5.2.8. Cinnamon

#### Available Evidence

Cinnamaldehyde is a compound found in cinnamon responsible for its particular flavor, which may improve metabolism owing to its reported hypoglycemic and lipid-lowering effects [136]. Two small randomized human clinical trials in healthy subjects showed that the acute ingestion of extracts of cinnamon (cinnamaldehyde [137] or cinnamyl isobutyrate, respectively [138]) increased energy expenditure (evaluated by indirect calorimetry) by ~3.6 kcal over 90 min from ingestion [137] or reduced short-term energy intake by 4.6% [138], when compared to placebo. These changes are too small to be clinically relevant. Animal studies have demonstrated that extracts of cinnamon elicit thermogenesis responses [139], reduced visceral adiposity, attenuated hyperphagia and normalized energy efficiency [140] and attenuated obesity through the modulation of genes implicated in the lipid metabolism pathways [141]. Currently, chronic studies conducted with the cinnamon amount usually consumed in an everyday diet are lacking.

#### Molecular Mechanisms of Action

In rats, cinnamon-linked increased rate of cold adaptive thermogenesis was due to the elevation in norepinephrine, blood levels of free fatty acid levels and increased expression of UCP1 in BAT [142]. Experimental studies have reported the ability of cinnamaldehyde in activating phospho-AMPK in adipose tissue [140], enhancing thermogenic and metabolic responses in human subcutaneous fat cells through a cAMP dependent protein kinase/p38 mitogen-activated protein kinase (p38 MAPK)-dependent pathway (involved in the transcription of thermogenic genes) [143] and inducing browning in mice subcutaneous adipocytes by increased expressions of UCP1 and other brown adipocyte markers and involvement of the  $\beta$ 3-adrenoreceptor activity [144]. Finally, cinnamaldehyde has been shown to activate the transient receptor potential ankyrin 1 (TRPA1), an ion channel located at the cellular surface, acting as a mechanical and chemical stress sensor, which is involved in adrenalin secretion [145].

#### Toxicity and Reactivity

Cinnamon is obtained from different tree species of the genus *Cinnamonum*: Chinese cinnamon (*Cinnamonum cassia* or *Cinnamonum aromaticum*), coming from the East and containing high level of coumarin, with potential harmful effects [146]; and Ceylon cinnamon (*Cinnamonum zeylanicum* or

Nutrients **2020**, 12, 1161 11 of 27

*Cinnamonum verum*), coming from Sri Lanka and Madagascar, which contains only trace amounts of coumarin. Hepatotoxicity, effects of coumarin on coagulation and potential interference with drugs and mild adverse events have been reported for Chinese cinnamon, while, the consumption of Ceylon cinnamon seems safe [147–149].

## 5.2.9. Chili Pepper (Capsicum Species)

## Available Evidence

Chili peppers are common food flavoring, which are also used as a traditional medicine in some cultures [150,151]. Chilis contain pungent capsaicinoids (capsaicin and dihydro-capsaicin), the major bioactive compounds responsible for the hot taste sensation, non-pungent capsaicin analogs, named capsinoids (e.g., capsiate, dihydro-capsiate and nordihydro-capsiate); and antioxidants, vitamins and carotenoids [150]. Studies in humans investigating a wide range of chili doses have shown that the weight-loss properties of chili are due to enhanced energy expenditure and thermogenesis [152,153]. Conflicting results have been found on the properties of capsaicin and capsiate in decreasing the respiratory quotient by enhancing fat oxidation, due to the different designs of the studies, the body composition and BMI of the subjects included and the habitual consumption of chili in their diet [152–154]. Interestingly, the consumption of 2.56 mg of capsaicin (1.03 g of dried red chili pepper) per meal was able to mitigate the unfavorable negative energy balance effect of decrease in DIT and REE induced by a 25% caloric restriction in humans [155]. However, the doses required to impact metabolism is high and out of the tolerated range for most people [137].

#### Molecular Mechanisms of Action

In mice, dietary capsaicin activated thermogenesis in WAT by up-regulating the expression of SIRT1 and PGC- $1\alpha$ , both of which increase the expression of UCP1 and bone morphogenetic protein-8b, resulting in energy dissipation by thermogenesis, increased EE and metabolic activity [156]. Both in mice and humans, capsaicin and capsinoids enhance energy expenditure by triggering BAT through multiple mechanisms, such as the stimulation of non-shivering thermogenesis by binding to TRPV1, stimulation of the SNS and catecholamine secretion from the adrenal gland [19,137,152–154,156,157].

## Toxicity and Reactivity

In humans, one milligram of capsaicin has neither adverse effects nor affects energy expenditure [137]. Side effects with higher doses of capsaicin, even when provided in capsule form (up to 135 mg/day) include low palatability, gastric distress, dyspepsia, anal burning, bowel irregularities and diarrhea [153]. Capsiate is more tolerable due to its non-pungent characteristics deriving from rapid hydrolysis in the oral cavity, with reduced accessibility to nociceptors [153,154].

# 5.2.10. Garcinia cambogia

# Available Evidence

*Garcinia cambogia* is an herbal product derived from the fruit of the Malabar tamarind tree (also called *Garcinia gummi-gutta*) native to India, Nepal and Sri Lanka [158]. The fruit rind is used either as food preservative, flavoring agent, food-bulking agent or traditional medicine in many Asian countries [159]. *Garcinia* contains xanthones, benzophenones, amino acids and organic acids, of which hydroxy-citric acid (HCA) accounts for 10%–30% of the weight of *Garcinia* fruit and 20%–60% of the extract [158].

Studies with different duration of administration and doses of *Garcinia cambogia* or its extract, were performed both in animals and humans with conflicting results. Favorable effects of *Garcinia cambogia* on glucose and lipid metabolism, as well as on appetite reduction, have been reported [159–161].

Nutrients **2020**, 12, 1161 12 of 27

However, no beneficial effect on EE has been found at different doses and durations of HCA supplementation in human trials, both in the short period and up to 12 weeks [162–164]. A recent meta-analysis of human trials failed to find a significant weight-loss effect of supplementation with *Garcinia cambogia* [165].

#### Molecular Mechanisms of Action

In animal studies, supplementation with HCA induced energy expenditure acceleration by the activation of the adiponectin AMPK signaling pathway [166] or through the regulation of thyroid hormone levels [167]. HCA inhibits serotonin uptake leading to satiety and reduced food intake and down-regulates ATP-citrate lyase, increasing fat oxidation and decreasing *de novo* lipogenesis [159–161].

#### Toxicity and Reactivity

Supplements containing a standardized dose (e.g., 300–500 mg) of *Garcinia cambogia*-derived HCA, consumed up to three times daily, are considered safe [160]. In one human trial, HCA has been supplemented with doses up to 5600 mg/day without adverse effects [168]. However, case reports have described severe hepatotoxicity, including acute liver failure requiring liver transplantation and acute necrotizing eosinophilic myocarditis in subjects using pure *Garcinia cambogia* supplements [169–172]. A natural product for weight loss containing *Garcinia cambogia* and a variety of other ingredients has been associated to fatigue, nausea, vomiting, colic, fever, chills, anorexia, abdominal pain, jaundice, increased levels of liver enzymes and bilirubin in healthy adults [173]. Liver toxicity has been associated with both cholestatic and hepatocellular patterns of injury.

## 5.2.11. Guarana (Paullinia cupana)

### Available Evidence

Guarana is a plant native to the Amazon basin, which is largely used by beverage industries [174]. Guarana seeds contain the highest percentage (2%–8%) of caffeine, compared to any other plant, a high concentration of polyphenols (particularly proanthocyanidins) and small quantities of other stimulant purine alkaloids, such as theobromine and theophylline [174,175]. Two human RCTs reported increased energy expenditure [176] and short-term weight and fat loss [177] after Guarana extract administration. In one animal study, Guarana seed powder supplementation prevented weight gain, insulin resistance and adipokine dysregulation induced by a Western diet [178]. However, conclusive results regarding Guarana supplementation on weight management are still lacking.

#### Molecular Mechanisms of Action

Guarana exerts an anti-adipogenic activity by down-regulating the expression of pro-adipogenic genes, up-regulating the expression of anti-adipogenic genes and increasing  $\beta$ -catenin nuclear translocation, which may contribute to adipogenesis inhibition [179]. Guarana induced BAT expansion, mitochondrial biogenesis, UCP1 overexpression, AMPK activation and minor changes in gut microbiota in rats [178]. Metabolic effects after Guarana supplementation are mainly due to its high caffeine content.

## Toxicity and Reactivity

The European Medication Agency (EMA) recommends a maximum intake of 2250 mg/day of Guarana extract, due to its high percentage of caffeine, which is considered safe up to a dose of 400 mg/day [64–66]. Supplements containing guarana, together with multiple ingredients—above all, high doses of caffeine—have determined agitation, anxiety, insomnia, aggressivity, decreased blood bicarbonate and tachycardia up to cardiorespiratory arrest [66,180]. In rats, pharmacological interactions have been reported after administration of guarana supplements, either with central nervous system stimulants or lamotrigine and amiodarone, leading to exacerbation of seizures and risk of arrythmias [66].

Nutrients **2020**, 12, 1161 13 of 27

#### 5.2.12. Brassicaceae

#### Available Evidence

Broccoli is a vegetable of the *Brassicaceae* (or *Cruciferae*) family, which contain sulfur-based compounds named glucosinolates [181]. These compounds are hydrolyzed to biologically active isothiocyanates (ITC) by the action of myrosinase, a vegetable enzyme present in the human gut microbiota [182,183]. Glucoraphanin, the predominant glucosinolate in broccoli, releases the ITC sulforaphane (SFN) [184]. A 100-g serving of fresh broccoli can release 37–75 mg of SFN, but a therapeutic dose of SFN may not be achieved by a regular diet, as transportation, storage conditions, preparations and cooking, may decrease the vegetable content of SFN [182]. In vitro and animal studies have reported a beneficial effect of SFN on lipid metabolism and thermogenesis [185–188], as well as on gut microbiota [189]. However, extrapolating these results to humans is difficult because studies are still lacking.

## Molecular Mechanisms of Action

In vitro studies have shown that SFN can induce apoptosis in adipocytes [185], inhibit adipocyte differentiation and promote lipolysis in adipocytes [186]. Both glucoraphanin and SFN exert thermogenic effects. SFN increased both mitochondrial biogenesis and function by up-regulating nuclear factor erythroid 2 (NF-E2)-related factor 2 (Nrf2) /SIRT1/ PGC-1 $\alpha$  signaling [187], as well as enhancing UCP1 expression through the activation of the Nrf2; thus promoting browning of WAT [188].

#### Toxicity and Reactivity

Brassicaceae are considered a goitrogenic food due to the ability of the goitrin, a molecule derived from myrosinase hydrolysis of the glucosinolates progoitrin, to inhibit iodine utilization by the thyroid [190]. It is worth noting that the effects of overactivation of the Nrf2-related metabolic pathways are controversial, as the worsening of insulin resistance, as well as glucose and lipid metabolism, have been reported in mice [191].

# 5.2.13. Nuts

Nut-rich diets have been proved to provide positive effects, both on cardiovascular health (owing to their content of mono- and polyunsaturated fats, flavonoids and vitamins) and on body weight, BMI or waist circumference [24,192]. At present, the role of nuts in the regulation of energy balance has not been extensively studied; however, favorable effects have been reported by a couple of RCTs [51,193]. In particular, a 28% increase in DIT 5 h after a meal rich in walnuts and a ~100 kcal higher BEE after 2 months of a nut-rich diet (independent of weight change) have been observed [193]. The role of peanuts is even more uncertain as, in a small RCT, the consumption of high-oleic peanuts increased DIT more than conventional peanuts, but similarly to controls consuming biscuits [194].

# 5.2.14. Apple Cider Vinegar

Apple cider vinegar is a rich source of polyphenols and acetic acid [195]. A systematic review and metanalysis of human trials failed to reach conclusive results on short and long-term blood glucose control after the administration of a wide range of dosages of apple cider vinegar [196]. At present, studies about the effects of apple cider vinegar on human energy expenditure are lacking. In Wistar rats subjected to a high-fat diet, the supplementation of apple cider vinegar (7 mL/kg/day) for 30 days reduced BMI, abdominal circumference and improved satiety [197]. In vitro, acetic acid upregulates the expression of genes for fatty acid oxidation enzymes and thermogenic proteins (e.g., ACO, CPT-1, and UCP2 through  $\alpha$ 2-AMPK/PPAR $\alpha$ -mediated pathway [198]. The lack of human studies on the role of apple cider vinegar on energy balance makes it impossible to draw conclusions about the potential effects of this food.

Nutrients **2020**, 12, 1161 14 of 27

#### 5.2.15. Spirulina

Spirulina refers to a large number of photosynthetic eubacterial species belonging to the phylum Cyanobacteria (*Arthrospira platensis* and A. *maxima*) [158]. These microscopic blue-green algae are a source of high-quality proteins, and contain nearly all essential amino acids, vitamins, minerals, fiber and bioactive compounds [199]. A metanalysis of five human trials (278 subjects) found that the chronic administration of spirulina at variable doses (from one to 4.5 g/day) significantly reduced body weight, body fat percentage and waist circumference [200]. Interestingly, the weight loss was not dose-dependent and was higher in patients with obesity rather than in those with overweight [201]. Many beneficial effects, both on glucose and lipid metabolism and oxidant status, have been described in human studies [200–202]. However, no data about the potential role of spirulina on energy expenditure are available; therefore, at present, no compelling evidence on this topic is available.

#### 5.2.16. Foods without Scientific Evidence to Date

A long list of foods that have been proposed as "fat-burning" or "slimming" agents have not been the subject of scientific studies supporting these supposed benefits. Among these are pineapple, bitter pumpkin (Momordica charantia), mangosteen, Griffonia simplicifolia, Rhodiola rosea, Hoodia gordonii, Fucus vesciculosus, Cissus quadrangularis, Irvingia gabonensis, yohimbine, Caralluma fimbriata, Coleus forskohlii and avocado (Persea americana). These foods do have other potential beneficial properties for human health but, to date, their effectiveness in inducing weight loss is far from proven.

#### 5.3. The Role of Diet Plans

A few human trials have compared the individual energy expenditure under different dietary regimens. A systematic review and meta-analysis of 32 controlled-feeding studies with 563 participants found no effects on TEE of low-carb versus low-fat diets with equivalent protein content [203]. A few RCTs found a significantly lower TEE decrease with low-carb diets when compared either to high-carb [204] or low-fat [205] diets. However, the pooled weighted mean difference in energy expenditure reported in the metanalysis was negligible (26 kcal/day) and favoring low-fat diets [203].

More recently, Ebbeling et al. measured TEE with doubly labeled water in 162 overweight/obese adults randomized to three diets with similar protein and energy content but different carbohydrate percentage (high, 60%; moderate, 40%; and low, 20%) [206]. TEE was increased by 52 kcal/day for every 10% decrease in the proportion of carbohydrates from the diet [206]. Participants following the low-carb diets showed significantly lower circulating leptin and ghrelin levels, affecting both hunger and energy expenditure [207,208]. This study supported the so-called "carbohydrate-insulin model", according to which a reduced proportion of dietary carbohydrate drives lower insulin secretion and increases fat mobilization and oxidation, thus leading to enhanced energy expenditure [209–211]. However, this trial has been criticized due to deviations from the planned analyses, the inclusion of subjects with excessive unaccounted energy and other methodological problems [212].

Therefore, more extensive and methodologically rigorous trials are needed before definitive conclusions on this topic can be reached. At present, the recommendation of combining a proven healthy diet with a daily exercise to obtain/maintain an adequate body muscle mass remains the best method to prevent a decline in energy expenditure after weight loss.

# 6. The Impact of the Gut Microbiota on the Human Energy Balance

The gut microbiota, which is the microbial community populating the digestive tract, regulates both metabolism and energy balance in a symbiotic relationship with the human host. Micro-organisms extract energy from foods that humans cannot digest, producing bioactive compounds such as short-chain fatty acids (SCFAs)—mainly acetate, propionate and butyrate—which supply energy to the intestinal epithelium and liver, providing  $\sim 10\%$  of the daily caloric requirement [213]. The microbiota regulates energy balance through different mechanisms: gut—brain axis control at both the level of

Nutrients **2020**, 12, 1161 15 of 27

intestinal nutrient-sensing mechanisms, as well as at the central nervous system integration sites; development of a low-grade chronic systemic and adipose inflammation together with abnormal gut permeability by an increased relative abundance of pathogenic bacteria; effects on the metabolism of bile acids, with the production of secondary bile acids activating thyroid hormones and oxygen consumption; and impaired secretion of gut peptides and hormones implicated in appetite regulation [214–216]. Alterations in gut–brain signaling can affect the regulation of food intake and SCFAs impact on the incretins and hormones implicated in energy homeostasis, such as glucagon-like-peptide 1, gastric inhibitory peptide, peptide YY, leptin and insulin [217]. The gut microbiota of obese mice has been shown to display a higher capacity to harvest energy from the diet and genes related to phosphotransferase systems involved in microbial carbohydrate processing have been found to be increased in both obese mice and humans [215]. Indeed, the relevance of these processes in the energy balance control has been discussed [218] and an increased pro-inflammatory microbiota, together with the impaired secretion of gut peptides and hormones, seem to be the main mechanisms linking dysbiosis to the occurrence of dysmetabolic diseases [215].

Data relative to the role of microbiota in energy expenditure are controversial. The SCFA turnover has been estimated to account for approximately 7% of REE [219], while gut microbiota composition was not associated with REE level [220].

Supplementation with butyrate enhanced energy expenditure in mice by induction of mitochondrial function in brown fat and skeletal muscle, with increased thermogenesis and fatty acid oxidation [221]. Supplementation with acetate, the most abundant SCFA in the colon (accounting for more than half of the total fecal SCFAs), induced browning by altering the expression of genes involved in beige adipogenesis [222]. An altered nutrient load induced rapid changes in the human gut microbiota composition, these changes being directly associated with stool energy loss in lean individuals, such that a 20% increase in Firmicutes and a corresponding decrease in Bacteroidetes was associated with an increased energy harvest of ≈150 kcal [8]. The bacterial endotoxin lipopolysaccharide—produced by the large gut community of Gram-negative bacteria—binds and activates Toll-like receptor 4, leading both to the repression of adaptive thermogenesis through endoplasmic reticulum stress-mediated mitochondrial dysfunction [223], and the suppression of white adipose tissue browning [224]. Intriguingly, obesity-induced alterations of the gut microbiome persist after successful dieting in obese mice and contribute to weight regain, as persistent dysbiosis contributes to diminishing post-dieting flavonoid levels and reducing energy expenditure [225].

Recently, a stratification of individuals in enterotypes by gut microbiota composition has been proposed, with the most important patterns being the P-type (dominated by Prevotella) and the B-type (dominated by Bacteroides), which probably exist as a continuum, rather than separate entities [226]. The P-type—characterized by hydrolase activity—has been associated with a high-fiber and resistant starch rich diet; while the B-type—characterized by saccharolytic and proteolytic capacity—has been associated with high-fat, low-fiber, Western-type diets [226]. In response to arabinoxylans from grain bran, P-type individuals produced larger amount of SCFAs (especially propionate) and showed higher weight loss and improvements in glucose metabolism when compared to B-type individuals [227–229]. On the other hand, B-type individuals lose more weight on a bifidogenic diet (rich in inulin and oligosaccharides) [226]. Should these data be confirmed in larger samples, they suggest a differential individual response to the same food, according to gut microbiota composition and ability to metabolize food, extracting more or less energy from it. This is in line with emerging concepts of a need for personalized nutrition [230]. Furthermore, complexity in individual microbiota introduces variability and errors in the measurement of energy expenditure, which usually are not considered and controlled for, making prediction of the effects of nutrition on the human energy balance extremely complex [11].

# 7. Conclusions

In Western societies, the availability of highly processed food and general lifestyle have concurred to generate an obesity pandemic. In attempts to address unavoidable weight gain, the general

Nutrients **2020**, 12, 1161 16 of 27

population has been fascinated by foods that can increase energy expenditure. However, only a few foods can potentially affect energy expenditure—usually when consumed in much higher amounts than those usually consumed. In humans, energy balance is complex and multifactorial and physiological compensation occurs with changes in energy intake and/or expenditure. Moreover, other factors such as microbiota composition and activity are involved, influencing food metabolism and nutrient utilization. Any attempts to classify diets and foods based on supposed roles in energy balance implies an excessive simplification of real biologic complexity, which we are just beginning to understand. Long-term and well-designed human intervention trials in different population groups are important to draw any conclusions on the effect of foods and dietary regimens in energy balance.

**Supplementary Materials:** The following are available online at <a href="http://www.mdpi.com/2072-6643/12/4/1161/s1">http://www.mdpi.com/2072-6643/12/4/1161/s1</a>, Table S1: Effects on energy expenditure and hypothesized mechanisms to induce weight loss.

**Author Contributions:** Conceptualization, S.B.; methodology, S.B and N.P; writing—original draft preparation, S.B., M.F., D.F., M.P., E.G. and N.P.; writing—review and editing, S.B., M.P. and N.P.; table and figure creation S.B. and M.P.; supervision, S.B., E.G. and N.P. All authors have read and agree to the published version of the manuscript.

**Funding:** This research did not receive any specific grant from funding agencies in the public, commercial or not-for-profit sectors.

Conflicts of Interest: All authors declare no conflict of interest.

#### References

- 1. Shader, R.I. Troublesome news, fake news, biased or incomplete news. *Clin. Ther.* **2018**, *40*, 1429–1434. [CrossRef]
- 2. Casazza, K.; Fontaine, K.R.; Astrup, A.; Birch, L.L.; Brown, A.W.; Bohan Brown, M.M.; Durant, N.; Dutton, G.; Foster, E.M.; Heymsfield, S.B.; et al. Myths, presumptions, and facts about obesity. *N. Engl. J. Med.* **2013**, *368*, 446–454. [CrossRef]
- 3. Hall, K.D.; Heymsfield, S.B.; Kemnitz, J.W.; Klein, S.; Schoeller, D.A.; Speakman, J.R. Energy balance and its components: Implications for body weight regulation. *Am. J. Clin. Nutr.* **2012**, *95*, 989–994. [CrossRef] [PubMed]
- 4. L'alimentazione. Available online: https://www.issalute.it/index.php/falsi-miti-e-bufale/l-alimentazione? limitstart=0 (accessed on 12 October 2019).
- Garvey, W.T.; Mechanick, J.I.; Brett, E.M.; Garber, A.J.; Hurley, D.L.; Jastreboff, A.M.; Nadolsky, K.; Pessah-Pollack, R.; Plodkowski, R. Reviewers of the AACE/ACE obesity clinical practice guidelines. American association of clinical endocrinologists and American College of Endocrinology comprehensive clinical practice guidelines for medical care of patients with obesity. *Endocr. Pract.* 2016, 22, 842–884. [CrossRef] [PubMed]
- 6. de Keersmaecker, J.; Roets, A. 'Fake news': Incorrect, but hard to correct: The role of cognitive ability on the impact of false information on social impressions. *Intelligence* **2017**, *65*, 107–110. [CrossRef]
- 7. Hopkins, M.; Blundell, J.E. Energy balance, body composition, sedentariness and appetite regulation: Pathways to obesity. *Clin. Sci.* **2016**, *130*, 1615–1628. [CrossRef]
- 8. Jumpertz, R.; Le, D.S.; Turnbaugh, P.J.; Trinidad, C.; Bogardus, C.; Gordon, J.I.; Krakoff, J. Energy-balance studies reveal associations between gut microbes, caloric load, and nutrient absorption in humans. *Am. J. Clin. Nutr.* **2011**, *94*, 58–65. [CrossRef]
- 9. Carneiro, I.P.; Elliott, S.A.; Siervo, M.; Padwal, R.; Bertoli, S.; Battezzati, A.; Prado, C.M. Is obesity associated with altered energy expenditure? *Adv. Nutr.* **2016**, *7*, 476–487. [CrossRef]
- 10. Yoo, S. Dynamic energy balance and obesity prevention. J. Obes. Metab. Syndr. 2018, 27, 203–212. [CrossRef]
- 11. Armstrong, L.E.; Casa, D.J.; Belval, L.N. Metabolism, bioenergetics and thermal physiology: Influences of the human intestinal microbiota. *Nutr. Res. Rev.* **2019**, *32*, 1–13. [CrossRef]
- 12. Westerterp, K.R. Exercise, energy balance and body composition. *Eur. J. Clin. Nutr.* **2018**, 72, 1246–1250. [CrossRef] [PubMed]
- 13. Westerterp, K.R. Control of energy expenditure in humans. *Eur. J. Clin. Nutr.* **2017**, *71*, 340–344. [CrossRef] [PubMed]

Nutrients **2020**, 12, 1161 17 of 27

14. Quatela, A.; Callister, R.; Patterson, A.; MacDonald-Wicks, L. The energy content and composition of meals consumed after an overnight fast and their effects on diet induced thermogenesis: A systematic review, meta-analyses and meta-regressions. *Nutrients* **2016**, *8*, 670. [CrossRef] [PubMed]

- 15. Lowell, B.B.; Spiegelman, B.M. Towards a molecular understanding of adaptive thermogenesis. *Nature* **2000**, 404, 652–660. [CrossRef]
- 16. Müller, M.J.; Bosy-Westphal, A. Adaptive thermogenesis with weight loss in humans. *Obesity* **2013**, 21, 218–228. [CrossRef]
- 17. Tansey, E.A.; Johnson, C.D. Recent advances in thermoregulation. *Adv. Physiol. Educ.* **2015**, *39*, 139–148. [CrossRef]
- 18. Rosenbaum, M.; Leibel, R.L. Adaptive thermogenesis in humans. *Int. J. Obes.* 2005 **2010**, 34, S47–S55. [CrossRef]
- 19. Saito, M.; Yoneshiro, T.; Matsushita, M. Activation and recruitment of brown adipose tissue by cold exposure and food ingredients in humans. *Best Pract. Res. Clin. Endocrinol. Metab.* **2016**, *30*, 537–547. [CrossRef]
- 20. Mele, L.; Bidault, G.; Mena, P.; Crozier, A.; Brighenti, F.; Vidal-Puig, A.; Del Rio, D. Dietary (Poly)phenols, brown adipose tissue activation, and energy expenditure: A narrative review. *Adv. Nutr.* **2017**, *8*, 694–704. [CrossRef]
- 21. Nedergaard, J.; Cannon, B. The browning of white adipose tissue: Some burning issues. *Cell Metab.* **2014**, 20, 396–407. [CrossRef]
- 22. Müller, M.J.; Enderle, J.; Bosy-Westphal, A. Changes in energy expenditure with weight gain and weight loss in humans. *Curr. Obes. Rep.* **2016**, *5*, 413–423. [CrossRef] [PubMed]
- 23. Capuano, E.; Oliviero, T.; Fogliano, V.; Pellegrini, N. Role of the food matrix and digestion on calculation of the actual energy content of food. *Nutr. Rev.* **2018**, *76*, 274–289. [CrossRef] [PubMed]
- 24. Tindall, A.M.; Petersen, K.S.; Lamendella, R.; Shearer, G.C.; Murray-Kolb, L.E.; Proctor, D.N.; Kris-Etherton, P.M. Tree nut consumption and adipose tissue mass: Mechanisms of action. *Curr. Dev. Nutr.* **2018**, 2, nzy069. [CrossRef] [PubMed]
- 25. Ros, E. Nuts and CVD. Br. J. Nutr. 2015, 11, S111–S120. [CrossRef]
- 26. Novotny, J.A.; Gebauer, S.K.; Baer, D.J. Discrepancy between the Atwater factor predicted and empirically measured energy values of almonds in human diets. *Am. J. Clin. Nutr.* **2012**, *96*, 296–301. [CrossRef]
- 27. Baer, D.J.; Gebauer, S.K.; Novotny, J.A. Walnuts consumed by healthy adults provide less available energy than predicted by the Atwater factors. *J. Nutr.* **2016**, *146*, 9–13. [CrossRef]
- 28. Baer, D.J.; Gebauer, S.K.; Novotny, J.A. Measured energy value of pistachios in the human diet. *Br. J. Nutr.* **2012**, *107*, 120–125. [CrossRef]
- 29. Baer, D.J.; Novotny, J.A. Metabolizable energy from cashew nuts is less than that predicted by atwater factors. *Nutrients* **2018**, *11*, 33. [CrossRef]
- 30. Capuano, E.; Pellegrini, N.; Ntone, E.; Nikiforidis, C.V. In vitro lipid digestion in raw and roasted hazelnut particles and oil bodies. *Food Funct.* **2018**, *9*, 2508–2516. [CrossRef]
- 31. Ellis, P.R.; Kendall, C.W.C.; Ren, Y.; Parker, C.; Pacy, J.F.; Waldron, K.W.; Jenkins, D.J.A. Role of Cell Walls in the bioaccessibility of lipids in almond seeds. *Am. J. Clin. Nutr.* **2004**, *80*, 604–613. [CrossRef]
- 32. Cassady, B.A.; Hollis, J.H.; Fulford, A.D.; Considine, R.V.; Mattes, R.D. Mastication of almonds: Effects of lipid bioaccessibility, appetite, and hormone response. *Am. J. Clin. Nutr.* **2009**, *89*, 794–800. [CrossRef] [PubMed]
- 33. Mandalari, G.; Parker, M.L.; Grundy, M.M.L.; Grassby, T.; Smeriglio, A.; Bisignano, C.; Raciti, R.; Trombetta, D.; Baer, D.J.; Wilde, P.J. Understanding the effect of particle size and processing on almond lipid bioaccessibility through microstructural analysis: From mastication to faecal collection. *Nutrients* **2018**, *10*, 213. [CrossRef] [PubMed]
- 34. Gebauer, S.K.; Novotny, J.A.; Bornhorst, G.M.; Baer, D.J. Food processing and structure impact the metabolizable energy of almonds. *Food Funct.* **2016**, *7*, 4231–4238. [CrossRef] [PubMed]
- 35. Traoret, C.J.; Lokko, P.; Cruz, A.C.R.F.; Oliveira, C.G.; Costa, N.M.B.; Bressan, J.; Alfenas, R.C.G.; Mattes, R.D. Peanut digestion and energy balance. *Int. J. Obes.* **2008**, *32*, 322–328. [CrossRef]
- 36. Levine, J.A. Measurement of energy expenditure. Public Health Nutr. 2005, 8, 1123–1132. [CrossRef]
- 37. Levine, J.A.; Schleusner, S.J.; Jensen, M.D. Energy expenditure of nonexercise activity. *Am. J. Clin. Nutr.* **2000**, 72, 1451–1454. [CrossRef]
- 38. Ho, K.K.Y. Diet-induced thermogenesis: Fake friend or foe? J. Endocrinol. 2018, 238, R185–R191. [CrossRef]

Nutrients **2020**, 12, 1161 18 of 27

39. Calcagno, M.; Kahleova, H.; Alwarith, J.; Burgess, N.N.; Flores, R.A.; Busta, M.L.; Barnard, N.D. The thermic effect of food: A Review. *J. Am. Coll. Nutr.* **2019**, *38*, 547–551. [CrossRef]

- 40. Camastra, S.; Bonora, E.; Del Prato, S.; Rett, K.; Weck, M.; Ferrannini, E. Effect of obesity and insulin resistance on resting and glucose-induced thermogenesis in man. EGIR (European Group for the Study of Insulin Resistance). *Int. J. Obes.* **1999**, *23*, 1307–1313. [CrossRef]
- 41. Bo, S.; Fadda, M.; Castiglione, A.; Ciccone, G.; De Francesco, A.; Fedele, D.; Guggino, A.; Parasiliti Caprino, M.; Ferrara, S.; Vezio Boggio, M.; et al. Is the timing of caloric intake associated with variation in diet-induced thermogenesis and in the metabolic pattern? A randomized cross-over study. *Int. J. Obes.* **2015**, *39*, 1689–1695. [CrossRef]
- 42. Bo, S.; Broglio, F.; Settanni, F.; Parasiliti Caprino, M.; Ianniello, A.; Mengozzi, G.; De Francesco, A.; Fadda, M.; Fedele, D.; Guggino, A.; et al. Effects of meal timing on changes in circulating epinephrine, norepinephrine, and acylated ghrelin concentrations: A pilot study. *Nutr. Diabetes* **2017**, *7*, 303. [CrossRef] [PubMed]
- 43. Martin, A.; Normand, S.; Sothier, M.; Peyrat, J.; Louche-Pelissier, C.; Laville, M. Is advice for breakfast consumption justified? Results from a short-term dietary and metabolic experiment in young healthy men. *Br. J. Nutr.* **2000**, *84*, 337–344. [CrossRef]
- 44. Soenen, S.; Martens, E.A.P.; Hochstenbach-Waelen, A.; Lemmens, S.G.T.; Westerterp-Plantenga, M.S. Normal protein intake is required for body weight loss and weight maintenance, and elevated protein intake for additional preservation of resting energy expenditure and fat free mass. *J. Nutr.* **2013**, *143*, 591–596. [CrossRef] [PubMed]
- 45. Nagai, N.; Sakane, N.; Moritani, T. Metabolic responses to high-fat or low-fat meals and association with sympathetic nervous system activity in healthy young men. *J. Nutr. Sci. Vitaminol.* **2005**, *51*, 355–360. [CrossRef] [PubMed]
- 46. Raben, A.; Agerholm-Larsen, L.; Flint, A.; Holst, J.J.; Astrup, A. Meals with similar energy densities but rich in protein, fat, carbohydrate, or alcohol have different effects on energy expenditure and substrate metabolism but not on appetite and energy intake. Am. J. Clin. Nutr. 2003, 77, 91–100. [CrossRef] [PubMed]
- 47. Ravn, A.M.; Gregersen, N.T.; Christensen, R.; Rasmussen, L.G.; Hels, O.; Belza, A.; Raben, A.; Larsen, T.M.; Toubro, S.; Astrup, A. Thermic effect of a meal and appetite in adults: An individual participant data meta-analysis of meal-test trials. *Food Nutr. Res.* **2013**, *57*, 19676. [CrossRef] [PubMed]
- 48. Kassis, A.; Godin, J.P.; Moille, S.E.; Nielsen-Moennoz, C.; Groulx, K.; Oguey-Araymon, S.; Praplan, F.; Beaumont, M.; Sauser, J.; Monnard, I.; et al. Effects of protein quantity and type on diet induced thermogenesis in overweight adults: A randomized controlled trial. *Clin. Nutr.* **2019**, *38*, 1570–1580. [CrossRef]
- 49. Clegg, M.E.; Golsorkhi, M.; Henry, C.J. Combined medium-chain triglyceride and chilli feeding increases diet-induced thermogenesis in normal-weight humans. *Eur. J. Nutr.* **2013**, *52*, 1579–1585. [CrossRef]
- 50. Kasai, M.; Nosaka, N.; Maki, H.; Suzuki, Y.; Takeuchi, H.; Aoyama, T.; Ohra, A.; Harada, Y.; Okazaki, M.; Kondo, K. Comparison of diet-induced thermogenesis of foods containing medium-versus long-chain triacylglycerols. *J. Nutr. Sci. Vitaminol.* **2002**, *48*, 536–540. [CrossRef]
- 51. Casas-Agustench, P.; López-Uriarte, P.; Bulló, M.; Ros, E.; Gómez-Flores, A.; Salas-Salvadó, J. Acute effects of three high-fat meals with different fat saturations on energy expenditure, substrate oxidation and satiety. *Clin. Nutr.* **2009**, *28*, 39–45. [CrossRef]
- 52. Zamora Navarro, S.; Pérez-Llamas, F. Errors and myths in feeding and nutrition: Impact on the problems of obesity. *Nutr. Hosp.* **2013**, *28*, 81–88. [CrossRef] [PubMed]
- 53. Couto, C.C.; Santos, T.F.; Mamede, A.M.G.N.; Oliveira, T.C.; Souza, A.M.; Freitas-Silva, O.; Oliveira, E.M.M. Coffea arabica and c. canephora discrimination in roasted and ground coffee from reference material candidates by real-time PCR. *Food Res. Int.* **2019**, *115*, 227–233. [CrossRef] [PubMed]
- 54. Nuhu, A.A. Bioactive micronutrients in coffee: Recent analytical approaches for characterization and quantification. *ISRN Nutr.* **2014**, 2014, 384230. [CrossRef] [PubMed]
- 55. Velickovic, K.; Wayne, D.; Leija, H.A.L.; Bloor, I.; Morris, D.E.; Law, J.; Budge, H.; Sacks, H.; Symonds, M.E.; Sottile, V. Caffeine exposure induces browning features in adipose tissue in vitro and in vivo. *Sci. Rep.* **2019**, 9,9104. [CrossRef] [PubMed]
- 56. Dulloo, A.G.; Geissler, C.A.; Horton, T.; Collins, A.; Miller, D.S. Normal caffeine consumption: Influence on thermogenesis and daily energy expenditure in lean and postobese human volunteers. *Am. J. Clin. Nutr.* **1989**, *49*, 44–50. [CrossRef] [PubMed]

Nutrients **2020**, 12, 1161 19 of 27

57. Koot, P.; Deurenberg, P. Comparison of changes in energy expenditure and body temperatures after caffeine consumption. *Ann. Nutr. Metab.* **1995**, *39*, 135–142. [CrossRef] [PubMed]

- 58. Belza, A.; Toubro, S.; Astrup, A. The effect of caffeine, green tea and tyrosine on thermogenesis and energy intake. *Eur. J. Clin. Nutr.* **2009**, *63*, 57–64. [CrossRef]
- 59. Gorji, Z.; Varkaneh, H.K.; Talaei, S.; Nazary-Vannani, A.; Clark, C.C.T.; Fatahi, S.; Rahmani, J.; Salamat, S.; Zhang, Y. The effect of green-coffee extract supplementation on obesity: A systematic review and dose-response meta-analysis of randomized controlled trials. *Phytomedicine Int. J. Phytother. Phytopharm.* **2019**, *63*, 153018. [CrossRef]
- 60. Ghadieh, H.E.; Smiley, Z.N.; Kopfman, M.W.; Najjar, M.G.; Hake, M.J.; Najjar, S.M. Chlorogenic acid/chromium supplement rescues diet-induced insulin resistance and obesity in mice. *Nutr. Metab.* **2015**, *12*, 19. [CrossRef]
- 61. Acar-Tek, N.; Ağagündüz, D.; Ayhan, B. Effect of green coffee consumption on resting energy expenditure, blood pressure, and body temperature in healthy women: A pilot study. *J. Am. Coll. Nutr.* **2018**, 37, 691–700. [CrossRef]
- 62. Dulloo, A.G. The search for compounds that stimulate thermogenesis in obesity management: From pharmaceuticals to functional food ingredients. *Obes. Rev.* **2011**, *12*, 866–883. [CrossRef] [PubMed]
- 63. Stohs, S.J.; Badmaev, V. A review of natural stimulant and non-stimulant thermogenic agents. *Phytother. Res.* **2016**, *30*, 732–740. [CrossRef] [PubMed]
- 64. EFSA Panel on Dietetic Products; Nutrition and Allergies (NDA). Scientific opinion on the safety of caffeine. *EFSA J.* **2015**, *13*, 4102. [CrossRef]
- 65. Wikoff, D.; Welsh, B.T.; Henderson, R.; Brorby, G.P.; Britt, J.; Myers, E.; Goldberger, J.; Lieberman, H.R.; O'Brien, C.; Peck, J.; et al. Systematic review of the potential adverse effects of caffeine consumption in healthy adults, pregnant women, adolescents, and children. *Food Chem. Toxicol.* 2017, 109, 585–648. [CrossRef] [PubMed]
- 66. Patrick, M.; Kim, H.A.; Oketch-Rabah, H.; Marles, R.J.; Roe, A.L.; Calderón, A.I. Safety of guarana seed as a dietary ingredient: A review. *J. Agric. Food Chem.* **2019**, *67*, 11281–11287. [CrossRef]
- 67. Hayat, K.; Iqbal, H.; Malik, U.; Bilal, U.; Mushtaq, S. Tea and its consumption: Benefits and risks. *Crit. Rev. Food Sci. Nutr.* **2015**, *55*, 939–954. [CrossRef]
- 68. Türközü, D.; Tek, N.A. A minireview of effects of green tea on energy expenditure. *Crit. Rev. Food Sci. Nutr.* **2017**, *57*, 254–258. [CrossRef]
- 69. Yang, C.S.; Wang, H.; Sheridan, Z.P. Studies on prevention of obesity, metabolic syndrome, diabetes, cardiovascular diseases and cancer by tea. *J. Food Drug Anal.* **2018**, *26*, 1–13. [CrossRef]
- 70. Seo, D.B.; Jeong, H.W.; Kim, Y.J.; Kim, S.; Kim, J.; Lee, J.H.; Joo, K.; Choi, J.K.; Shin, S.S.; Lee, S.J. Fermented green tea extract exhibits hypolipidaemic effects through the inhibition of pancreatic lipase and promotion of energy expenditure. *Br. J. Nutr.* **2017**, *117*, 177–186. [CrossRef]
- 71. Dinh, T.C.; Thi Phuong, T.N.; Minh, L.B.; Minh Thuc, V.T.; Bac, N.D.; Van Tien, N.; Pham, V.H.; Show, P.L.; Tao, Y.; Nhu Ngoc, V.T.; et al. The effects of green tea on lipid metabolism and its potential applications for obesity and related metabolic disorders—An existing update. *Diabetes Metab. Syndr.* **2019**, *13*, 1667–1673. [CrossRef]
- 72. Huang, J.; Wang, Y.; Xie, Z.; Zhou, Y.; Zhang, Y.; Wan, X. The anti-obesity effects of green tea in human intervention and basic molecular studies. *Eur. J. Clin. Nutr.* **2014**, *68*, 1075–1087. [CrossRef] [PubMed]
- 73. Dulloo, A.G.; Duret, C.; Rohrer, D.; Girardier, L.; Mensi, N.; Fathi, M.; Chantre, P.; Vandermander, J. Efficacy of a green tea extract rich in catechin polyphenols and caffeine in increasing 24-h energy expenditure and fat oxidation in humans. *Am. J. Clin. Nutr.* **1999**, *70*, 1040–1045. [CrossRef] [PubMed]
- 74. Yoneshiro, T.; Matsushita, M.; Hibi, M.; Tone, H.; Takeshita, M.; Yasunaga, K.; Katsuragi, Y.; Kameya, T.; Sugie, H.; Saito, M. Tea catechin and caffeine activate brown adipose tissue and increase cold-induced thermogenic capacity in humans. *Am. J. Clin. Nutr.* **2017**, *105*, 873–881. [CrossRef] [PubMed]
- 75. Janssens, P.L.H.R.; Hursel, R.; Westerterp-Plantenga, M.S. Long-term green tea extract supplementation does not affect fat absorption, resting energy expenditure, and body composition in adults. *J. Nutr.* **2015**, *145*, 864–870. [CrossRef] [PubMed]
- 76. Jurgens, T.M.; Whelan, A.M.; Killian, L.; Doucette, S.; Kirk, S.; Foy, E. Green tea for weight loss and weight maintenance in overweight or obese adults. *Cochrane Database Syst. Rev.* **2012**, *12*, CD008650. [CrossRef]
- 77. Hu, J.; Webster, D.; Cao, J.; Shao, A. The safety of green tea and green tea extract consumption in adults—Results of a systematic review. *Regul. Toxicol. Pharmacol.* **2018**, 95, 412–433. [CrossRef]

Nutrients **2020**, 12, 1161 20 of 27

78. Cooper, K.A.; Campos-Giménez, E.; Jiménez Alvarez, D.; Rytz, A.; Nagy, K.; Williamson, G. Predictive relationship between polyphenol and nonfat cocoa solids content of chocolate. *J. Agric. Food Chem.* **2008**, *56*, 260–265. [CrossRef]

- 79. Baggott, M.J.; Childs, E.; Hart, A.B.; de Bruin, E.; Palmer, A.A.; Wilkinson, J.E.; de Wit, H. Psychopharmacology of theobromine in healthy volunteers. *Psychopharmacology* **2013**, 228, 109–118. [CrossRef]
- 80. Massaro, M.; Scoditti, E.; Carluccio, M.A.; Kaltsatou, A.; Cicchella, A. Effect of Cocoa products and its polyphenolic constituents on exercise performance and exercise-induced muscle damage and inflammation: A review of clinical trials. *Nutrients* **2019**, *11*, 1471. [CrossRef]
- 81. Hatano, T.; Miyatake, H.; Natsume, M.; Osakabe, N.; Takizawa, T.; Ito, H.; Yoshida, T. Proanthocyanidin glycosides and related polyphenols from cacao liquor and their antioxidant effects. *Phytochemistry* **2002**, *59*, 749–758. [CrossRef]
- 82. Actis-Goretta, L.; Lévèques, A.; Giuffrida, F.; Romanov-Michailidis, F.; Viton, F.; Barron, D.; Duenas-Paton, M.; Gonzalez-Manzano, S.; Santos-Buelga, C.; Williamson, G.; et al. Elucidation of (-)-epicatechin metabolites after ingestion of chocolate by healthy humans. *Free Radic. Biol. Med.* **2012**, *53*, 787–795. [CrossRef] [PubMed]
- 83. Matsui, N.; Ito, R.; Nishimura, E.; Yoshikawa, M.; Kato, M.; Kamei, M.; Shibata, H.; Matsumoto, I.; Abe, K.; Hashizume, S. Ingested Cocoa can prevent high-fat diet-induced obesity by regulating the expression of genes for fatty acid metabolism. *Nutrition* **2005**, *21*, 594–601. [CrossRef] [PubMed]
- 84. Kord-Varkaneh, H.; Ghaedi, E.; Nazary-Vanani, A.; Mohammadi, H.; Shab-Bidar, S. Does Cocoa/dark chocolate supplementation have favorable effect on body weight, body mass index and waist circumference? A systematic review, meta-analysis and dose-response of randomized clinical trials. *Crit. Rev. Food Sci. Nutr.* **2019**, *59*, 2349–2362. [CrossRef] [PubMed]
- 85. Strat, K.M.; Rowley, T.J.; Smithson, A.T.; Tessem, J.S.; Hulver, M.W.; Liu, D.; Davy, B.M.; Davy, K.P.; Neilson, A.P. Mechanisms by which Cocoa flavanols improve metabolic syndrome and related disorders. *J. Nutr. Biochem.* **2016**, *35*, 1–21. [CrossRef] [PubMed]
- 86. Bowser, S.M.; Moore, W.T.; McMillan, R.P.; Dorenkott, M.R.; Goodrich, K.M.; Ye, L.; O'Keefe, S.F.; Hulver, M.W.; Neilson, A.P. High-molecular-weight Cocoa procyanidins possess enhanced insulin-enhancing and insulin mimetic activities in human primary skeletal muscle cells compared to smaller procyanidins. *J. Nutr. Biochem.* 2017, 39, 48–58. [CrossRef] [PubMed]
- 87. Yamashita, Y.; Okabe, M.; Natsume, M.; Ashida, H. Cacao liquor procyanidins prevent postprandial hyperglycaemia by increasing glucagon-like peptide-1 activity and AMP-activated protein kinase in mice. *J. Nutr. Sci.* **2019**, *8*, e2. [CrossRef]
- 88. Yamashita, Y.; Okabe, M.; Natsume, M.; Ashida, H. Prevention mechanisms of glucose intolerance and obesity by cacao liquor procyanidin extract in high-fat diet-fed C57BL/6 mice. *Arch. Biochem. Biophys.* **2012**, 527, 95–104. [CrossRef]
- 89. Viollet, B.; Foretz, M.; Guigas, B.; Horman, S.; Dentin, R.; Bertrand, L.; Hue, L.; Andreelli, F. Activation of AMP-activated protein kinase in the liver: A new strategy for the management of metabolic hepatic disorders. *J. Physiol.* **2006**, *574*, 41–53. [CrossRef]
- 90. Franco, R.; Oñatibia-Astibia, A.; Martínez-Pinilla, E. Health benefits of methylxanthines in cacao and chocolate. *Nutrients* **2013**, *5*, 4159–4173. [CrossRef]
- 91. Fredholm, B.B.; Lindgren, E. The effect of alkylxanthines and other phosphodiesterase inhibitors on adenosine-receptor mediated decrease in lipolysis and cyclic AMP accumulation in rat fat cells. *Acta Pharmacol. Toxicol.* **1984**, *54*, 64–71. [CrossRef]
- 92. Harpaz, E.; Tamir, S.; Weinstein, A.; Weinstein, Y. The effect of caffeine on energy balance. *J. Basic Clin. Physiol. Pharmacol.* **2017**, *28*, 1–10. [CrossRef] [PubMed]
- 93. Abt, E.; Sam, J.F.; Gray, P.; Robin, L.P. Cadmium and lead in cocoa powder and chocolate products in the US market. *Food Addit. Contam. Part B Surveill.* **2018**, *11*, 92–102. [CrossRef] [PubMed]
- 94. Lo Dico, G.M.; Galvano, F.; Dugo, G.; D'ascenzi, C.; Macaluso, A.; Vella, A.; Giangrosso, G.; Cammilleri, G.; Ferrantelli, V. Toxic metal levels in cocoa powder and chocolate by ICP-MS method after microwave-assisted digestion. *Food Chem.* **2018**, 245, 1163–1168. [CrossRef] [PubMed]
- 95. Bracesco, N.; Sanchez, A.G.; Contreras, V.; Menini, T.; Gugliucci, A. Recent advances on ilex paraguariensis research: Minireview. *J. Ethnopharmacol.* **2011**, *136*, 378–384. [CrossRef]

Nutrients **2020**, 12, 1161 21 of 27

96. Cho, A.S.; Jeon, S.M.; Kim, M.J.; Yeo, J.; Seo, K.I.; Choi, M.S.; Lee, M.K. Chlorogenic acid exhibits anti-obesity property and improves lipid metabolism in high-fat diet-induced-obese mice. *Food Chem. Toxicol.* **2010**, *48*, 937–943. [CrossRef]

- 97. Martinet, A.; Hostettmann, K.; Schutz, Y. Thermogenic effects of commercially available plant preparations aimed at treating human obesity. *Phytomedicine* **1999**, *6*, 231–238. [CrossRef]
- 98. Alkhatib, A. Yerba maté (Illex Paraguariensis) ingestion augments fat oxidation and energy expenditure during exercise at various submaximal intensities. *Nutr. Metab.* **2014**, *11*, 42. [CrossRef]
- 99. Choi, M.S.; Park, H.J.; Kim, S.R.; Kim, D.Y.; Jung, U.J. Long-term dietary supplementation with yerba mate ameliorates diet-induced obesity and metabolic disorders in mice by regulating energy expenditure and lipid metabolism. *J. Med. Food* **2017**, *20*, 1168–1175. [CrossRef]
- 100. Arçari, D.P.; Santos, J.C.; Gambero, A.; Ribeiro, M.L. The in vitro and in vivo effects of yerba mate (Ilex paraguariensis) extract on adipogenesis. *Food Chem.* **2013**, *141*, 809–815. [CrossRef]
- 101. Wood Dos Santos, T.; Cristina Pereira, Q.; Teixeira, L.; Gambero, A.; Villena, A.J.; Lima Ribeiro, M. Effects of polyphenols on thermogenesis and mitochondrial biogenesis. *Int. J. Mol. Sci.* **2018**, *19*, 2757. [CrossRef]
- 102. de Andrade, F.; de Albuquerque, C.A.C.; Maraschin, M.; da Silva, E.L. Safety assessment of yerba mate (Ilex paraguariensis) dried extract: Results of acute and 90 days subchronic toxicity studies in rats and rabbits. *Food Chem. Toxicol.* **2012**, *50*, 328–334. [CrossRef] [PubMed]
- 103. de Sousa, W.R.; Lourenço, B.H.L.B.; Reis, M.d.P.; Donadel, G.; Marques, M.A.A.; Cardozo Junior, E.L.; Jacomassi, E.; Belettini, S.T.; Lívero, F.A.D.R.; Gasparotto Junior, A.; et al. Evaluation of reproductive toxicology of aqueous extract of yerba mate (Ilex paraguariensis a. st.-hil.), a traditional south american beverage. *J. Med. Food* 2019, 22, 97–101. [CrossRef] [PubMed]
- 104. Preuss, H.G.; Di Ferdinando, D.; Bagchi, M.; Bagchi, D. Citrus aurantium as a thermogenic, weight-reduction replacement for ephedra: An overview. *J. Med.* **2002**, *33*, 247–264. [PubMed]
- 105. Stohs, S.J. Safety, Efficacy, and mechanistic studies regarding Citrus aurantium (Bitter Orange) extract and p-Synephrine. *Phytother. Res. PTR* **2017**, *31*, 1463–1474. [CrossRef]
- 106. Ríos-Hoyo, A.; Gutiérrez-Salmeán, G. New dietary supplements for obesity: What we currently know. *Curr. Obes. Rep.* **2016**, *5*, 262–270. [CrossRef]
- 107. Stohs, S.J.; Preuss, H.G.; Keith, S.C.; Keith, P.L.; Miller, H.; Kaats, G.R. Effects of p-Synephrine alone and in combination with selected bioflavonoids on resting metabolism, blood pressure, heart rate and self-reported mood changes. *Int. J. Med. Sci.* 2011, *8*, 295–301. [CrossRef]
- 108. Gougeon, R.; Harrigan, K.; Tremblay, J.F.; Hedrei, P.; Lamarche, M.; Morais, J.A. Increase in the Thermic effect of food in women by adrenergic amines extracted from Citrus aurantium. *Obes. Res.* **2005**, *13*, 1187–1194. [CrossRef]
- 109. Stohs, S.J.; Preuss, H.G.; Shara, M. A Review of the human clinical studies involving Citrus aurantium (Bitter Orange) extract and its primary protoalkaloid p-Synephrine. *Int. J. Med. Sci.* **2012**, *9*, 527–538. [CrossRef]
- 110. Kliszczewicz, B.; Bechke, E.; Williamson, C.; Green, Z.; Bailey, P.; McLester, J.; McLester, C. Citrus Aurantium and caffeine complex versus placebo on biomarkers of metabolism: A double blind crossover design. *J. Int. Soc. Sports Nutr.* **2019**, *16*, 4. [CrossRef]
- 111. Guo, J.; He, Z.; Wu, S.; Zeng, M.; Chen, J. Binding of aromatic compounds with soy protein isolate in an aqueous model: Effect of pH. *J. Food Biochem.* **2019**, 43, e12817. [CrossRef]
- 112. Park, J.; Kim, H.L.; Jung, Y.; Ahn, K.S.; Kwak, H.J.; Um, J.Y. Bitter orange (Citrus aurantium Linné) improves obesity by regulating adipogenesis and thermogenesis through AMPK activation. *Nutrients* 2019, 11, 1988. [CrossRef] [PubMed]
- 113. Bakhiya, N.; Ziegenhagen, R.; Hirsch-Ernst, K.I.; Dusemund, B.; Richter, K.; Schultrich, K.; Pevny, S.; Schäfer, B.; Lampen, A. Phytochemical compounds in sport nutrition: Synephrine and hydroxycitric acid (HCA) as examples for evaluation of possible health risks. *Mol. Nutr. Food Res.* 2017, *61*, 1601020. [CrossRef] [PubMed]
- 114. Ginger|plant. Encycl. Br.. 2019. Available online: https://www.britannica.com/plant/ginger (accessed on 30 October 2019).
- 115. Mansour, M.S.; Ni, Y.M.; Roberts, A.L.; Kelleman, M.; Roychoudhury, A.; St-Onge, M.P. Ginger consumption enhances the thermic effect of food and promotes feelings of satiety without affecting metabolic and hormonal parameters in overweight men: A pilot study. *Metabolism* **2012**, *61*, 1347–1352. [CrossRef] [PubMed]

Nutrients **2020**, 12, 1161 22 of 27

116. Gregersen, N.T.; Belza, A.; Jensen, M.G.; Ritz, C.; Bitz, C.; Hels, O.; Frandsen, E.; Mela, D.J.; Astrup, A. Acute effects of mustard, horseradish, black pepper and ginger on energy expenditure, appetite, ad libitum energy intake and energy balance in human subjects. *Br. J. Nutr.* **2013**, *109*, 556–563. [CrossRef] [PubMed]

- 117. Nammi, S.; Sreemantula, S.; Roufogalis, B.D. Protective effects of ethanolic extract of zingiber officinale rhizome on the development of metabolic syndrome in high-fat diet-fed rats. *Basic Clin. Pharmacol. Toxicol.* **2009**, *104*, 366–373. [CrossRef] [PubMed]
- 118. Kim, S.; Lee, M.S.; Jung, S.; Son, H.Y.; Park, S.; Kang, B.; Kim, S.Y.; Kim, I.H.; Kim, C.T.; Kim, Y. Ginger extract ameliorates obesity and inflammation via regulating microrna-21/132 expression and AMPK activation in white adipose tissue. *Nutrients* **2018**, *10*, 1567. [CrossRef]
- 119. Sayed, S.; Ahmed, M.; El-Shehawi, A.; Alkafafy, M.; Al-Otaibi, S.; El-Sawy, H.; Farouk, S.; El-Shazly, S. ginger water reduces body weight gain and improves energy expenditure in rats. *Foods* **2020**, *9*, 38. [CrossRef]
- 120. Misawa, K.; Hashizume, K.; Yamamoto, M.; Minegishi, Y.; Hase, T.; Shimotoyodome, A. Ginger extract prevents high-fat diet-induced obesity in mice via activation of the peroxisome proliferator-activated receptor δ pathway. *J. Nutr. Biochem.* **2015**, 26, 1058–1067. [CrossRef]
- 121. Wang, J.; Li, D.; Wang, P.; Hu, X.; Chen, F. Ginger prevents obesity through regulation of energy metabolism and activation of browning in high-fat diet-induced obese mice. *J. Nutr. Biochem.* **2019**, 70, 105–115. [CrossRef]
- 122. Deng, X.; Zhang, S.; Wu, J.; Sun, X.; Shen, Z.; Dong, J.; Huang, J. Promotion of mitochondrial biogenesis via activation of AMPK-PGC1<sup>α</sup> signaling pathway by ginger (Zingiber Officinale Roscoe) extract, and its major active component 6-gingerol. *J. Food Sci.* **2019**, *84*, 2101–2111. [CrossRef]
- 123. Wang, J.; Zhang, L.; Dong, L.; Hu, X.; Feng, F.; Chen, F. 6-gingerol, a functional polyphenol of ginger, promotes browning through an AMPK-dependent pathway in 3T3-L1 adipocytes. *J. Agric. Food Chem.* **2019**, 67, 14056–14065. [CrossRef] [PubMed]
- 124. Stanisiere, J.; Mousset, P.Y.; Lafay, S. How safe is ginger rhizome for decreasing nausea and vomiting in women during early pregnancy? *Foods* **2018**, *7*, 50. [CrossRef] [PubMed]
- 125. Turmeric|Description, History,&Uses. *Encycl. Br.* 2019. Available online: https://www.britannica.com/plant/turmeric (accessed on 30 October 2019).
- 126. Hay, E.; Lucariello, A.; Contieri, M.; Esposito, T.; De Luca, A.; Guerra, G.; Perna, A. Therapeutic effects of turmeric in several diseases: An overview. *Chem. Biol. Interact.* **2019**, *310*, 108729. [CrossRef] [PubMed]
- 127. Pan, Y.; Zhao, D.; Yu, N.; An, T.; Miao, J.; Mo, F.; Gu, Y.; Zhang, D.; Gao, S.; Jiang, G. Curcumin improves glycolipid metabolism through regulating peroxisome proliferator activated receptor γ signalling pathway in high-fat diet-induced obese mice and 3T3-L1 adipocytes. *R. Soc. Open Sci.* **2017**, *4*, 170917. [CrossRef] [PubMed]
- 128. Ding, L.; Li, J.; Song, B.; Xiao, X.; Zhang, B.; Qi, M.; Huang, W.; Yang, L.; Wang, Z. Curcumin rescues high fat diet-induced obesity and insulin sensitivity in mice through regulating srebp pathway. *Toxicol. Appl. Pharmacol.* 2016, 304, 99–109. [CrossRef]
- 129. Wang, S.; Wang, X.; Ye, Z.; Xu, C.; Zhang, M.; Ruan, B.; Wei, M.; Jiang, Y.; Zhang, Y.; Wang, L.; et al. Curcumin promotes browning of white adipose tissue in a norepinephrine-dependent way. *Biochem. Biophys. Res. Commun.* 2015, 466, 247–253. [CrossRef]
- 130. Lone, J.; Choi, J.H.; Kim, S.W.; Yun, J.W. Curcumin induces brown fat-like phenotype in 3T3-L1 and primary white adipocytes. *J. Nutr. Biochem.* **2016**, 27, 193–202. [CrossRef]
- 131. Peron, G.; Sut, S.; Dal Ben, S.; Voinovich, D.; Dall'Acqua, S. Untargeted UPLC-MS metabolomics reveals multiple changes of urine composition in healthy adult volunteers after consumption of Curcuma Longa L. Extract. *Food Res. Int. Ott. Ont* **2020**, 127, 108730. [CrossRef]
- 132. Zingg, J.-M.; Hasan, S.T.; Nakagawa, K.; Canepa, E.; Ricciarelli, R.; Villacorta, L.; Azzi, A.; Meydani, M. Modulation of CAMP levels by high-fat diet and curcumin and regulatory effects on CD36/FAT scavenger receptor/fatty acids transporter gene expression. *BioFactors Oxf. Engl.* 2017, 43, 42–53. [CrossRef]
- 133. Qiu, P.; Man, S.; Li, J.; Zhang, L.; Yu, P.; Gao, W. Overdose intake of curcumin initiates the unbalanced state of bodies. *J. Agric. Food Chem.* **2016**, *64*, 2765–2771. [CrossRef]
- 134. Soleimani, V.; Sahebkar, A.; Hosseinzadeh, H. Turmeric (Curcuma Longa) and its major constituent (curcumin) as nontoxic and safe substances: Review. *Phytother. Res. PTR* **2018**, 32, 985–995. [CrossRef] [PubMed]
- 135. Fernández-Aceñero, M.J.; Ortega Medina, L.; Maroto, M. Herbal drugs: Friend or foe? *J. Clin. Exp. Hepatol.* **2019**, *9*, 409–411. [CrossRef] [PubMed]

Nutrients **2020**, 12, 1161 23 of 27

136. Cinnamaldehyde. PubChem. Available online: https://pubchem.ncbi.nlm.nih.gov/compound/637511 (accessed on 30 October 2019).

- 137. Michlig, S.; Merlini, J.M.; Beaumont, M.; Ledda, M.; Tavenard, A.; Mukherjee, R.; Camacho, S.; le Coutre, J. Effects of TRP channel agonist ingestion on metabolism and autonomic nervous system in a randomized clinical trial of healthy subjects. *Sci. Rep.* **2016**, *6*, 20795. [CrossRef] [PubMed]
- 138. Hochkogler, C.M.; Hoi, J.K.; Lieder, B.; Müller, N.; Hans, J.; Widder, S.; Ley, J.P.; Somoza, V. Cinnamyl isobutyrate decreases plasma glucose levels and total energy intake from a standardized breakfast: A randomized, crossover intervention. *Mol. Nutr. Food Res.* **2018**, *62*, 1701038. [CrossRef] [PubMed]
- 139. Pandit, C.; Anilakumar, K.R. Cold adaptive thermogenesis following consumption of certain pungent spice principles: A validation study. *J. Therm. Biol.* **2017**, *64*, 35–40. [CrossRef] [PubMed]
- 140. Neto, J.G.O.; Boechat, S.K.; Romão, J.S.; Pazos-Moura, C.C.; Oliveira, K.J. Treatment with cinnamaldehyde reduces the visceral adiposity and regulates lipid metabolism, autophagy and endoplasmic reticulum stress in the liver of a rat model of early obesity. *J. Nutr. Biochem.* 2020, 77, 108321. [CrossRef]
- 141. Kaur, N.; Chugh, H.; Tomar, V.; Sakharkar, M.K.; Dass, S.K.; Chandra, R. Cinnamon attenuates adiposity and affects the expression of metabolic genes in diet-induced obesity model of zebrafish. *Artif. Cells Nanomed. Biotechnol.* **2019**, *47*, 2930–2939. [CrossRef]
- 142. Pandit, C.; Latha, S.S.; Rani, T.U.; Anilakumar, K.R. Pepper and cinnamon improve cold induced cognitive impairment via increasing non-shivering thermogenesis; a study. *Int. J. Hyperth.* **2018**, *35*, 518–527. [CrossRef]
- 143. Jiang, J.; Emont, M.P.; Jun, H.; Qiao, X.; Liao, J.; Kim, D.I.; Wu, J. Cinnamaldehyde induces fat cell-autonomous thermogenesis and metabolic reprogramming. *Metabolism* **2017**, 77, 58–64. [CrossRef]
- 144. Kwan, H.Y.; Wu, J.; Su, T.; Chao, X.-J.; Liu, B.; Fu, X.; Chan, C.L.; Lau, R.H.Y.; Tse, A.K.W.; Han, Q.B.; et al. Cinnamon induces browning in subcutaneous adipocytes. *Sci. Rep.* **2017**, *7*, 1–12. [CrossRef]
- 145. Clapham, D.E.; Julius, D.; Montell, C.; Schultz, G. International union of pharmacology. XLIX. Nomenclature and structure-function relationships of transient receptor potential channels. *Pharmacol. Rev.* **2005**, *57*, 427–450. [CrossRef] [PubMed]
- 146. Woehrlin, F.; Fry, H.; Abraham, K.; Preiss-Weigert, A. Quantification of flavoring constituents in cinnamon: High variation of coumarin in cassia bark from the german retail market and in authentic samples from Indonesia. *J. Agric. Food Chem.* **2010**, *58*, 10568–10575. [CrossRef] [PubMed]
- 147. Brancheau, D.; Patel, B.; Zughaib, M. Do cinnamon supplements cause acute hepatitis? *Am. J. Case Rep.* **2015**, 16, 250–254. [CrossRef] [PubMed]
- 148. Altschuler, J.A.; Casella, S.J.; MacKenzie, T.A.; Curtis, K.M. The effect of cinnamon on A1c among adolescents with type 1 diabetes. *Diabetes Care* **2007**, *30*, 813–816. [CrossRef]
- 149. Ranasinghe, P.; Jayawardena, R.; Pigera, S.; Wathurapatha, W.S.; Weeratunga, H.D.; Premakumara, G.A.S.; Katulanda, P.; Constantine, G.R.; Galappaththy, P. Evaluation of pharmacodynamic properties and safety of cinnamomum zeylanicum (Ceylon Cinnamon) in healthy adults: A phase i clinical trial. *BMC Complement. Altern. Med.* **2017**, *17*, 550. [CrossRef]
- 150. Maji, A.K.; Banerji, P. Phytochemistry and gastrointestinal benefits of the medicinal spice, Capsicum annuum L. (chilli): A review. *J. Complement. Integr. Med.* **2016**, *13*, 97–122. [CrossRef]
- 151. Zheng, J.; Zheng, S.; Feng, Q.; Zhang, Q.; Xiao, X. Dietary capsaicin and its anti-obesity potency: From mechanism to clinical implications. *Biosci. Rep.* **2017**, *37*, BSR20170286. [CrossRef]
- 152. Varghese, S.; Kubatka, P.; Rodrigo, L.; Gazdikova, K.; Caprnda, M.; Fedotova, J.; Zulli, A.; Kruzliak, P.; Büsselberg, D. Chili pepper as a body weight-loss food. *Int. J. Food Sci. Nutr.* **2017**, *68*, 392–401. [CrossRef]
- 153. Ludy, M.J.; Moore, G.E.; Mattes, R.D. The Effects of capsaicin and capsiate on energy balance: Critical review and meta-analyses of studies in humans. *Chem. Senses* **2012**, *37*, 103–121. [CrossRef]
- 154. Zsiborás, C.; Mátics, R.; Hegyi, P.; Balaskó, M.; Pétervári, E.; Szabó, I.; Sarlós, P.; Mikó, A.; Tenk, J.; Rostás, I.; et al. Capsaicin and capsiate could be appropriate agents for treatment of obesity: A meta-analysis of human studies. *Crit. Rev. Food Sci. Nutr.* **2018**, *58*, 1419–1427. [CrossRef]
- 155. Janssens, P.L.H.R.; Hursel, R.; Martens, E.A.P.; Westerterp-Plantenga, M.S. Acute effects of capsaicin on energy expenditure and fat oxidation in negative energy balance. *PLoS ONE* **2013**, *8*, e67786. [CrossRef] [PubMed]
- 156. Baskaran, P.; Krishnan, V.; Ren, J.; Thyagarajan, B. Capsaicin induces browning of white adipose tissue and counters obesity by activating TRPV1 channel-dependent mechanisms. *Br. J. Pharmacol.* **2016**, 173, 2369–2389. [CrossRef] [PubMed]

Nutrients **2020**, 12, 1161 24 of 27

157. Yoneshiro, T.; Saito, M. Transient receptor potential activated brown fat thermogenesis as a target of food ingredients for obesity management. *Curr. Opin. Clin. Nutr. Metab. Care* **2013**, *16*, 625–631. [CrossRef] [PubMed]

- 158. LiverTox: Clinical and Research Information on Drug-Induced Liver Injury [Internet]. National Institute of Diabetes and Digestive and Kidney Diseases: Bethesda, MD, USA, 2012. Available online: https://www.ncbi.nlm.nih.gov/books/NBK547852/ (accessed on 21 April 2020).
- 159. Semwal, R.B.; Semwal, D.K.; Vermaak, I.; Viljoen, A. A comprehensive scientific overview of Garcinia cambogia. *Fitoterapia* **2015**, *102*, 134–148. [CrossRef]
- 160. Fassina, P.; Scherer Adami, F.; Terezinha Zani, V.; Kasper Machado, I.C.; Garavaglia, J.; Quevedo Grave, M.T.; Ramos, R.; Morelo Dal Bosco, S. The effect of Garcinia cambogia as coadjuvant in the weight loss process. *Nutr. Hosp.* **2015**, *32*, 2400–2408. [CrossRef]
- 161. Haber, S.L.; Awwad, O.; Phillips, A.; Park, A.E.; Pham, T.M. Garcinia cambogia for weight loss. *Am. J. Health. Syst. Pharm.* **2018**, 75, 17–22. [CrossRef]
- 162. Kriketos, A.D.; Thompson, H.R.; Greene, H.; Hill, J.O. (-)-Hydroxycitric acid does not affect energy expenditure and substrate oxidation in adult males in a post-absorptive state. *Int. J. Obes.* 1999, 23, 867–873. [CrossRef]
- 163. Vasques, C.A.R.; Rossetto, S.; Halmenschlager, G.; Linden, R.; Heckler, E.; Fernandez, M.S.P.; Alonso, J.L.L. Evaluation of the pharmacotherapeutic efficacy of Garcinia cambogia plus Amorphophallus konjac for the treatment of obesity. *Phytother. Res.* **2008**, 22, 1135–1140. [CrossRef]
- 164. Kovacs, E.M.; Westerterp-Plantenga, M.S.; Saris, W.H. The effects of 2-week ingestion of (–)-hydroxycitrate and (–)-hydroxycitrate combined with medium-chain triglycerides on satiety, fat oxidation, energy expenditure and body weight. *Int. J. Obes.* 2001, 25, 1087–1094. [CrossRef]
- 165. Payab, M.; Hasani-Ranjbar, S.; Shahbal, N.; Qorbani, M.; Aletaha, A.; Haghi-Aminjan, H.; Soltani, A.; Khatami, F.; Nikfar, S.; Hassani, S.; et al. Effect of the herbal medicines in obesity and metabolic syndrome: A systematic review and meta-analysis of clinical trials. *Phytother. Res.* **2019**, *97*, e8825. [CrossRef]
- 166. Li, L.; Zhang, H.; Yao, Y.; Yang, Z.; Ma, H. (-)-Hydroxycitric acid suppresses lipid droplet accumulation and accelerates energy metabolism via activation of the adiponectin-AMPK signaling pathway in broiler chickens. *J. Agric. Food Chem.* **2019**, *67*, 3188–3197. [CrossRef] [PubMed]
- 167. Han, N.; Li, L.; Peng, M.; Ma, H. (-)-Hydroxycitric acid nourishes protein synthesis via altering metabolic directions of amino acids in male rats. *Phytother. Res.* **2016**, *30*, 1316–1329. [CrossRef] [PubMed]
- 168. Anton, S.D.; Shuster, J.; Leeuwenburgh, C. Investigations of botanicals on food intake, satiety, weight loss, and oxidative stress: A study protocol of a double-blind, placebo-controlled, crossover study. *Zhong Xi Yi Jie He Xue Bao* **2011**, *9*, 1190–1198. [CrossRef] [PubMed]
- 169. Corey, R.; Werner, K.T.; Singer, A.; Moss, A.; Smith, M.; Noelting, J.; Rakela, J. acute liver failure associated with Garcinia cambogia use. *Ann. Hepatol.* **2016**, *15*, 123–126. [CrossRef] [PubMed]
- 170. Melendez-Rosado, J.; Snipelisky, D.; Matcha, G.; Stancampiano, F. Acute hepatitis induced by pure Garcinia Cambogia. *J. Clin. Gastroenterol.* **2015**, *49*, 449–450. [CrossRef]
- 171. Lunsford, K.E.; Bodzin, A.S.; Reino, D.C.; Wang, H.L.; Busuttil, R.W. Dangerous dietary supplements: Garcinia cambogia-associated hepatic failure requiring transplantation. *World J. Gastroenterol.* **2016**, 22, 10071–10076. [CrossRef]
- 172. Allen, S.F.; Godley, R.W.; Evron, J.M.; Heider, A.; Nicklas, J.M.; Thomas, M.P. Acute necrotizing eosinophilic myocarditis in a patient taking Garcinia cambogia extract successfully treated with high-dose corticosteroids. *Can. J. Cardiol.* **2014**, *30*, e13–e15. [CrossRef]
- 173. Lobb, A. Hepatoxicity associated with weight-loss supplements: A case for better post-marketing surveillance. *World J. Gastroenterol. WJG* **2009**, *15*, 1786–1787. [CrossRef]
- 174. Marques, L.L.M.; Panizzon, G.P.; Aguiar, B.A.A.; Simionato, A.S.; Cardozo-Filho, L.; Andrade, G.; de Oliveira, A.G.; Guedes, T.A.; Mello, J.C. Guaraná (Paullinia cupana) seeds: Selective supercritical extraction of phenolic compounds. *Food Chem.* **2016**, *212*, 703–711. [CrossRef]
- 175. Smith, N.; Atroch, A.L. Guaraná's journey from regional tonic to aphrodisiac and global energy drink. *Evid. Based Complement. Altern. Med. ECAM* **2010**, 7, 279–282. [CrossRef]
- 176. Bérubé-Parent, S.; Pelletier, C.; Doré, J.; Tremblay, A. Effects of encapsulated green tea and guarana extracts containing a mixture of epigallocatechin-3-gallate and caffeine on 24 h energy expenditure and fat oxidation in Men. *Br. J. Nutr.* **2005**, *94*, 432–436. [CrossRef] [PubMed]

Nutrients **2020**, 12, 1161 25 of 27

177. Boozer, C.N.; Nasser, J.A.; Heymsfield, S.B.; Wang, V.; Chen, G.; Solomon, J.L. An herbal supplement containing ma huang-guarana for weight loss: A randomized, double-blind trial. *Int. J. Obes.* **2001**, 25, 316–324. [CrossRef] [PubMed]

- 178. Bortolin, R.C.; Vargas, A.R.; de Miranda Ramos, V.; Gasparotto, J.; Chaves, P.R.; Schnorr, C.E.; da Boit Martinello, K.; Silveira, A.K.; Gomes, H.M.; Rabelo, T.K.; et al. Guarana supplementation attenuated obesity, insulin resistance, and adipokines dysregulation induced by a standardized human western diet via brown adipose tissue activation. *Phytother. Res. PTR* **2019**, *33*, 1394–1403. [CrossRef] [PubMed]
- 179. Lima, N.D.S.; Numata, E.D.P.; Mesquita, L.M.S.; Dias, P.H.; Vilegas, W.; Gambero, A.; Ribeiro, M.L. Modulatory effects of guarana (Paullinia cupana) on adipogenesis. *Nutrients* **2017**, *9*, 635. [CrossRef]
- 180. Ciszowski, K.; Biedroń, W.; Gomólka, E. Acute caffeine poisoning resulting in atrial fibrillation after guarana extract overdose. *Przegl. Lek.* **2014**, *71*, 495–498.
- 181. Ishida, M.; Hara, M.; Fukino, N.; Kakizaki, T.; Morimitsu, Y. Glucosinolate Metabolism, Functionality and Breeding for the Improvement of Brassicaceae Vegetables. *Breed. Sci.* **2014**, *64*, 48–59. [CrossRef]
- 182. Martins, T.; Colaço, B.; Venâncio, C.; Pires, M.J.; Oliveira, P.A.; Rosa, E.; Antunes, L.M. Potential effects of sulforaphane to fight obesity. *J. Sci. Food Agric.* **2018**, *98*, 2837–2844. [CrossRef]
- 183. Glade, M.J.; Meguid, M.M. A Glance at ... Broccoli, glucoraphanin, and sulforaphane. *Nutrition* **2015**, *31*, 1175–1178. [CrossRef]
- 184. Kushad, M.M.; Brown, A.F.; Kurilich, A.C.; Juvik, J.A.; Klein, B.P.; Wallig, M.A.; Jeffery, E.H. Variation of glucosinolates in vegetable crops of Brassica oleracea. *J. Agric. Food Chem.* **1999**, 47, 1541–1548. [CrossRef]
- 185. Yao, A.; Shen, Y.; Wang, A.; Chen, S.; Zhang, H.; Chen, F.; Chen, Z.; Wei, H.; Zou, Z.; Shan, Y.; et al. Sulforaphane induces apoptosis in adipocytes via Akt/P70s6k1/Bad inhibition and ERK activation. *Biochem. Biophys. Res. Commun.* **2015**, 465, 696–701. [CrossRef]
- 186. Lee, J.H.; Moon, M.H.; Jeong, J.K.; Park, Y.G.; Lee, Y.J.; Seol, J.W.; Park, S.Y. Sulforaphane induced adipolysis via hormone sensitive lipase activation, regulated by AMPK signaling pathway. *Biochem. Biophys. Res. Commun.* 2012, 426, 492–497. [CrossRef] [PubMed]
- 187. Lei, P.; Tian, S.; Teng, C.; Huang, L.; Liu, X.; Wang, J.; Zhang, Y.; Li, B.; Shan, Y. Sulforaphane improves lipid metabolism by enhancing mitochondrial function and biogenesis in vivo and in vitro. *Mol. Nutr. Food Res.* **2019**, *63*, 1800795. [CrossRef] [PubMed]
- 188. Nagata, N.; Xu, L.; Kohno, S.; Ushida, Y.; Aoki, Y.; Umeda, R.; Fuke, N.; Zhuge, F.; Ni, Y.; Nagashimada, M.; et al. Glucoraphanin ameliorates obesity and insulin resistance through adipose tissue browning and reduction of metabolic endotoxemia in mice. *Diabetes* 2017, 66, 1222–1236. [CrossRef] [PubMed]
- 189. Xu, L.; Nagata, N.; Ota, T. Glucoraphanin: A broccoli sprout extract that ameliorates obesity-induced inflammation and insulin resistance. *Adipocyte* **2018**, 7, 218–225. [CrossRef] [PubMed]
- 190. Felker, P.; Bunch, R.; Leung, A.M. Concentrations of thiocyanate and goitrin in human plasma, their precursor concentrations in brassica vegetables, and associated potential risk for hypothyroidism. *Nutr. Rev.* **2016**, 74, 248–258. [CrossRef]
- 191. Xu, J.; Kulkarni, S.R.; Donepudi, A.C.; More, V.R.; Slitt, A.L. Enhanced Nrf2 activity worsens insulin resistance, impairs lipid accumulation in adipose tissue, and increases hepatic steatosis in leptin-deficient mice. *Diabetes* **2012**, *61*, 3208–3218. [CrossRef] [PubMed]
- 192. Flores-Mateo, G.; Rojas-Rueda, D.; Basora, J.; Ros, E.; Salas-Salvadó, J. Nut intake and adiposity: Meta-analysis of clinical trials. *Am. J. Clin. Nutr.* **2013**, *97*, 1346–1355. [CrossRef]
- 193. Agebratt, C.; Ström, E.; Romu, T.; Dahlqvist-Leinhard, O.; Borga, M.; Leandersson, P.; Nystrom, F.H. A randomized study of the effects of additional fruit and nuts consumption on hepatic fat content, cardiovascular risk factors and basal metabolic rate. *PLoS ONE* **2016**, *11*, e0147149. [CrossRef]
- 194. Duarte Moreira Alves, R.; Boroni Moreira, A.P.; Silva Macedo, V.; Brunoro Costa, N.M.; Gonçalves Alfenas, R.d.C.; Bressan, J. High-oleic peanuts increase diet-induced thermogenesis in overweight and obese men. *Nutr. Hosp.* **2014**, 29, 1024–1032. [CrossRef]
- 195. Gheflati, A.; Bashiri, R.; Ghadiri-Anari, A.; Reza, J.Z.; Kord, M.T.; Nadjarzadeh, A. The effect of apple vinegar consumption on glycemic indices, blood pressure, oxidative stress, and homocysteine in patients with type 2 diabetes and dyslipidemia: A randomized controlled clinical trial. *Clin. Nutr. ESPEN* **2019**, *33*, 132–138. [CrossRef]

Nutrients **2020**, 12, 1161 26 of 27

196. Siddiqui, F.J.; Assam, P.N.; de Souza, N.N.; Sultana, R.; Dalan, R.; Chan, E.S. Diabetes control: Is vinegar a promising candidate to help achieve targets? *J. Evid.-Based Integr. Med.* **2018**, 23, 2156587217753004. [CrossRef] [PubMed]

- 197. Bouderbala, H.; Kaddouri, H.; Kheroua, O.; Saidi, D. Anti-obesogenic effect of apple cider vinegar in rats subjected to a high fat diet. *Ann. Cardiol. Angeiol.* **2016**, *65*, 208–213. [CrossRef] [PubMed]
- 198. Kondo, T.; Kishi, M.; Fushimi, T.; Kaga, T. Acetic acid upregulates the expression of genes for fatty acid oxidation enzymes in liver to suppress body fat accumulation. *J. Agric. Food Chem.* **2009**, *57*, 5982–5986. [CrossRef] [PubMed]
- 199. Kulshreshtha, A.; Zacharia, A.J.; Jarouliya, U.; Bhadauriya, P.; Prasad, G.B.K.S.; Bisen, P.S. Spirulina in health care management. *Curr. Pharm. Biotechnol.* **2008**, *9*, 400–405. [CrossRef]
- 200. Moradi, S.; Ziaei, R.; Foshati, S.; Mohammadi, H.; Nachvak, S.M.; Rouhani, M.H. Effects of spirulina supplementation on obesity: A systematic review and meta-analysis of randomized clinical trials. *Complement. Ther. Med.* 2019, 47, 102211. [CrossRef]
- 201. Park, W.S.; Kim, H.J.; Li, M.; Lim, D.H.; Kim, J.; Kwak, S.S.; Kang, C.M.; Ferruzzi, M.G.; Ahn, M.J. Two classes of pigments, carotenoids and c-phycocyanin, in Spirulina powder and their antioxidant activities. *Molecules* **2018**, 23, 2065. [CrossRef]
- 202. Hamedifard, Z.; Milajerdi, A.; Reiner, Ž.; Taghizadeh, M.; Kolahdooz, F.; Asemi, Z. The effects of Spirulina on glycemic control and serum lipoproteins in patients with metabolic syndrome and related disorders: A systematic review and meta-analysis of randomized controlled trials. *Phytother. Res.* 2019, 33, 2609–2621. [CrossRef]
- 203. Hall, K.D.; Guo, J. Obesity energetics: Body weight regulation and the effects of diet composition. *Gastroenterology* **2017**, *152*, 1718–1727.e3. [CrossRef]
- 204. Martens, E.A.; Gonnissen, H.K.; Gatta-Cherifi, B.; Janssens, P.L.; Westerterp-Plantenga, M.S. Maintenance of energy expenditure on high-protein vs. high-carbohydrate diets at a constant body weight may prevent a positive energy balance. *Clin. Nutr.* **2015**, *34*, 968–975. [CrossRef]
- 205. Ebbeling, C.B.; Swain, J.F.; Feldman, H.A.; Wong, W.W.; Hachey, D.L.; Garcia-Lago, E.; Ludwig, D.S. Effects of dietary composition on energy expenditure during weight-loss maintenance. *JAMA* **2012**, 307, 2627–2634. [CrossRef]
- 206. Ebbeling, C.B.; Feldman, H.A.; Klein, G.L.; Wong, J.M.W.; Bielak, L.; Steltz, S.K.; Luoto, P.K.; Wolfe, R.R.; Wong, W.W.; Ludwig, D.S. Effects of a low carbohydrate diet on energy expenditure during weight loss maintenance: Randomized trial. *BMJ* 2018, 363, k4583. [CrossRef] [PubMed]
- 207. Cummings, D.E.; Foster-Schubert, K.E.; Overduin, J. Ghrelin and energy balance: Focus on current controversies. *Curr. Drug Targets* **2005**, *6*, 153–169. [CrossRef]
- 208. Mihalache, L.; Gherasim, A.; Niţă, O.; Ungureanu, M.C.; Pădureanu, S.S.; Gavril, R.S.; Arhire, L.I. Effects of ghrelin in energy balance and body weight homeostasis. *Hormones (Athens)* **2016**, *15*, 186–196. [CrossRef] [PubMed]
- 209. Hall, K.D. A review of the carbohydrate-insulin model of obesity. *Eur. J. Clin. Nutr.* **2017**, 71, 323–326. [CrossRef] [PubMed]
- 210. Ludwig, D.S.; Friedman, M.I. Increasing adiposity: Consequence or cause of overeating? *JAMA* **2014**, *311*, 2167–2168. [CrossRef]
- 211. Taubes, G. The Science of obesity: What do we really know about what makes us fat? An essay by Gary Taubes. *BMJ* **2013**, *346*, f1050. [CrossRef]
- 212. Hall, K.D.; Guo, J.; Speakman, J.R. Do Low-carbohydrate diets increase energy expenditure? *Int. J. Obes.* **2019**, *43*, 2350–2354. [CrossRef]
- 213. Nieuwdorp, M.; Gilijamse, P.W.; Pai, N.; Kaplan, L.M. Role of the microbiome in energy regulation and metabolism. *Gastroenterology* **2014**, *146*, 1525–1533. [CrossRef]
- 214. Duca, F.A.; Lam, T.K.T. Gut microbiota, nutrient sensing and energy balance. *Diabetes Obes. Metab.* **2014**, *16*, 68–76. [CrossRef]
- 215. Turnbaugh, P.J.; Ley, R.E.; Mahowald, M.A.; Magrini, V.; Mardis, E.R.; Gordon, J.I. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* **2006**, *444*, 1027–1031. [CrossRef]
- 216. Bohan, R.; Tianyu, X.; Tiantian, Z.; Ruonan, F.; Hongtao, H.; Qiong, W.; Chao, S. Gut microbiota: A potential manipulator for host adipose tissue and energy metabolism. *J. Nutr. Biochem.* **2019**, *64*, 206–217. [CrossRef] [PubMed]

Nutrients **2020**, 12, 1161 27 of 27

217. Rosenbaum, M.; Knight, R.; Leibel, R.L. The gut microbiota in human energy homeostasis and obesity. *Trends Endocrinol. Metab.* **2015**, *26*, 493–501. [CrossRef] [PubMed]

- 218. Murphy, E.F.; Cotter, P.D.; Healy, S.; Marques, T.M.; O'Sullivan, O.; Fouhy, F.; Clarke, S.F.; O'Toole, P.W.; Quigley, E.M.; Stanton, C.; et al. Composition and energy harvesting capacity of the gut microbiota: Relationship to diet, obesity and time in mouse models. *Gut* 2010, 59, 1635–1642. [CrossRef] [PubMed]
- 219. Pouteau, E.; Nguyen, P.; Ballèvre, O.; Krempf, M. Production rates and metabolism of short-chain fatty acids in the colon and whole body using stable isotopes. *Proc. Nutr. Soc.* **2003**, *62*, 87–93. [CrossRef]
- 220. Kocełak, P.; Zak-Gołąb, A.; Zahorska-Markiewicz, B.; Aptekorz, M.; Zientara, M.; Martirosian, G.; Chudek, J.; Olszanecka-Glinianowicz, M. Resting energy expenditure and gut microbiota in obese and normal weight subjects. *Eur. Rev. Med. Pharmacol. Sci.* 2013, 17, 2816–2821.
- 221. Gao, Z.; Yin, J.; Zhang, J.; Ward, R.E.; Martin, R.J.; Lefevre, M.; Cefalu, W.T.; Ye, J. Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* **2009**, *58*, 1509–1517. [CrossRef]
- 222. Hanatani, S.; Motoshima, H.; Takaki, Y.; Kawasaki, S.; Igata, M.; Matsumura, T.; Kondo, T.; Senokuchi, T.; Ishii, N.; Kawashima, J.; et al. Acetate Alters expression of genes involved in beige adipogenesis in 3T3-L1 cells and obese KK-Ay mice. *J. Clin. Biochem. Nutr.* **2016**, *59*, 207–214. [CrossRef]
- 223. Okla, M.; Wang, W.; Kang, I.; Pashaj, A.; Carr, T.; Chung, S. Activation of Toll-like Receptor 4 (TLR4) attenuates adaptive thermogenesis via endoplasmic reticulum stress. *J. Biol. Chem.* **2015**, 290, 26476–26490. [CrossRef]
- 224. Cao, W.; Huang, H.; Xia, T.; Liu, C.; Muhammad, S.; Sun, C. Homeobox A5 promotes white adipose tissue browning through inhibition of the Tenascin C/Toll-Like Receptor 4/Nuclear Factor Kappa B inflammatory signaling in mice. *Front. Immunol.* **2018**, *9*, 647. [CrossRef]
- 225. Thaiss, C.A.; Itav, S.; Rothschild, D.; Meijer, M.T.; Levy, M.; Moresi, C.; Dohnalová, L.; Braverman, S.; Rozin, S.; Malitsky, S.; et al. Persistent microbiome alterations modulate the rate of post-dieting weight regain. *Nature* **2016**, *540*, *544*–*551*. [CrossRef]
- 226. Christensen, L.; Roager, H.M.; Astrup, A.; Hjorth, M.F. Microbial enterotypes in personalized nutrition and obesity management. *Am. J. Clin. Nutr.* **2018**, *108*, 645–651. [CrossRef]
- 227. Hjorth, M.F.; Roager, H.M.; Larsen, T.M.; Poulsen, S.K.; Licht, T.R.; Bahl, M.I.; Zohar, Y.; Astrup, A. Pre-treatment microbial Prevotella-to-Bacteroides ratio, determines body fat loss success during a 6-month randomized controlled diet intervention. *Int. J. Obes.* 2005 2018, 42, 580–583. [CrossRef]
- 228. Kovatcheva-Datchary, P.; Nilsson, A.; Akrami, R.; Lee, Y.S.; de Vadder, F.; Arora, T.; Hallen, A.; Martens, E.; Björck, I.; Bäckhed, F. Dietary fiber-induced improvement in glucose metabolism is associated with increased abundance of Prevotella. *Cell Metab.* 2015, 22, 971–982. [CrossRef] [PubMed]
- 229. Benítez-Páez, A.; Kjølbæk, L.; Pulgar, E.M.G.; Brahe, L.K.; Astrup, A.; Matysik, S.; Schött, H.F.; Krautbauer, S.; Liebisch, G.; Boberska, J.; et al. A multi-omics approach to unraveling the microbiome-mediated effects of arabinoxylan oligosaccharides in overweight humans. *mSystems* **2019**, *4*, e00209–e00219. [CrossRef] [PubMed]
- 230. Johnson, A.J.; Vangay, P.; Al-Ghalith, G.A.; Hillmann, B.M.; Ward, T.L.; Shields-Cutler, R.R.; Kim, A.D.; Shmagel, A.K.; Syed, A.N.; Walter, J.; et al. Daily sampling reveals personalized diet-microbiome associations in humans. *Cell Host Microbe* **2019**, 25, 789–802.e5. [CrossRef] [PubMed]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).