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# Thermogenic adipose tissue in older adults with obesity: a narrative review of mechanisms, brown fat resistance, and the translational relevance of exercise and nutrition

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Population aging is accelerating, and obesity is becoming more common in older adults, creating a growing clinical burden. Brown adipose tissue (BAT), together with inducible beige adipocytes in white fat depots, supports adaptive thermogenesis through substrate oxidation and helps clear glucose and lipids from the circulation. Aging and obesity often coexist, reducing BAT volume and activity while lowering the browning capacity of white adipose tissue. Thermogenic responsiveness to cold exposure, exercise, and diet-related signals appears attenuated in this context. This pattern is often described as brown fat resistance. In real-life settings, it may make older adults' responses to lifestyle interventions less consistent and less pronounced. This narrative review synthesizes evidence from animal models and human studies to delineate the principal mechanisms by which exercise and nutrition shape BAT and beige adipose biology. Relevant English-language articles published up to December 2025 were identified through PubMed, Scopus, and Web of Science, and screened according to their topical and methodological relevance. We also examine how age-related limits, including sarcopenia, chronic low-grade inflammation, weaker sympathetic and  $\beta$ -adrenergic signaling, and endocrine imbalance, raise the activation threshold and reduce thermogenic responses. Much mechanistic evidence comes from rodent studies or young, metabolically healthy populations, whereas human findings in obese older adults remain heterogeneous because of differences in endpoints, cold-stimulation protocols, phenotype characterization, and small sample sizes. Based on this evidence, we outline an integrated framework that links structured exercise with thermogenesis-supportive nutrition, while cautioning against overinterpretation of surrogate imaging readouts. The aim is to make thermogenic adipose activation more feasible in obese older adults by improving whole-body metabolic conditions and substrate handling, strengthening inter-organ communication, and increasing adipose tissue sensitivity to external triggers. We also highlight the need for future trials to use clear prespecified stratification suitable for older adults, and to establish standard safety checks and

monitoring plans that support practical and individualized strategies targeting BAT and beige adipose tissue.

#### KEYWORDS

beige adipocytes, brown adipose tissue, brown fat resistance, exercise, nutrition

## Introduction

Global population aging is accelerating, and obesity is becoming more common (1, 2). Older adults now make up a growing share of people living with obesity, and the impact on cardiovascular and metabolic health in later life is becoming clearer (3). Excess adiposity worsens insulin resistance and dyslipidaemia and commonly coexists with age related loss of skeletal muscle mass and function, producing the clinical phenotype termed sarcopenic obesity (4, 5).

Adipose tissue remodeling is a critical, yet frequently overlooked, driver of metabolic decline with aging (6). Thermogenic adipocytes, including classical brown adipocytes in brown adipose tissue and inducible beige adipocytes within white adipose depots, dissipate energy as heat and contribute to systemic glucose and lipid disposal (7). In humans, brown adipose tissue persists into adulthood and is typically identified in the supraclavicular, cervical, mediastinal, and paravertebral regions (8–10). With advancing age, brown adipose tissue volume and activity generally decrease, whereas visceral white adipose tissue more readily expands and acquires an inflammatory phenotype with reduced metabolic competence (11). Landmark PET/CT studies published in 2009 demonstrated metabolically active brown adipose tissue in adults, overturning the prior assumption that it was restricted to infants and small mammals (9, 10). Subsequent work confirmed that both classical brown adipose tissue and recruitable beige adipocytes express uncoupling protein 1 (UCP1), are enriched in mitochondria, and, when activated, can increase energy expenditure and improve metabolic regulation (12). Collectively, these observations support thermogenic adipose tissue as a biologically plausible target for interventions aimed at obesity and related metabolic disorders. However, existing syntheses have largely treated the exercise, nutrition, and mechanistic BAT literatures separately, mostly drawing on studies in young, metabolically healthy individuals. The translational implications for obese older adults, whose BAT biology is shaped by aging, adiposity, comorbidity, and polypharmacy, therefore remain insufficiently examined. As a result, it is still unclear which candidate stimuli remain biologically plausible under age- and obesity-related constraints, and which are unlikely to succeed without prior improvement of the systemic milieu.

Most mechanistic insights into brown adipose tissue activation, white adipose tissue browning, and thermogenesis-promoting nutrients are derived from animal models and younger cohorts. Therefore, interpretation in obese older adults should emphasize endpoint context, phenotype differences, and feasibility constraints. Older individuals often differ in physiological reserve, cardiometabolic comorbidity burden, medication exposure, and body composition, which can modify thermogenic readouts and

intervention responses. Age-associated systemic alterations may raise the activation threshold of thermogenic adipose tissue, which can make lifestyle effects more heterogeneous across older adults (13).

In this review, we integrate available evidence on exercise and nutrition interventions relevant to brown adipose tissue activation in older adults and argue that intervention development should be anchored in the central concept of age related brown fat resistance. For this review, age-related brown fat resistance refers to a tissue-level phenotype in obese older adults characterized, relative to metabolically matched younger individuals, by a higher threshold for thermogenic recruitment, weaker responses to physiological cues, and reduced durability of induced beige phenotypes after stimulus withdrawal. A fuller definition and its distinction from related constructs are presented in the section on age-related brown fat resistance. Using this core concept, the present review aims to inform intervention programs for obese older adults that are both biologically sound and clinically feasible. The present synthesis differs from earlier narrative reviews in three ways. First, it brings the exercise and nutrition literatures together within a single framework of age-related brown fat resistance, rather than treating them separately. Second, it makes explicit where findings from rodent studies or younger cohorts may not translate well to obese older adults. Third, it extends this mechanistic synthesis into a phenotype-informed, safety-aware intervention framework that prioritizes preparation of the systemic milieu over high-intensity or pharmacologic stimuli that may be poorly tolerated. Taken together, these features shift the focus from whether BAT can be activated in principle to the conditions under which such activation may be achievable and clinically meaningful in obese older adults.

## Methods

This narrative review was conducted with reference to the Scale for the Assessment of Narrative Review Articles (SANRA). PubMed/MEDLINE, Scopus, and Web of Science Core Collection were searched from database inception to 31 December 2025 for English-language, peer-reviewed publications. The search strategy combined terms related to thermogenic adipose tissue with terms related to aging or obesity, exercise, and nutrition, using AND between concept blocks and OR within each block. Searches were performed in title and abstract fields in PubMed, TITLE-ABS-KEY in Scopus, and topic fields in Web of Science. The database-specific search records, search-field settings, and Boolean concept blocks are summarized in the [Supplementary Material 1](#) to improve methodological transparency, while recognizing that this article is a structured narrative review rather than a systematic review or meta-analysis.

Eligible records included peer-reviewed original studies and relevant reviews examining exercise-related, nutrition-related, or age-related effects on brown or beige adipose tissue. Exclusion criteria were abstracts, editorials, letters, preprints, duplicate records, studies without a clear focus on brown or beige adipose tissue, studies limited to fetal or neonatal populations, and records for which the full text was not accessible.

After deduplication, two authors independently screened titles, abstracts, and keywords, followed by detailed relevance assessment of potentially relevant records, including full-text review when necessary. Disagreements were resolved through discussion or, when needed, consultation with a third author. Reference lists of included articles and key reviews were also screened manually. Given the methodological heterogeneity of the included literature, findings were synthesized narratively across four themes: thermogenic adipose biology, age-related brown fat resistance, exercise-mediated modulation, and the effects of nutrition or bioactive compounds. The selection process is summarized in a simplified PRISMA-style flow diagram in [Supplementary Figure 1](#).

Figure preparation. All schematic figures ([Figures 1–5](#)) were created with BioRender (BioRender.com; Science Suite Inc., Toronto, Canada) under a BioRender Academic Publication License granted to H.Q., which permits publication in academic journals and sublicensing under open-access models such as CC-BY 4.0, consistent with the open-access policy of *Frontiers in Nutrition*. The corresponding citation URLs are provided in the legend of each figure, and signed Confirmation of Publication and Licensing Rights documents have been provided to the Editorial Office.

## Thermogenic adipose tissue: brown and beige fat

### Brown adipose tissue: canonical and non-canonical thermogenic pathways

Brown adipose tissue (BAT) is a principal adaptive thermogenic organ that contributes to thermoregulation and energy homeostasis by increasing substrate oxidation and converting chemical energy into heat (14). BAT is richly vascularized and receives dense sympathetic innervation (15). Brown adipocytes characteristically contain multilocular lipid droplets and abundant mitochondria, with uncoupling protein 1 (UCP1) localized to the inner mitochondrial membrane (16).

In addition to heat production, BAT activation promotes the clearance of circulating fatty acids and glucose, thereby modulating systemic lipid and glycemic homeostasis (17). Because human investigations of thermogenic adipose responses largely depend on imaging based readouts and surrogate physiological measures, [Box 1](#) summarizes the interpretation of these endpoints and the major sources of between study variability.

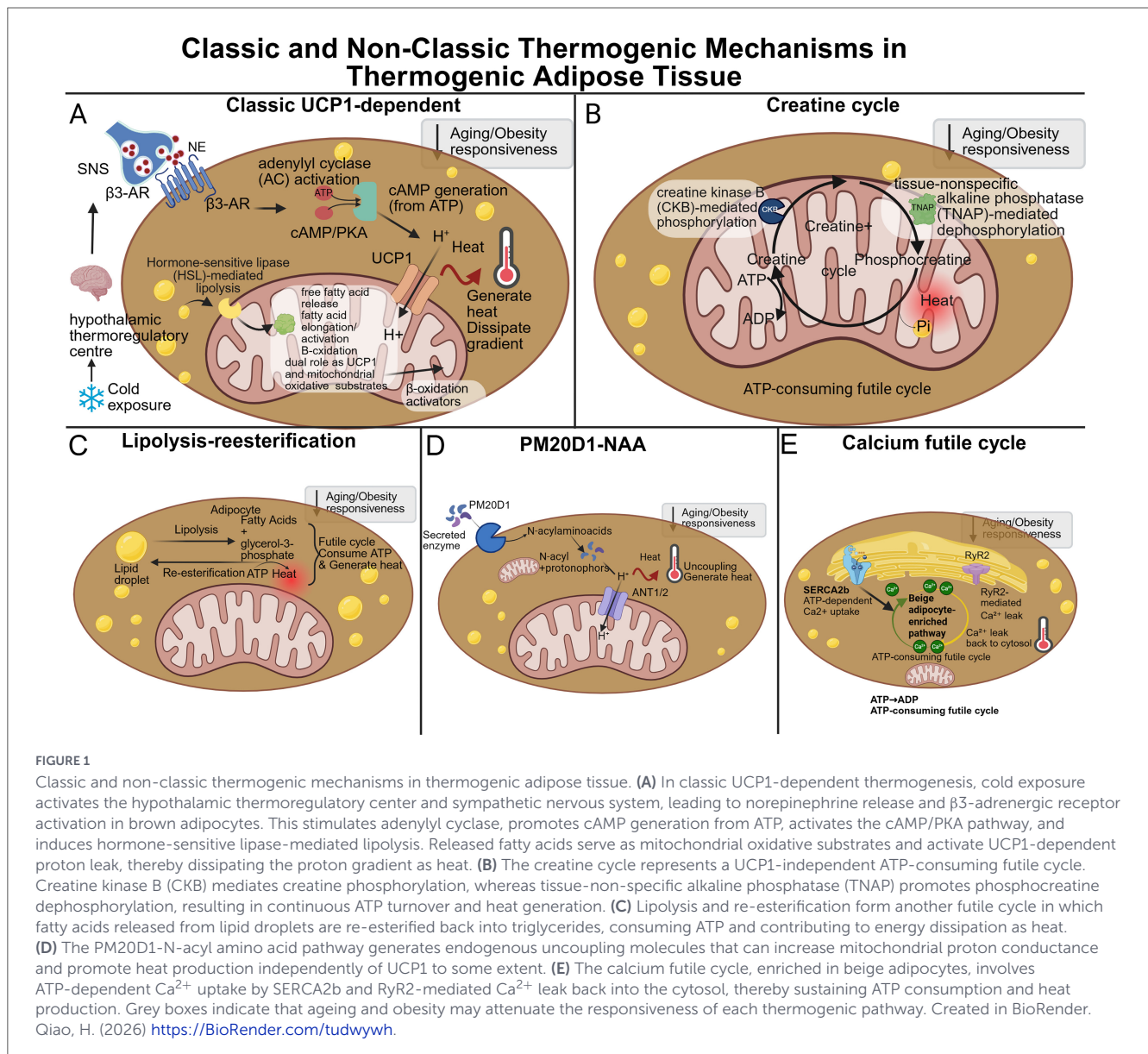
Canonical BAT thermogenesis is initiated predominantly by cold triggered sympathetic outflow (18). Upon cold exposure, sympathetic nerve terminals innervating BAT release norepinephrine (NE), which binds  $\beta$ 3-adrenergic receptors on brown adipocytes. Receptor activation engages G $\alpha$ s-coupled signaling to stimulate adenylate cyclase, which catalyzes the

conversion of ATP to cyclic AMP (cAMP); the resulting elevation of intracellular cAMP activates protein kinase A (PKA), which in turn stimulates lipolysis and upregulates thermogenic gene programs (19). PKA-driven lipolysis of intracellular triglycerides releases free fatty acids (FFAs); long-chain FFAs are further modified by adipocyte-expressed fatty-acid elongases and, upon reaching the mitochondrial inner membrane, function both as mitochondrial oxidative substrates and as activators of UCP1 dependent proton conductance across the inner mitochondrial membrane (20). By increasing proton leak, UCP1 uncouples electron transport from ATP synthesis and dissipates the electrochemical gradient as heat (21). In aggregate, fatty acid oxidation together with UCP1 mediated uncoupling constitutes the core UCP1 dependent thermogenic mechanism in BAT.

Available human evidence indicates that BAT detectability and cold-activated BAT activity decline with advancing age and greater adiposity, and preliminary cold-exposure PET data further suggest lower BAT uptake in older than in younger men; accordingly, obese older adults would be expected to exhibit a blunted BAT response to cold exposure (22–25). This attenuated responsiveness is linked not only to lower detectable BAT volume but also to functional deficits, including a higher activation threshold, reduced tissue perfusion, and compromised mitochondrial performance (11). Structural loss and functional impairment often develop at the same time, so imaging-based estimates may not fully capture true physiological capacity. These limits together can restrict BAT recruitment in obese older adults and reduce its role in energy balance and metabolic homeostasis. As a result, approaches that depend on a strong sympathetic drive to activate BAT may show less consistent efficacy in this population. This pattern reflects a recurring inconsistency in the field. Rodent cold-exposure studies typically show robust BAT recruitment, increased UCP1 expression, and higher energy expenditure, whereas human studies, particularly in obese or older adults, often report small, heterogeneous, or null effects on  $^{18}\text{F}$ -FDG-defined BAT activity. The gap likely reflects both interspecies differences and the fact that rodent experiments are usually conducted in young, lean animals under short, intense cold challenges, conditions far removed from the aging, polypharmacy-exposed, and chronically inflamed background of obese older adults. Rodent findings should therefore be treated as hypothesis-generating, and human data should remain the main basis for clinical interpretation. [Box 2](#) expands on these translational considerations, and [Table 1](#) summarizes the preclinical and clinical evidence base for the principal exercise and nutrition interventions discussed in the subsequent sections.

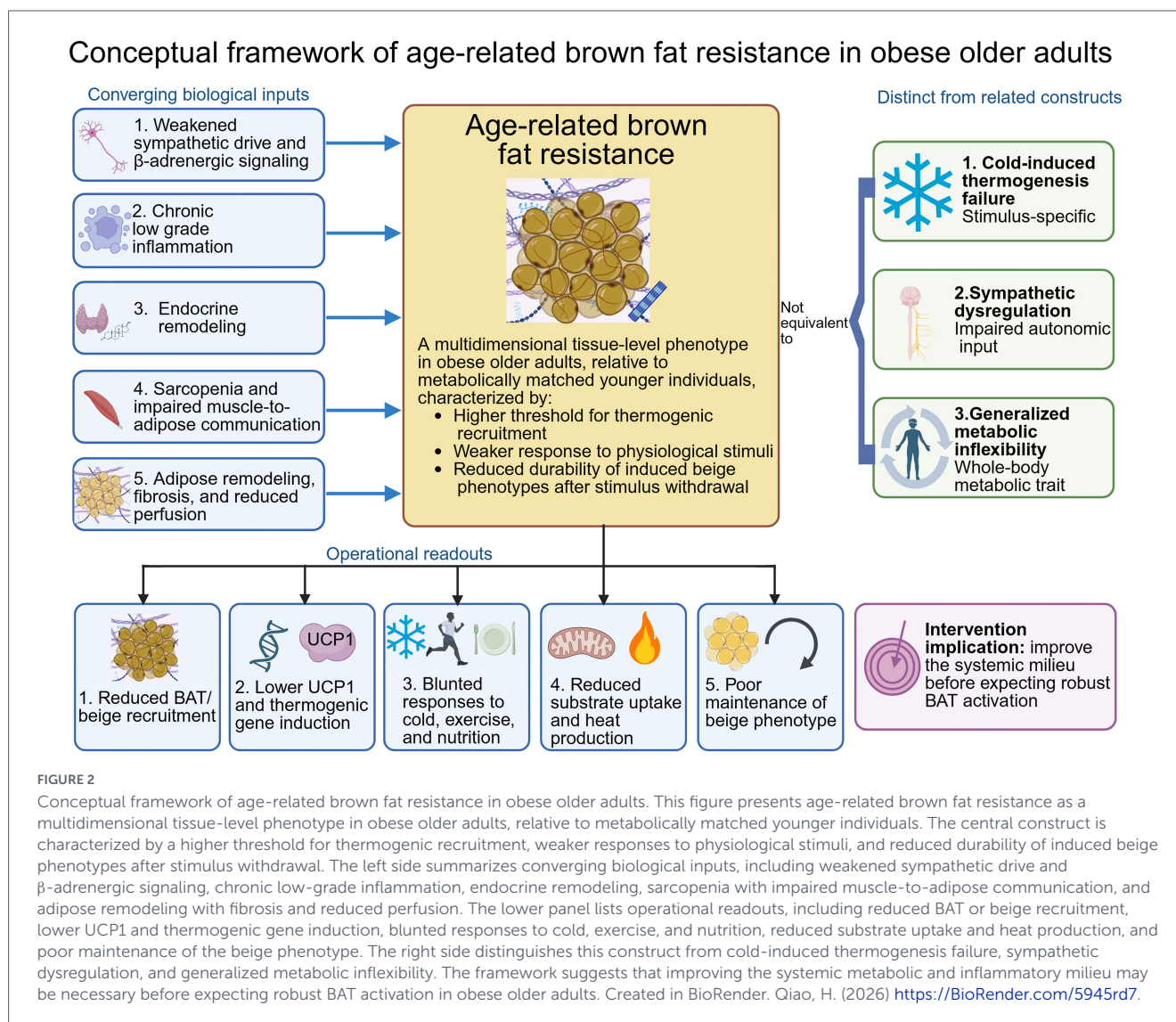
Pharmacological approaches aligned with canonical BAT activation typically target  $\beta$ 3 adrenergic receptors to recapitulate cold driven sympathetic signaling (26). Available evidence suggests that  $\beta$ 3 adrenergic agonists can modestly increase BAT activity and yield metabolic benefits (27, 28), yet reported effects in humans are heterogeneous (29). Several studies report cardiovascular safety signals (e.g., elevations in heart rate and blood pressure), which may necessitate careful participant selection and rigorous safety monitoring in obese older adults with cardiometabolic risk (30–32). Accordingly, evaluation in well-characterized cohorts with prespecified safety surveillance may be the most informative near-term pathway for this population.

As shown in [Figure 1](#), in addition to UCP1 dependent uncoupling, brown and beige adipose tissue have been reported to



engage multiple UCP1 independent routes of energy dissipation (33). These mechanisms typically operate through elevated ATP demand or inefficient substrate cycling and may partially compensate when UCP1 mediated thermogenesis is constrained (34). Four illustrative pathways are frequently discussed. First, the creatine linked phosphocreatine cycle can increase ATP turnover and thereby promote heat production (35). Seminal work from the Kazak group has defined the molecular basis of this futile cycle: mitochondrial creatine kinase B (CKB) utilizes ATP to phosphorylate creatine to phosphocreatine (36), while mitochondrially localized tissue-non-specific alkaline phosphatase (TNAP) hydrolyzes phosphocreatine back to creatine and inorganic phosphate (37). The coordinated, opposing activity of CKB and TNAP drives continuous ADP regeneration that fuels thermogenic respiration in the absence of net chemical work. Preclinical studies associate this pathway with improved cold tolerance and higher energy expenditure, whereas findings in humans remain variable (38). Second, a futile lipid cycle, characterized by simultaneous lipolysis and re-esterification, consumes ATP

continuously and dissipates energy as heat (39), and several experimental models suggest that this process may be upregulated when UCP1 capacity is limited (40, 41). Third, the PM20D1 associated N-acyl amino acid pathway generates bioactive lipids that act as endogenous uncouplers, increasing proton conductance and supporting thermogenesis independently of UCP1 to a certain extent (42). Fourth, as originally revealed by the Kajimura group, a calcium futile cycle operates prominently in beige adipocytes: SERCA2b actively imports Ca<sup>2+</sup> into the endoplasmic/sarcoplasmic reticulum at the expense of ATP hydrolysis, while Ca<sup>2+</sup> is simultaneously released back into the cytosol via ryanodine receptor 2 (RyR2), generating a continuous ATP-consuming Ca<sup>2+</sup> cycle that produces heat and contributes to systemic glucose homeostasis even when UCP1 is absent (43). A recent follow-up study from the same group identified the ER-membrane peptide C4orf3 (also known as another-regulin, ALN) as a molecular determinant of this process; C4orf3/ALN uncouples SERCA2b-mediated Ca<sup>2+</sup> transport from its ATP hydrolysis, rendering the SERCA2b-C4orf3 complex exothermic,



and its genetic loss diminishes adipose thermogenesis and increases adiposity in mice (44). Current human studies have not yet established the relative contribution of these non-canonical pathways, particularly in obese older adults. Mechanistic studies using standardized endpoints would help clarify their clinical relevance for metabolic outcomes.

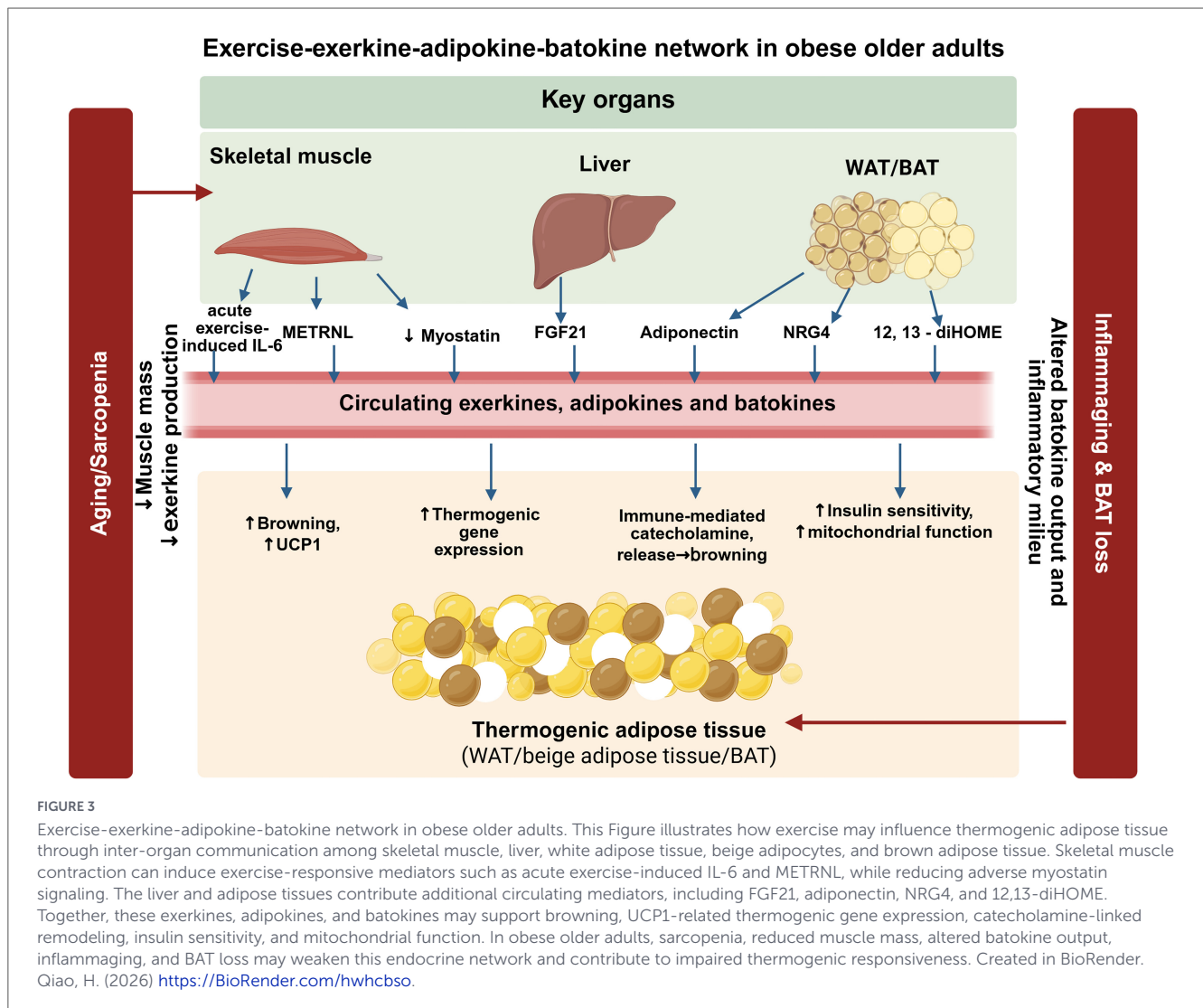
Accordingly, in obese older adults, a pragmatic strategy is to prioritize sustained exercise and dietary modification rather than depending on high intensity, short lived stimuli to elicit transient BAT activation (45). Over time, these interventions can improve basal metabolic function and the local tissue milieu, which may increase adipose responsiveness to physiologic cues and support more consistent, durable metabolic benefits.

## Adipose tissue browning, whitening, and loss of plasticity

In addition to classical brown adipose tissue, white adipose tissue in adult humans and rodents can acquire UCP1 positive beige adipocytes in response to specific stimuli, thereby gaining

inducible thermogenic capacity (46, 47). Beige adipocytes exhibit features intermediate between white and brown fat, and their lipid droplet architecture can shift between multilocular and unilocular patterns as environmental conditions change (48). This responsiveness indicates that browning is not a fixed conversion, but a reversible and dynamic process that can regress after stimulus withdrawal (49).

Two principal cellular sources have been proposed. Their relative contribution appears to vary by depot, and much of the mechanistic evidence derives from rodent models (50). Beige adipocytes may arise from precursor populations within white adipose depots, and under browning stimuli such as cold exposure or  $\beta$ 3-adrenergic stimulation, adipocytes within these depots may acquire beige-like features (51, 52). In subcutaneous adipose tissue, particularly in preclinical models, prolonged cold exposure or  $\beta$ 3-adrenergic stimulation can increase beige adipocyte recruitment and thermogenic activation, and this phenotype can lose thermogenic gene expression and multilocular morphology and shift toward a less thermogenic state, including a masked beige-like state described in rodent studies, once the stimulus is removed (51, 52). These observations indicate that maintenance of the beige phenotype depends on continued stimulation and that



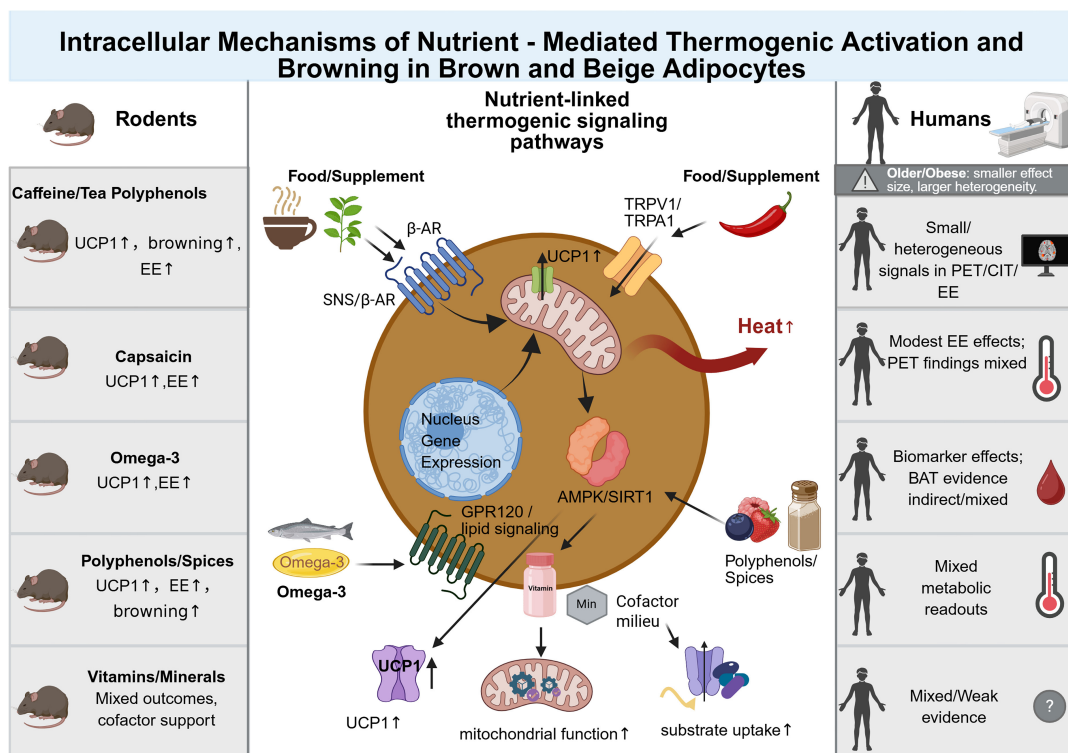
this plasticity may be especially vulnerable to aging- and obesity-related constraints.

Beige adipocyte induction depends on several regulatory inputs that act together, rather than one single stepwise pathway (50). Sympathetic signaling plays a major upstream role, while endocrine factors and nuclear receptor driven gene programs can also turn on the browning machinery (53). Exercise is one modifiable factor, and it may support white adipose browning through muscle to adipose communication (54). Available evidence suggests that some myokines can raise UCP1 expression and increase mitochondrial gene networks (55). The variation across human studies likely reflects differences in design, stimulus strength, tissue sampling, and outcome measures. Individual differences, depot specific adipose features, and prior exposure history also shape beige adipocyte recruitment and functional capacity (56, 57).

When these mechanisms are considered in humans, thermogenic adipose tissue must be viewed as a heterogeneous system. Adult human thermogenic adipose depots, particularly in the supraclavicular region, may contain overlapping features of classical brown and recruitable beige/brite adipocytes (58). In experimental and preclinical settings, beige cell recruitment during white fat browning is more commonly observed in

subcutaneous depots, whereas visceral fat typically shows more limited browning capacity (59, 60). Adiposity-related adipose niche remodeling may further shape depot heterogeneity (61). Even when thermogenic adipose tissue is difficult to detect at baseline, prolonged cold acclimation can still lead to measurable metabolic activation, suggesting that many adults retain an inducible thermogenic reserve.

Human studies also lack shared criteria for defining brown and beige adipogenesis. Some studies focus on tissue based evidence, such as histology and gene signals like higher UCP1 and mitochondrial gene expression, while other studies infer activity from imaging based metabolic readouts (62, 63). These methods reflect different biological aspects, so differences in methods likely add to the mixed results seen across populations. A clear example of this definitional heterogeneity is the dissociation reported in several adult cohorts between UCP1 abundance in supraclavicular adipose biopsies and  $^{18}\text{F}$ -FDG uptake in the same depot. High UCP1 expression does not always coincide with increased glucose uptake under cold stimulation, and detectable  $^{18}\text{F}$ -FDG uptake may occur without clearly enriched UCP1 signal. Because these measures reflect different aspects of thermogenic biology, studies relying on only one readout may reach different conclusions about



**FIGURE 4**  
 Intracellular Mechanisms of nutrient-mediated thermogenic activation and browning in brown and beige adipocytes. This figure summarizes candidate thermogenic nutrients and dietary bioactive compounds according to their main pathway nodes and compares the strength of evidence between rodent and human studies. The left column summarizes typical rodent findings, in which caffeine or tea polyphenols, capsaicin, omega-3 fatty acids, polyphenols or spices, and selected vitamins or minerals have been associated with changes in UCP1 expression, browning markers, energy expenditure, or mitochondrial support. The central panel illustrates putative nutrient-linked signaling pathways, including sympathetic and β-adrenergic signaling, TRPV1/TRPA1 activation, GPR120 or lipid-mediated signaling, AMPK/SIRT1-related mitochondrial adaptation, and cofactor support for substrate uptake and oxidative metabolism. The right column highlights that human evidence is generally smaller, more heterogeneous, and often based on surrogate readouts such as PET uptake, cold-induced thermogenesis, energy expenditure, circulating biomarkers, or metabolic endpoints. In obese older adults, these nutrients should therefore be interpreted as adjunctive modulators of a permissive metabolic milieu rather than stand-alone thermogenic interventions. Created in BioRender. Qiao, H. (2026) <https://BioRender.com/5cyedzb>.

BAT activation. In obese older adults, this limitation is amplified by the low baseline detectability of thermogenic adipose tissue, which may partly account for conflicting results across otherwise comparable studies.

Beige adipocytes also rely more on continued external input to remain stable than classical brown adipose tissue. Cold exposure, β3 adrenergic agonists, and exercise can support beige adipocyte recruitment, yet this phenotype can fade over time, and de-browning may occur when these inputs stop (64–66). This instability matters in aging and obesity. Aging can lower baseline beige adipocyte levels and reduce the ability to recruit and maintain them during stimulation, while obesity related energy excess, low physical activity, and chronic metabolic stress can further push adipose depots toward a less thermogenic and less flexible state (67–69). In obese older adults, these combined effects often appear as higher activation thresholds and weaker durability of the beige phenotype, which limits the role of thermogenic adipose tissue in energy balance and metabolic homeostasis.

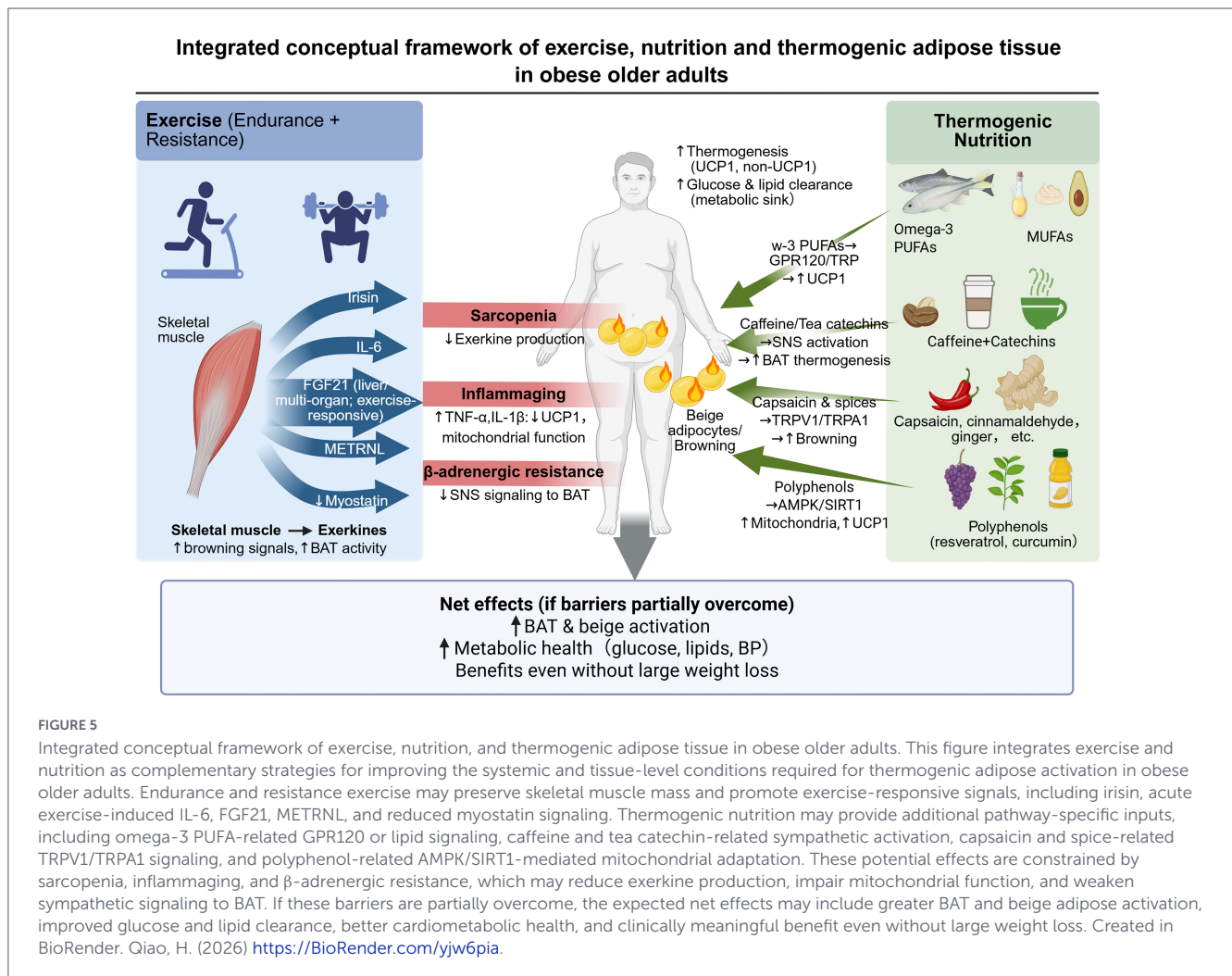
In summary, obese older adults often show a reduced ability to start and sustain white adipose browning, which is a key feature of brown fat resistance. The next sections use this framework to clarify when exercise and nutrition strategies are most likely to help,

with the aim of guiding practical weight management approaches for obese older adults.

### Age-related brown fat resistance: mechanisms in obese older adults

In older adults, both the ability to recruit brown adipose tissue and the tendency of white adipose tissue to undergo browning generally decline with age, a pattern often described as brown fat resistance (70, 71). This impairment in thermogenic function likely reflects converging mechanisms, including weakened sympathetic signaling, chronic low-grade inflammation, reduced endocrine support, sarcopenia with impaired muscle–adipose communication, and structural remodeling of adipose tissue (72–74).

For this review, age-related brown fat resistance is defined as a multidimensional, tissue-level phenotype in obese older adults that, compared with metabolically matched younger individuals, shows a higher threshold for thermogenic recruitment, a weaker response to physiological stimuli, and less durable maintenance of induced beige phenotypes.



This construct is distinct from failure of cold-induced thermogenesis, which is stimulus-specific, from sympathetic dysregulation, which reflects impaired autonomic input, and from generalized metabolic inflexibility, which describes a whole-body metabolic trait rather than a thermogenic adipose phenotype. Instead, brown fat resistance refers to the integrated tissue-level consequence of concurrent decline in sympathetic drive, endocrine support, muscle-to-adipose signaling, and adipose niche integrity with aging and obesity. Figure 2 summarizes these inputs and their measurable readouts, and each is discussed below.

## Sympathetic drive and impaired signaling

Aging is associated with alterations in autonomic regulation that collectively reduce effective sympathetic input to thermogenic adipose tissue (75). Although basal sympathetic tone may rise as a compensatory response to weight gain or insulin resistance,  $\beta$  adrenergic receptor responsiveness and downstream signal transduction within adipose depots commonly decline. In parallel, local neural density and norepinephrine signaling efficiency in aged fat can be reduced (76–78). Consequently, greater sympathetic activity in obese older adults does not always yield proportionate increases in adipose thermogenesis.

These changes help explain why canonical stimuli, including cold exposure and catecholaminergic activation, often show limited capacity to sustain robust induction of UCP1 and related thermogenic gene programs in aged adipocytes. Attenuated norepinephrine signaling may also impair BAT perfusion and substrate delivery during cold challenge, further limiting the thermogenic response (79–81).

## Chronic low-grade inflammation

With advancing age, the adipose tissue microenvironment shifts toward a more pro-inflammatory profile, characterized by increased immune cell infiltration and higher levels of mediators such as TNF  $\alpha$  and IL 1 $\beta$  (82, 83). This inflammatory milieu can impair mitochondrial function and blunt the induction of thermogenic gene programs, including UCP1. In parallel, inflammation promotes fibrotic remodeling, which disrupts tissue architecture and makes the microenvironment less conducive to browning and sustained heat production. As a result, effective thermogenic activation often requires stronger or more prolonged stimuli.

Chronic low grade inflammation can blunt thermogenic gene induction, and cytokine dynamics should be interpreted in

**TABLE 1** Preclinical versus clinical evidence for the principal lifestyle, nutritional, and pharmacological-adjunct interventions discussed, with recommendations for obese older adults.

Intervention	Preclinical (rodent) evidence	Human evidence	Recommendation in obese older adults
Aerobic exercise	In rodent models, endurance training has been reported to increase adipocyte precursor populations in BAT; UCP1-related findings are not uniform across studies.	RCTs show improved fitness and body composition; FDG-PET BAT signals often null in young healthy adults; data in obese older adults are limited.	Core intervention. Benefit should be judged from cardiometabolic and functional endpoints rather than BAT imaging.
Resistance training	Not emphasized as a distinct preclinical BAT literature in this review.	Most established clinical value is lean mass preservation; BAT imaging evidence is mixed.	Core intervention, particularly for sarcopenic obesity. Thermogenic benefit is indirect.
Caffeine	Mechanism: adenosine receptor blockade and increased sympathetic signaling.	Short-term studies show small rises in resting energy use and modest increases in supraclavicular skin temperature.	Optional adjunct. Careful safety checks in older adults (sleep, anxiety, blood pressure).
Capsaicin/TRPV1 agonists	Mechanism: TRPV1 channel activation with engagement of sympathetic-related signaling.	Modest effects on BMI, body weight, and waist circumference in meta-analysis of RCTs in adults with overweight or obesity.	Adjunctive dietary component; effects are modest and tolerance should be considered.
Resveratrol	Multiple rodent studies associate resveratrol with increased adipose oxidative capacity and UCP1-related pathway upregulation.	Small human studies report thermogenic gene-expression changes, whereas weight and metabolic outcomes remain inconsistent.	Not recommended as routine intervention; bioavailability limits translation.
Curcumin	Browning markers and AMPK activation reported in rodents.	Small RCTs show modest weight or glycemic effects; direct human BAT evidence lacking.	Evidence that curcumin produces sustained thermogenic benefit in humans remains preliminary.
Omega-3 PUFA (EPA/DHA)	Upregulates UCP1-related markers and mitochondrial pathways in rodents.	Most consistent effects at the population level are on cardiometabolic biomarkers (triglycerides, inflammation); effects on BAT are indirect.	May be considered within a Mediterranean-style dietary pattern; $\geq 250$ mg/day EPA + DHA is a commonly used nutritional target rather than a BAT-specific dose.
Vitamin D	Depot- and context-dependent thermogenic effects in rodents.	No clear association between 25(OH)D and BAT metabolism in recent healthy adult cohort.	Correct deficiency only; do not frame as thermogenic intervention.
Iron	Mouse studies link iron handling to BAT mitochondrial respiration; ferritin heavy-chain knockout improves adaptive thermogenesis.	Human TFRC expression has been associated with obesity-related adipose phenotypes and UCP1 expression; observational only.	Human evidence should be interpreted with caution; phenotype-specific effects likely across aging and obesity.
B vitamins/thiamine	<i>In vitro</i> and rodent data on thermogenic cofactor roles.	Emerging human adipose SLC19A3 data; deficiency common in bariatric cohorts.	Correct deficiency; no role for high-dose supplementation.
$\beta$ 3-adrenergic agonists (e.g., mirabegron)	$\beta$ 3-AR activation recapitulates cold-driven sympathetic signaling to BAT (mechanistic basis).	Modest, heterogeneous BAT activation in humans; cardiovascular safety signals reported.	Investigational. Use only with careful cardiovascular monitoring and phenotype-informed selection.

light of exposure context. The former more commonly reflects ongoing metabolic stress and is associated with constraints on thermogenesis, whereas the latter may function as a context dependent regulator of energy metabolism (84–86). Overall, persistent inflammation is generally associated with an inflammatory–fibrotic milieu that hinders initiation and maintenance of thermogenic transcriptional programs.

## Endocrine and hormonal environment changes

Age related endocrine remodeling may reduce the sensitivity of brown and beige adipose tissue to external stimuli. Lower thyroid hormone availability or diminished tissue responsiveness has been linked in some studies to impaired thermogenic capacity, consistent with the role of triiodothyronine in regulating UCP1 expression and promoting mitochondrial biogenesis (87, 88). Declining sex steroid levels may also contribute. For example, estrogen related variation has been associated with redistribution of adipose depots

and differences in detectable thermogenic fat, although results across cohorts are not fully concordant (89–91).

Alterations in the growth hormone and insulin like growth factor 1 axis may further influence thermogenic potential indirectly by affecting skeletal muscle maintenance and overall metabolic status (92). In parallel, insulin resistance and compensatory hyperinsulinemia are more common in later life, and this metabolic context may reduce the efficiency with which thermogenic adipose tissue is recruited in response to physiologic cues (93). Taken together, attenuation of sympathetic and thyroid linked signaling, combined with broader hormonal shifts in aging, may generate an endocrine milieu that is less permissive for effective thermogenic fat activation.

## Sarcopenia and weakened muscle-fat communication

Aging is commonly accompanied by declines in skeletal muscle mass and function, which reduce resting energy expenditure and

**BOX 1** Interpreting human evidence on thermogenic adipose tissue

(BAT/TAT)

Endpoints

- <sup>18</sup>F-FDG PET/CT uptake is a proxy of glucose utilization and BAT detectability under specific conditions; it does not always translate into quantified heat production and is influenced by insulin/feeding state and skeletal muscle glucose uptake.
- Fatty-acid tracers/oxidative metrics more closely reflect substrate utilization, but protocols are heterogeneous and less common in older cohorts.
- Indirect calorimetry and skin temperature are relatively non-specific and can be influenced by ambient temperature, clothing, and peripheral blood flow.
- Circulating 'brown/beige' factors are often correlational or acute signals and, on their own, are insufficient to confirm sustained tissue thermogenesis without complementary endpoints.
- Protocol Heterogeneity
- Ambient temperature, season, cold-stimulation dose/duration, and shivering control determine detectability and effect size.
- Phenotype/medication confounding
- Adiposity, insulin resistance/diabetes, thyroid status, inflammation, and perfusion modify BAT readouts.
- $\beta$ -blockers and other drugs affecting sympathetic tone or metabolism may attenuate BAT responsiveness, which is common in older adults.
- Depot Biology
- Human depots exhibit heterogeneity; aging/obesity-associated remodeling diminishes recruitable capacity and imaging signal.
- Interpretation hierarchy
- In obese older adults, prioritize body composition (muscle preservation), insulin sensitivity, cardiometabolic markers, and function over BAT surrogates.
- Minimal reporting checklist
- Cold/ambient temperature + shivering control; season.
- Fasting/feeding status; recent physical activity; sleep/caffeine intake control.
- Key phenotype/comorbidities; medication background.
- Endpoint type + limitations; safety/tolerability.

**BOX 2** Preclinical-to-clinical translation in obese older adults

A recurring challenge in interpreting this field is that the mechanistic framework of thermogenic adipose tissue has been established largely in young, lean rodents under short, intense cold challenges below thermoneutrality, whereas the clinical population of interest is older, obese, exposed to polypharmacy, and chronically inflamed, and is usually studied under near-thermoneutral conditions. Three specific translational gaps therefore require caution: (i) the short, intense cold exposures used in preclinical work rarely match the milder, more variable exposures that are tolerable in older adults, so effect sizes cannot be assumed to scale directly; (ii) preclinical studies are dominated by UCP1-related readouts, whereas human studies more often rely on <sup>18</sup>F-FDG uptake, a surrogate of glucose uptake that does not always correspond to UCP1 expression; and (iii) most cited preclinical studies are conducted in young, lean animals, so findings have to be extrapolated to the older, obese, polypharmacy-exposed phenotype of the target clinical population. Throughout this review, preclinical findings are therefore treated as hypothesis-generating, and statements about clinical applicability are anchored, where possible, in human evidence. When only animal data are available, this is stated explicitly, and [Table 1](#) summarizes the evidence base for the principal interventions discussed.

may attenuate exercise related metabolic signaling from muscle to adipose tissue. Experimental work suggests that exercise evoked myogenic mediators can participate in beige adipocyte recruitment and modulate thermogenic responses, yet much of the supporting evidence is currently limited to animal models and *in vitro* systems (94). In aging, particularly in the setting of sarcopenia and habitual inactivity, impaired muscle secretory output and weakened muscle-to-adipose signaling may hinder engagement of thermogenic programs within adipose depots, thereby contributing to brown fat resistance; however, direct evidence in older adults remains limited, and much of the mechanistic support derives from animal studies (95–97).

## Adipose tissue remodeling and fibrosis

With advancing age, adipose extracellular matrix remodeling becomes more prominent and is commonly accompanied by progressive fibrosis, reduced angiogenic capacity, and greater vulnerability to local hypoxia (98, 99). Increased collagen deposition and cross linking stiffen the matrix and reshape the mechanical microenvironment, which can impair beige adipocyte recruitment and survival and weaken maintenance of thermogenic transcriptional programs (100, 101). At the same time, diminished perfusion limits oxygen and nutrient delivery, exacerbating metabolic stress and cellular injury, intensifying inflammatory signaling, and promoting further fibrotic remodeling. Reduced

capillary density additionally constrains the substrate and oxygen supply required for high rate oxidative metabolism, thereby making sustained thermogenic activation more difficult to achieve (102, 103).

In summary, these interacting constraints likely underlie the widely observed phenotype of brown fat resistance in obese older adults. Compared with younger individuals, recruitment of thermogenic adipose tissue in response to common triggers such as cold exposure, physical training, or diet based cues is typically blunted and shows greater interindividual heterogeneity. Accordingly, a key limitation in obese older adults is often not the lack of stimuli, but reduced tissue sensitivity and a diminished capacity to translate signals into a robust thermogenic program.

In this context, exercise and nutritional strategies may be more readily implementable. Interventions that reduce chronic inflammatory burden, improve microvascular perfusion and substrate handling, preserve skeletal muscle function, and support adrenergic signaling alongside mitochondrial adaptation may help create a more permissive physiological milieu for initiating and sustaining thermogenic activity.

Human evidence in obese older adults is frequently inferred from imaging based readouts and surrogate physiological measures, and protocol differences in temperature control, stimulation dose, and participant phenotype can produce divergent conclusions. Future studies should prioritize standardized reporting of ambient and cold exposure conditions, prespecified subgroup stratification, and mechanistic endpoints that better link thermogenic biology to clinically meaningful metabolic and functional outcomes.

## Exercise and thermogenic adipose tissue

Direct intervention evidence on exercise-induced thermogenic adipose adaptations in obese older adults remains limited; accordingly, the following discussion integrates available human

findings with mechanistic and translational evidence from broader adult and preclinical literature. An overview of the exercise–exerkine–adipokine–batokine network discussed in this section is provided in [Figure 3](#). Regular physical activity plays a key role in healthy aging and supports body composition, cardiovascular risk factors, and insulin sensitivity (104–106). Skeletal muscle contraction raises energy use and heat production (107), which may briefly lower the need for brown adipose tissue mediated heat generation. At the same time, regular exercise activates signaling pathways that can help maintain the longer term function of both brown and beige adipose tissue (108, 109).

Even with these mechanistic considerations, reported exercise effects on human BAT remain variable across cohorts and protocols. In obese older adults, exercise most often leads to better body composition, improved metabolic control, and stronger physical function. Whether BAT imaging signals or other surrogate indices change reproducibly with exercise should be interpreted in light of [Box 1](#), with careful attention to endpoint limitations and variability in study protocols and experimental conditions.

Three factors may underlie these heterogeneous findings: age-related biological changes, differences in metabolic status, and methodological variation across studies. Declines in  $\beta$ 3-adrenergic signaling, myokine output, and mitochondrial biogenesis may raise the threshold for thermogenic remodeling (110–112), while inflammation, insulin resistance, and visceral adiposity may suppress thermogenic programs, including UCP1-related pathways, and may impair PGC-1 $\alpha$ -associated mitochondrial regulation (62, 113). At the same time, differences in imaging modality, cold-acclimation protocol, post-exercise assessment, training dose, and biopsy depot can influence whether such changes are detected. These findings suggest that thermogenic responses are conditional rather than absent, supporting the depot-, phenotype-, and protocol-specific interpretation adopted below.

In obese older adults characterized by brown fat resistance, exercise is best conceptualized as a multi target intervention (114–116). Resistance training and combined training support preservation of skeletal muscle mass and may influence metabolic regulation through exercise evoked myogenic signals, including transient elevations in irisin and IL 6 (117–119). Aerobic training improves endothelial function and insulin sensitivity (120, 121), thereby facilitating substrate delivery to thermogenic depots, and sustained training can reduce inflammatory inhibition of thermogenic gene programs (122, 123). Accordingly, even when imaging derived indices of BAT show minimal change, as discussed in [Box 1](#), exercise based programs may still yield meaningful improvements in obesity related metabolic health in older populations.

## Endocrine mediators of exercise: myokines and brown adipokines

A key mechanism by which exercise modulates adipose biology involves myokine secretion. During contraction, skeletal muscle releases peptides and metabolites into the circulation that can act on distal targets, including adipose depots (124). Several myokines have been implicated in promoting thermogenic activity.

**Myokine irisin:** Irisin was first identified as a PGC-1 $\alpha$ -dependent myokine generated by proteolytic cleavage of FNDC5

and linked to brown-fat-like development of white adipose tissue (125). In adipocyte studies, irisin has been reported to engage p38 MAPK and ERK signaling, thereby increasing PGC-1 $\alpha$  activity and UCP1-related transcription and supporting the browning program in white adipocytes (126). In mouse models, exogenous irisin administration can increase energy expenditure and ameliorate diet induced metabolic dysfunction (127). In humans, studies have reported exercise associated changes in circulating irisin and correlations with imaging derived indices of brown adipose activity (128, 129), but findings are inconsistent across assays, training paradigms, and participant characteristics, highlighting the need for further validation. In obese older adults, particularly those with concomitant sarcopenia, irisin responsiveness may depend strongly on training modality and dose (130). Whether irisin-related signaling translates into durable thermogenic phenotypic changes in obese older adults requires further well-controlled studies.

**Interleukin 6 (IL 6):** IL 6 is a prototypical exercise responsive cytokine that rises transiently during endurance exercise and is associated with increased lipolysis, greater circulating fatty acid availability, and short term improvements in insulin sensitivity (131). In thermogenic adipose tissue, IL 6 has been implicated in regulation of heat production and systemic metabolic control. In animal models, IL 6 deficiency is associated with impaired cold induced thermogenesis (132), and transplantation studies suggest that adipose derived IL 6 can modulate glucose homeostasis through pathways linked to FGF21 (133). It remains unclear whether exercise-induced IL-6 modifies FGF21 production through intermediary organs (e.g., the liver) and thereby influences brown adipose function.

**Meteorin like (METRNL):** METRNL is released from skeletal muscle in response to exercise and is also expressed in adipose tissue during cold exposure (134). Preclinical studies indicate that METRNL can improve insulin sensitivity and attenuate inflammation and may facilitate beige adipocyte recruitment through immune mediated adipose remodeling, including shifts toward an anti inflammatory milieu enriched in M2 like macrophage profiles (135). In obese mouse models, increased METRNL expression has been associated with greater white adipose browning and improved metabolic parameters (134). Human investigations have reported associations between circulating METRNL and markers linked to thermogenic adipose activity (136), and some studies observe higher levels after endurance training (137). In obese older adults, the durability of METRNL-related signaling is likely shaped by inflammatory status and training dose, which may influence the persistence of beige-like phenotypes.

**Fibroblast growth factor 21 (FGF21):** FGF21 is produced by multiple organs and contributes to systemic metabolic regulation, with circulating concentrations influenced by exercise and cold exposure. It has been proposed to facilitate white adipose browning, increase UCP1 related transcription, and enhance fatty acid oxidative capacity (138–140). Although higher basal FGF21 levels are frequently observed in older adults, this does not necessarily indicate increased pathway signaling, and several studies suggest that stimulus responsiveness may be altered in a manner that depends on the underlying metabolic phenotype (141–143). Exercise associated FGF21 signaling may support adaptations relevant to browning resistance by improving substrate handling and modulating catecholamine sensitivity. Whether older adults

with obesity exhibit functional FGF21 resistance, and how this axis responds to exercise based interventions, remains a key question for well-characterized elderly cohorts.

**Neuregulin 4 (NRG4):** Unlike the mediators described above, NRG4 is not a myokine but a brown adipokine — an epidermal growth factor (EGF)-family ligand that is highly enriched in BAT, is further induced during brown adipocyte differentiation and cold-driven recruitment of beige adipocytes in white adipose tissue, and signals principally through ErbB4/ErbB3 receptors on the liver and other peripheral targets (144). Gain- and loss-of-function studies in rodents established that NRG4 attenuates hepatic *de novo* lipogenesis via suppression of SREBP-1c, protects against diet-induced insulin resistance and hepatic steatosis, and supports a healthier systemic adipokine profile alongside enhanced fuel oxidation (144, 145). Adipose expression and circulating concentrations of NRG4 are consistently reduced in rodent and human obesity, in metabolic syndrome, and with advancing age, suggesting that circulating NRG4 may track the functional status of thermogenic adipose tissue (146). Although NRG4 originates from BAT rather than contracting muscle, it responds to exercise training: in a 12-week randomized trial in men with obesity, high-intensity interval training and circuit resistance training elicited greater increases in serum NRG4 than moderate-intensity continuous training, alongside parallel gains in body composition and cardiometabolic risk factors (147). This exercise responsiveness is biologically consistent with evidence that structured training can partially restore BAT functionality and positions NRG4 as a candidate circulating read-out of brown-fat-liver crosstalk in obese older adults. Human observational data nonetheless remain heterogeneous, with some clinical studies reporting discordant associations between circulating NRG4 and insulin sensitivity, so translation to aging populations will require confirmation with prespecified endpoints and standardized assays (146).

Collectively, these myogenic mediators together with the brown adipokine NRG4 support endocrine communication between skeletal muscle, thermogenic adipose depots, and the liver and represent a major route by which exercise modulates lipid metabolism and thermogenic capacity. Repeated training may partially offset sarcopenia related deficits in inter tissue signaling and improve the systemic milieu that enables adipose metabolic adaptation.

Myokine and adipokine signaling patterns are strongly shaped by age and metabolic status (148). Aging is associated with lower PGC-1 $\alpha$  expression and a blunted FNDC5/irisin response, and chronic inflammation and insulin resistance may further impair adipose responsiveness to myokine signaling. Findings are also shaped by methodological factors, including sampling time, assay specificity, fasting state, circadian timing, and concurrent medication use. Accordingly, the inconsistent results reported in obese older adults likely reflect phenotype-dependent biology superimposed on measurement variability rather than a true absence of exercise-responsive signaling.

Several investigations propose that exercise associated signaling contributes to initiation or persistence of thermogenic transcriptional programs within adipose tissue (149–151). Human evidence is mainly derived from observational designs and surrogate outcomes, so causal interpretation should remain cautious, especially in obese older adults. Correlational signals should be presented as hypotheses that require confirmation with

standardized endpoints and longitudinal designs. Thermogenic adipose tissue may also release mediators that support bidirectional regulation between fat and other organs. For example, experimental studies associate brown adipose derived interleukin 6 with improved systemic glucose homeostasis (152–154). In addition, 12,13-dihydroxy-9Z-octadecenoic acid (12,13-diHOME), an oxylipin derived from linoleic acid and released by BAT in response to cold exposure or exercise, has been reported to increase fatty acid uptake into BAT and skeletal muscle (155). Together, these observations support the possibility of coordinated cross-tissue signaling between skeletal muscle and thermogenic adipose depots.

## Effects of different exercise modalities

Aerobic exercise and resistance training may affect brown and beige adipose function through partially distinct mechanisms, and in applied settings a combined program is often more appropriate. Aerobic training commonly improves cardiorespiratory performance and insulin sensitivity and can enhance substrate handling, thereby creating systemic conditions that may facilitate recruitment of thermogenic adipose depots (156). The magnitude of exercise evoked myokine signaling varies by age and metabolic phenotype.

In rodent models, endurance training can attenuate high fat diet induced metabolic dysfunction and, in some settings, has been reported to increase the abundance of adipocyte precursor populations within brown adipose tissue (157). These observations raise the possibility that endurance exercise influences adipose plasticity and cellular reserve, with human studies showing variable signals depending on study design and endpoint selection. Findings across animal studies are not uniform. Some report increased UCP1 related expression in subcutaneous adipose tissue, whereas others observe minimal change, with discrepancies plausibly attributable to differences in ambient temperature, training load, and species or strain characteristics (158, 159).

Human studies suggest that moderate-intensity aerobic exercise alone does not consistently produce sustained increases in FDG-PET-derived indices of brown adipose activity (160). In a large randomized trial, 24 weeks of combined endurance and resistance training improved cardiorespiratory fitness and body composition in young participants, while no measurable changes were detected in brown fat volume or glucose uptake under the specific endpoints and conditions used (161). One frequently cited interpretation is that exercise induced heat production and higher energy expenditure may transiently reduce the need for non-shivering thermogenesis, such that imaging signals may not fully reflect functional adaptations (162). Because this trial enrolled young, metabolically healthy individuals, response patterns may differ in older adults or in those with cardiometabolic impairment. More broadly, the discrepancy between null imaging findings and positive preclinical data likely reflects the interaction of cohort phenotype, intervention dose, and environmental conditions. Rodent studies are typically conducted in young animals below thermoneutrality, where exercise can readily augment BAT browning, whereas human trials are usually performed near thermoneutrality, where this effect is less readily detected. In obese older adults, low baseline BAT detectability, a higher effective thermoneutral zone, and common use of  $\beta$ -adrenergic dampening

medications may further blunt imaging-detectable activation. As a result, phenotypically heterogeneous older cohorts are likely to show variable group-level effects even when individual responders are present.

For obese older adults, aerobic training may be more likely to enhance thermogenic adipose performance through indirect mechanisms, including reductions in visceral adiposity, improved insulin sensitivity, and better tissue perfusion that supports substrate delivery. Therefore, even if its acute mobilizing effect is weaker than cold exposure, sustained training can optimize the systemic milieu and allow residual brown or beige adipose tissue to operate closer to its functional capacity. Divergent conclusions across studies are often attributable to differences in endpoint properties and experimental conditions.

Resistance training primarily promotes preservation and accrual of skeletal muscle mass. By mitigating sarcopenia, it can enhance metabolic reserve and may support production and amplification of exercise responsive myogenic mediators (163). Available data also suggest that resistance exercise can influence anabolic endocrine profiles, potentially contributing to a more permissive hormonal milieu (164).

Evidence linking resistance training to BAT imaging or functional thermogenic outcomes is mixed, and most studies have not focused on obese older adults. High intensity resistance sessions can induce transient catecholamine elevations, which could in principle participate in thermogenic recruitment, but implementation in older adults must be guided by safety considerations and dose feasibility (165, 166). For this population, the most established clinical value of resistance training is maintenance or increase of lean mass, support of resting metabolic rate, and indirect facilitation of thermogenic adipose function through improved substrate handling and a more favorable systemic metabolic environment.

Combined exercise regimens, particularly programs integrating aerobic and resistance components, are widely implemented to improve cardiorespiratory capacity and muscular strength concurrently. Conceptually, this approach is well suited to the multi domain needs of obesity in later life, as it can enhance vascular function and insulin sensitivity while preserving or increasing skeletal muscle mass.

Preclinical data further suggest that pairing exercise with adjunct interventions may augment browning related phenotypes (167). In some animal models, polyphenol supplements paired with exercise training have been linked to larger increases in browning markers in white adipose tissue than either approach on its own (168). This pattern supports the idea of testing combined strategies in human studies. For now, these results should be framed as working hypotheses and research priorities, not as firm conclusions that can be applied directly to people.

When prescribing exercise for obese older adults, safety and a plan that fits the individual matter most. Frail people, and those with heart or blood vessel problems, may not cope well with high intensity exercise or strong cold exposure. A practical starting plan can include two to three light to moderate resistance sessions each week that work the main muscle groups, along with moderate aerobic activity such as brisk walking or water based exercise. A person can increase the load and time step by step as physical capacity improves.

The main goal is to protect muscle and heart health through long term adherence, which can improve whole body conditions that support thermogenic responses. Exercise can improve blood glucose control, lower inflammation, and support blood vessel structure and function.

Across available exercise studies, heterogeneity in training dose, baseline adipose detectability, and endpoint selection remains a key barrier to synthesis in obese older adults. Future trials should incorporate consistent thermogenic outcome definitions, control for temperature and recent activity, and evaluate functional and cardiometabolic endpoints alongside thermogenic surrogates to improve interpretability and translational value. Human evidence in obese older adults remains limited in quantity, and the available studies often involve small sample sizes, with substantial heterogeneity in temperature settings, direct assessments of thermogenic function, and clinically relevant outcome measures. Future studies should plan subgroup analyses in advance, so researchers can look at sex, menopausal status, medication use, baseline detectability of thermogenic adipose tissue, and the presence of sarcopenic obesity. This approach can help identify who is more likely to respond and can guide more suitable exercise plans. For individuals unable to achieve sufficient training volume, pharmacological options or exercise-mimetic approaches may be considered, with implementation best guided by structured safety monitoring and phenotype-informed stratification.

Taken together, the literature suggests that the apparent inconsistency in exercise induced thermogenic responses largely reflects three factors: age related biology, metabolic status, and methodological variability. Reduced sympathetic signaling,  $\beta$  adrenergic desensitization, lower myokine secretion, and declining mitochondrial and progenitor cell reserve may limit responsiveness, while chronic inflammation, insulin resistance, and ectopic lipid accumulation further constrain thermogenic programs. At the same time, differences in training dose, cold exposure and imaging protocols, assay specificity, adipose depot, and measurement timing influence whether such changes are detected. This framework may help distinguish true non-response from design related failure to detect an existing effect and improve cross study synthesis in obese older adults.

## Nutritional factors and thermogenic adipose tissue

Direct evidence for nutrition-driven activation of thermogenic adipose tissue in obese older adults is scarce; therefore, this section emphasizes mechanistic plausibility and translational relevance, while distinguishing these from confirmed intervention effects in older populations. Dietary patterns and nutrient exposures can influence energy expenditure and adipose tissue biology. Beyond total energy intake, selected foods and bioactive constituents may augment thermogenic responses by engaging pathways related to brown adipose tissue activity or by facilitating browning within white adipose depots. Because nutritional strategies can be implemented alongside exercise and are generally more sustainable over extended periods, this section frames them as adjunct modules that support thermogenic phenotypes rather than as primary substitutes for cold exposure or pharmacological stimulation.

The following sections synthesize principal categories of thermogenesis supportive nutrients, outline their putative mechanisms, and integrate findings from experimental models and human studies, with a conceptual overview provided in [Figure 4](#).

Rather than discussing each compound individually, we group candidate nutrients according to their predominant mechanism of action in thermogenic adipose tissue. Four broad categories can be identified: (i) activators of sympathetic and TRP channel signaling that engage the upstream neural input to BAT; (ii) modulators of mitochondrial biogenesis and UCP1 transcription acting through the AMPK-SIRT1-PGC-1 $\alpha$  axis; (iii) lipid-derived mediators and substrates that influence inter-organ fuel handling; and (iv) nuclear receptor ligands and enzymatic cofactors that establish a permissive metabolic baseline. This framework helps clarify which compounds act on the same node and are therefore unlikely to produce additive effects, as well as which act upstream or downstream, a distinction that is particularly relevant in obese older adults, in whom upstream sympathetic drive and mitochondrial reserve are already compromised. [Table 1](#) outlines the corresponding preclinical and clinical evidence for representative compounds in each category, together with the recommendations for obese older adults.

## Activators of sympathetic and TRP channel signaling

Caffeine is common in coffee, tea, and many commercial drinks, and it blocks adenosine receptors while also increasing sympathetic signaling ([169](#), [170](#)). Short term studies usually show small rises in resting energy use and modest increases in supraclavicular skin temperature ([171](#)). Some experimental work also suggests that regular intake, especially when paired with tea catechins, may relate to higher cold induced thermogenic readouts, even though study designs and outcome measures differ widely across reports ([172](#)). Older adults still need careful safety checks, since caffeine can disrupt sleep, worsen anxiety symptoms, and raise blood pressure.

Capsaicin is the pungent compound in chili peppers, and it activates transient receptor potential vanilloid 1 (TRPV1) channels while also engaging sympathetic related signaling ([173](#), [174](#)). Human studies often report small increases in energy use or modest rises in cold related thermogenic readouts after capsaicin exposure, which suggests a possible effect, although the overall size is usually limited ([175](#)). A recent systematic review and meta-analysis of randomized trials in overweight or obese adults reported only modest average effects on BMI, body weight, and waist circumference, and the trials varied widely in design and dosing ([176](#)). Other pungent and aromatic compounds, such as cinnamaldehyde and gingerol, have been linked in some short term studies to small shifts in energy use or substrate oxidation ([177](#), [178](#)).

## Modulators of mitochondrial biogenesis and UCP1 transcription

Polyphenols are abundant in plant based foods, including fruits, vegetables, tea, and wine, and are commonly linked to antioxidant

and anti-inflammatory properties ([179](#)). Emerging evidence indicates that selected polyphenols and other phytochemicals may support BAT related phenotypes or facilitate white adipose browning indirectly by modulating mitochondrial adaptation, sympathetic signaling pathways, or adipocyte transcriptional programs ([180](#)).

Resveratrol, a polyphenol abundant in grapes and red wine, is among the most intensively investigated dietary bioactives. In multiple rodent studies, resveratrol supplementation has been associated with increased adipose oxidative capacity and upregulation of UCP1 related thermogenic pathways ([181](#)). Enhanced expression of thermogenesis associated genes has also been reported in neonatal or early life intervention paradigms ([182](#)); however, these developmental models may not translate directly to obesity in later life and should be interpreted cautiously.

In adult obese mice, resveratrol administration has been linked to higher oxygen consumption, increased UCP1 expression in brown adipose tissue, and improvements in glucose metabolic phenotypes, with similar observations reported in models of more severe metabolic disruption ([183](#), [184](#)). Mechanistically, resveratrol is proposed to engage SIRT1 and AMPK signaling, thereby promoting PGC 1 $\alpha$  dependent mitochondrial biogenesis and oxidative metabolism programs that align with UCP1 related thermogenic phenotypic shifts ([185](#), [186](#)).

In humans, replication has been variable across trials, likely reflecting differences in dosing, bioavailability, baseline phenotype, and endpoints. In some trials involving obese men, prolonged high dose resveratrol supplementation increased expression of thermogenic markers in subcutaneous adipose tissue, including UCP1, PRDM16, and PGC 1 $\alpha$ , together with changes consistent with SIRT1 pathway engagement ([187](#)). In contrast, other studies have not detected meaningful reductions in body weight or clear improvements in metabolic outcomes.

Resveratrol has also been proposed to modulate energy balance through effects on the gut microbiome and bile acid signaling ([188](#)), although direct supportive evidence in humans remains relatively limited. In older adults, key constraints relate primarily to limited bioavailability and long term tolerability. Accordingly, rather than advocating routine supplementation, a more informative research direction is development of formulations or structural analogs with improved systemic exposure, followed by evaluation using reproducible mechanistic and clinical endpoints in metabolically compromised elderly populations.

Tea catechins, particularly epigallocatechin gallate (EGCG) from green tea, are also widely studied polyphenols. Evidence from intervention studies suggests that catechin rich preparations may increase fat oxidation and produce small increases in energy expenditure, which has motivated their evaluation in weight management ([189](#)). In some reports, co administration of EGCG with caffeine has been associated with greater cold related thermogenic readouts, consistent with possible interaction at the level of sympathetic signaling ([190](#), [191](#)), although effect sizes are heterogeneous across studies.

Curcumin, the principal polyphenolic constituent of turmeric, has been investigated for potential links to thermogenic phenotypes. In animal studies, curcumin supplementation has been associated with higher UCP1 expression and increased browning markers in white adipose tissue, accompanied by reduced adiposity and improved insulin sensitivity ([192](#), [193](#)). Proposed

mechanisms include activation of AMPK related signaling and dampening of inflammatory pathways.

From a clinical perspective, low bioavailability remains a major limitation, and the evidence that curcumin directly improves human BAT function or produces sustained increases in thermogenic output remains preliminary. Some small clinical trials report added improvements in body weight or glycemic measures (194, 195), yet studies with direct and repeatable human endpoints are still needed to confirm any thermogenic benefit. New formulations that improve solubility, or the use of absorption enhancers taken together with curcumin, may raise systemic exposure, but stronger evidence is needed to define their real clinical value (196).

## Lipid-derived signaling and substrate supply

Dietary fat composition can influence substrate handling and signaling within thermogenic adipose depots. Relative to saturated fatty acids, monounsaturated and polyunsaturated fatty acids are more often examined for associations with brown and beige adipose phenotypes (197). In practice, improving fat quality is more consistently linked to favorable cardiometabolic profiles, including better lipid parameters, reduced inflammatory tone, and enhanced insulin sensitivity, and may therefore provide a metabolic foundation that supports thermogenic responsiveness (198).

Long chain omega 3 polyunsaturated fatty acids, particularly eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) from fish oil, are the most frequently discussed examples. In animal models, omega 3 supplementation is often accompanied by increased expression of UCP1 related markers in brown adipose tissue or subcutaneous white adipose tissue and by upregulation of mitochondrial oxidative pathways (199). Mechanistic studies further propose that omega 3 fatty acids can modulate adipocyte signaling through fatty acid receptor mediated pathways and, in some contexts, influence sympathetic input to adipose tissue, with transient receptor potential channel related mechanisms also suggested (200, 201).

Across population level studies, the most consistent effects of omega 3 supplementation are observed in cardiometabolic biomarkers, including reductions in triglycerides and changes in inflammation related measures. Some reports further suggest that fish oil may increase postprandial energy expenditure or related metabolic readouts, potentially reflecting enhanced substrate mobilization (202), although findings are not uniform.

Beyond omega 3 fatty acids, oleic acid is also of interest. Oleic acid is a monounsaturated fat found in high amounts in olive oil. Researchers often interpret these findings as a shift toward more oxidative metabolic programming in fat cells when diets high in oleic acid replace diets high in saturated fat (203). Proposed mechanisms include changes in lipid handling through nuclear receptor driven gene activity, together with more efficient  $\beta$  oxidation that may provide fuel for uncoupling linked energy dissipation.

Omega 3 fatty acids have stronger evidence for lowering triglycerides (204). From an inter organ communication perspective, thermogenic adipose tissue both generates and responds to lipid derived mediators. A prominent example is 12,13

diHOME, an oxidized linoleic acid metabolite that has been linked to brown adipose activation during cold exposure. Experimental and translational findings suggest that 12,13 diHOME may enhance fatty acid uptake by skeletal muscle, thereby coordinating substrate utilization between brown adipose tissue and muscle (205).

Interindividual variation in circulating 12,13-diHOME appears substantial across human studies, and observed associations may depend on baseline metabolic state and study design, including temperature exposure and sampling conditions. In this context, dietary linoleic acid is best interpreted as a background substrate rather than a direct lever for increasing 12,13-diHOME in a predictable manner. A more appropriate approach is to maintain a balanced intake of essential fatty acids and to avoid treating single fatty acids or their metabolites as actionable standalone intervention targets.

## Nuclear receptor-mediated transcriptional programming: Vitamins A and D

Vitamins A and D exert their effects through the ligand-activated nuclear receptors RAR and VDR, which are involved in adipocyte differentiation and the transcriptional regulation of oxidative and thermogenic programs. Because the strength of these signals depends on receptor occupancy rather than simply on nutrient intake, supraphysiological exposure does not consistently translate into greater thermogenic effects. In clinical practice, the most meaningful application of both vitamins remains the correction of deficiency.

Vitamin A and its metabolites, particularly retinoic acid, are closely linked to adipocyte differentiation, adipose tissue metabolism, and thermogenic programming, as supported by mechanistic studies in mice and analyses of human adipose tissue retinoic acid biosynthesis (206, 207). Experimental work shows that retinoic acid signaling can change oxidative metabolism and thermogenesis related gene programs under certain conditions, as seen in mouse studies where adipocyte retinoic acid receptor  $\alpha$  signaling increased energy use and reduced metabolic injury (208). These effects depend strongly on dose and setting, making it hard to translate them into lasting clinical benefit in obese older adults, even though a recent mouse study reported that brown fat-specific overexpression of retinol-binding protein 4 (RBP4) can increase cold-induced thermogenesis (209). In addition, preclinical studies have shown that retinoid-based interventions may reduce adiposity and improve obesity-related metabolic phenotypes in mice; for example, all-trans retinoic acid administration has been reported to reduce fat accumulation, and more recent work further showed reduced body weight and white adipose tissue accumulation in obese mice (210, 211).

In clinical practice, vitamin A supplements are best used to correct deficiency. Treatment of low intake or malabsorption can help maintain normal metabolism and tissue health. Many older adults already have enough liver stores, so high dose supplementation without a clear need may raise the risk of adverse events, including hepatotoxicity. A safer approach prioritizes dietary sources of provitamin A carotenoids, such as beta carotene rich vegetables and tubers, and reserves supplements for confirmed deficiency after professional assessment and follow up monitoring.

Vitamin D signaling connects with pathways involved in energy metabolism, and vitamin D insufficiency is common in obese older adults, which has led to interest in possible links with thermogenic adipose phenotypes. Vitamin D receptor signaling has been implicated in adipocyte differentiation, lipogenesis, and inflammatory responses (212), and mechanistic work has further shown that vitamin D can regulate fatty acid composition in subcutaneous adipose tissue through Elov3 (213). The evidence base still comes mainly from mechanistic work and animal models; human studies vary in design, dosing, and endpoint selection (Box 1). Recent human work also reported no clear association between circulating 25-hydroxyvitamin D and BAT metabolism in healthy adults (214).

Clinicians generally view vitamin D as a basic part of nutritional care in older adults. Sunlight exposure, diet, and supplements can correct deficiency and support skeletal and immune health, and these effects may help metabolic flexibility in a broader way. Evidence for thermogenesis remains limited, and current human data do not consistently show that routine vitamin D supplementation at standard doses produces lasting increases in BAT thermogenic function in adults. At the same time, disease-context-specific animal data suggest that vitamin D repletion can attenuate aberrant adipose browning and thermogenic gene expression, as shown in a mouse model of CKD-associated cachexia (215). Animal studies still report depot specific and context dependent thermogenic changes when vitamin D status is altered in rats and mice (216, 217). Vitamin D should therefore not be framed as a main thermogenic intervention.

In older adults, supplementation should focus on correcting deficiency and should match the individual clinical setting, including renal function, prior hypercalcemia, nephrolithiasis risk, and current medications. Clinicians can monitor serum calcium and 25-hydroxyvitamin D when clinical circumstances make it necessary.

## Micronutrient cofactors for mitochondrial and oxidative metabolism: B vitamins, choline, iron, and zinc

B vitamins, choline, iron, and zinc all serve as essential cofactors in one-carbon metabolism, the electron transport chain, and redox enzymes that support the oxidative capacity required for UCP1-mediated thermogenesis. Their role is therefore better understood as permissive rather than stimulatory. Adequate levels help avoid a metabolic ceiling, whereas supraphysiological dosing does not appear to provide additional thermogenic drive and may instead introduce off-target risks, including zinc-induced copper deficiency and iron overload-related dysmetabolism. In obese older adults, the more appropriate clinical focus is to identify and correct deficiency rather than to use supplementation as a means of enhancing thermogenesis.

B vitamins and choline play key roles in cellular energy use and one carbon metabolism, and they act as cofactors that shape substrate handling and mitochondrial processes. *In vitro* studies suggest that some components can influence beige related features under specific experimental settings and within narrow dose ranges (218), and recent work supports this by showing that thiamine availability can affect thermogenic activation in

human adipocytes (219). Beyond adipocyte experiments, thiamine deficiency has also been reported in individuals with obesity and in bariatric surgery candidates, including preoperative cohorts and medically complicated obesity populations (220–223). These findings support careful perioperative assessment and correction of thiamine deficiency in the bariatric setting rather than assuming adequate thiamine status. Recent human adipose studies further indicate that SLC19A3, which encodes thiamine transporter 2, shows adipose-enriched expression, increases during adipocyte differentiation, and is detectable in both brown and white adipose depots, strengthening the relevance of thiamine handling for adipocyte function (224). These findings still depend heavily on experimental context, so they do not support high dose supplementation as a way to boost thermogenesis.

In older adults, clinical care should focus on finding and correcting nutritional deficiencies. Low plasma thiamine concentrations have been reported in type 1 and type 2 diabetes (225), and high-dose thiamine supplementation has been reported to improve glucose tolerance in hyperglycemic individuals (226). In addition, case reports of Wernicke's encephalopathy have described severe hypothermia responding to thiamine treatment (227), underscoring the systemic consequences of marked deficiency. Vitamin B<sub>12</sub> insufficiency and malabsorption are relatively common, and many people may also have low habitual choline intake. Balanced dietary patterns, or a suitable multivitamin chosen through clinical assessment, can help close these gaps and support overall metabolic health. People with multiple comorbidities and polypharmacy also need individualized safety planning. Periodic assessment of vitamin B<sub>12</sub> status may be appropriate in long term users of metformin or proton pump inhibitors.

Iron is required for many parts of the mitochondrial electron transport chain. Iron supports energy metabolism by enabling ATP production and normal electron transport chain function (228). Disrupted iron homeostasis, marked by high serum ferritin and excess iron build up in the liver, adipose tissue, and skeletal muscle, can worsen adipose dysfunction (229, 230). Mitochondrial biogenesis and function also rely on iron. Experimental studies show that excess iron in adipose tissue during browning, induced by  $\beta$ 3 AR activation, is linked to higher mitochondrial formation and respiration (231), and recent mouse work also suggests that iron supplementation can promote adipocyte thermogenesis through lipolysis related pathways (232). Mouse studies also show that altering adipose iron handling can raise mitochondrial respiration and adipokine expression in BAT (233), and newer ferritin heavy chain knockout work has reported improved adaptive thermogenesis and better metabolic profiles (234). Iron deficiency can impair beige fat formation, while iron chelation can enhance beige fat differentiation and metabolic activity.

Human evidence should, however, be interpreted with caution. In human adipose tissue, TFRC expression has been reported to increase in browning fat but to decrease in adipose tissue from overweight individuals. TFRC was also negatively correlated with body mass index and positively correlated with UCP1, suggesting that reduced TFRC may be associated with an impaired thermogenic phenotype in obesity rather than higher TFRC levels in overweight individuals *per se* (235). More recent work further showed that adrenergic stimulation increased TFRC expression in human brown adipocytes and that transferrin was constitutively secreted by these cells, supporting a role for iron handling during

thermogenic activation (236). Together, these findings support the biological plausibility of iron related regulation in beige adipocyte development, with phenotype specific effects likely across aging and obesity.

Zinc is an essential trace element that supports many enzymes and is closely tied to immune function and nervous system stability. Zinc status may shape metabolic adaptation in an indirect way because it supports tissue repair and the broader neuroimmune environment. In older adults, supplementation should mainly target deficiency correction, and large scale clinical data still show that zinc deficiency becomes more common with age (237, 238). Mild insufficiency is relatively common and can often be improved through foods such as nuts, legumes, and seafood. Supplements should be reserved for confirmed deficiency and used to restore levels back to the normal physiological range, since animal studies also suggest that zinc exposure can affect adipose lipid handling and inflammation in ways that may not always be beneficial. Excess zinc can also reduce copper absorption and cause secondary copper deficiency, so dosing should aim for adequacy rather than high exposure, and recent clinical reports continue to describe zinc induced copper deficiency in older adults (239).

Nutritional studies relevant to thermogenic adipose responses often vary in dosing, formulation, intervention duration, and concurrent lifestyle context, which complicates attribution to single nutrients. Future work should focus on pragmatic dietary patterns, combine reproducible biomarkers with interpretable thermogenic endpoints, and evaluate feasibility and safety in older adults with common comorbidities and medication exposure.

## Synergistic interventions and clinical implications

For obese older adults, combined endurance and resistance training should form the foundation of intervention, as skeletal muscle contraction promotes exercise-responsive mediators, including irisin, IL-6, FGF21, and METRN1, while reducing adverse myostatin signaling, thereby helping shape the endocrine and paracrine milieu that supports browning signals and BAT responsiveness. We suggest moderate-intensity aerobic training at 40% to 60% VO<sub>2</sub> reserve, or an RPE of 12–14, for 150–300 min per week, together with progressive resistance training 2–3 times per week involving major muscle groups and 8–12 repetitions per set. This combination addresses both metabolic flexibility and preservation of muscle mass. Exercise prescriptions should be individualized according to comorbidity profile. Low-impact modalities such as aquatic exercise or recumbent cycling may be more suitable for individuals with osteoarthritis or balance impairment. In those with cardiovascular disease, intensity should be advanced cautiously after appropriate pre-participation screening. Supervised initiation may be particularly appropriate in the setting of frailty, recent hospitalization, or insulin-treated diabetes. Adherence may be improved by using shorter, more frequent sessions and home-based options, given the mobility and transportation barriers commonly faced by this population. [Figure 5](#) summarizes this integrated framework, including the principal exercise- and nutrition-related inputs and the barriers most likely to constrain net effect.

Dietary optimization can complement these effects by influencing both substrate availability and pathway-specific signaling. Adequate protein intake, at 1.0–1.2 g/kg/day and increasing to 1.2–1.5 g/kg/day when combined with resistance training, distributed across meals at about 25–30 g per meal, helps support muscle protein synthesis and may counteract sarcopenia-related reductions in exerkine production. A Mediterranean-style dietary pattern provides omega-3 PUFAs, with at least 250 mg/day of EPA plus DHA as a general nutritional target rather than a BAT-specific dose; these lipids may support cardiometabolic health and lipid-mediated signaling relevant to thermogenic tone, while MUFAs offer additional substrate and signaling support. Adjunctive thermogenic nutrition strategies may also include moderate caffeine or tea catechin intake to support sympathetic BAT activation, culinary spices such as capsaicin, cinnamaldehyde, and ginger that engage TRPV1- or TRPA1-related browning pathways, and polyphenols including resveratrol and curcumin, which may promote mitochondrial biogenesis and UCP1-permissive signaling through AMPK and SIRT1. Human effect sizes remain modest and appear to depend on context. Caution is needed in the setting of chronic kidney disease, where protein intake may require adjustment, as well as in patients using anticoagulants, those with arrhythmia or uncontrolled hypertension, and those receiving multiple drugs with narrow therapeutic indices.

Clinical benefit ultimately depends on whether three principal barriers can be at least partly addressed: sarcopenia, which reduces exerkine production; inflammaging, in which elevated TNF- $\alpha$  and IL-1 $\beta$  impair mitochondrial function and suppress UCP1; and  $\beta$ -adrenergic resistance, which limits sympathetic signaling to BAT. In selected patients, lifestyle intervention may be combined with pharmacotherapy. GLP-1 receptor agonists can support clinically meaningful weight loss, but concurrent resistance training and adequate protein intake are needed to reduce accompanying lean-mass loss, whereas  $\beta$ 3-adrenergic agonists remain investigational. Variation in response is likely related to baseline metabolic phenotype, sex, body composition, medication burden, and inflammaging status. Biomarker-guided stratification using circulating mediators such as 12,13-diHOME, FGF21, and irisin, together with functional and metabolic endpoints rather than weight or imaging alone, may help identify those most likely to respond. When these barriers are at least partly overcome, the overall effects may include greater BAT and beige activation, improved glucose, lipid, and blood pressure profiles, and clinically meaningful benefit even in the absence of substantial weight loss.

## Conclusion

Late-life obesity is accompanied by an age-related decline in the recruitable capacity of brown and beige adipose tissue, arising from impaired sympathetic signaling, chronic low-grade inflammation, endocrine remodeling, and weaker muscle-derived cues. Combined exercise and nutritional strategies remain the most defensible basis for clinical management, and their metabolic and functional benefits, including improved glucose disposal, lipid clearance, and lower cardiometabolic risk, may still be clinically meaningful even when weight loss is modest. Intervention assessment should

therefore give priority to metabolic and functional outcomes rather than weight change alone.

Several important gaps remain. First, adequately powered trials in obese adults aged 65 years and older are still scarce, and much of the current evidence is extrapolated from younger or non-obese populations. Second, standardized definitions of thermogenic adipose tissue activation that integrate histological, transcriptomic, and imaging-based criteria are still lacking, which continues to contribute to inconsistent findings. Third, predictors of response and tools for stratification remain underdeveloped, and key questions about who benefits, to what extent, and through which mechanisms have yet to be answered. Fourth, the relative contributions of UCP1-dependent and UCP1-independent thermogenic pathways in older adults, their interaction with sarcopenic obesity, inflammaging, and polypharmacy, and the durability of induced beige adipocyte phenotypes after stimulus withdrawal require further mechanistic and translational investigation.

Future work should therefore focus on four priorities: randomized trials in obese older adults with prespecified subgroup analyses by sex, frailty status, and comorbidity profile; harmonized outcome frameworks linking circulating biomarkers such as 12,13-diHOME, FGF21, and irisin to clinically meaningful endpoints; head-to-head comparisons of exercise and nutrition combinations versus pharmacological adjuncts such as GLP-1 receptor agonists; and safety and adherence data extending to at least 12 months in high-risk subgroups. Until these gaps are addressed, brown and beige adipose tissue should be viewed as a promising but still provisional therapeutic lens in late-life obesity.

## Author contributions

FW: Conceptualization, Formal analysis, Visualization, Writing – original draft. HQ: Investigation, Methodology, Writing – review & editing. HZ: Formal analysis, Investigation, Writing – original draft. YZ: Investigation, Supervision, Writing – original draft. YN: Validation, Writing – review & editing. XH: Project administration, Supervision, Writing – review & editing.

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## References

- Freitas HR, Malva JO. Getting ahead of the ageing curve: learning from EU experiences for a healthier demographic transition. *Front Public Health*. (2025) 13:1678040. doi: 10.3389/fpubh.2025.1678040
- Ullrich-Kniffka N, Schöley J. Population age structure dependency of the excess mortality P-score. *Popul Health Metr*. (2024) 22:25. doi: 10.1186/s12963-024-00346-w
- Henney AE, Wilding JPH, Alam U, Cuthbertson DJ. Obesity pharmacotherapy in older adults: a narrative review of evidence. *Int J Obes*. (2025) 49:369–80. doi: 10.1038/s41366-024-01529-z
- Kerr AG, Andersson DP, Dahlman I, Rydén M, Arner P. Adipose insulin resistance associates with dyslipidemia independent of liver resistance and involves early

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## Conflict of interest

The author(s) declared that this work was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fnut.2026.1818342/full#supplementary-material>

- hormone signaling. *Arterioscler Thromb Vasc Biol.* (2023) 43:1054–65. doi: 10.1161/atvbaha.123.319227
5. Wei S, Nguyen TT, Zhang Y, Ryu D, Gariani K. Sarcopenic obesity: epidemiology, pathophysiology, cardiovascular disease, mortality, and management. *Front Endocrinol.* (2023) 14:1185221. doi: 10.3389/fendo.2023.1185221
6. Liu J, Huang Q, Liu F. Fat talks first: How adipose tissue sets the pace of aging? *Life Med.* (2025) 4:lnaf028. doi: 10.1093/lifemedi/lnaf028
7. Malicka A, Ali A, MacCannell ADV, Roberts LD. Brown and beige adipose tissue-derived metabolite and lipokine inter-organ signalling in health and disease. *Exp Physiol.* (2025) 110:918–35. doi: 10.1113/ep092008
8. Li J, Wang L, Wu S, Yang G, Zhang LJ. Imaging brown adipose tissue: current state and future perspective. *Theranostics.* (2025) 15:9001–18. doi: 10.7150/thno.111643
9. Virtanen KA, Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, et al. Functional brown adipose tissue in healthy adults. *N Engl J Med.* (2009) 360:1518–25. doi: 10.1056/NEJMoa0808949
10. Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB, et al. Identification and importance of brown adipose tissue in adult humans. *N Engl J Med.* (2009) 360:1509–17. doi: 10.1056/NEJMoa0810780
11. Tang S, Geng Y, Lin Q. Adipose tissue aging as a risk factor for metabolic organ abnormalities: mechanistic insights and the role of exercise interventions. *Lipids Health Dis.* (2025) 24:274. doi: 10.1186/s12944-025-02695-3
12. Dobre MZ, Virgolic B, Timnea O. Key roles of brown, subcutaneous, and visceral adipose tissues in obesity and insulin resistance. *Curr Issues Mol Biol.* (2025) 47:343. doi: 10.3390/cimb47050343
13. Luo Y, Song Y, Zeng M, Li B, Li Y, Dong Z. The past, present, and future of adipose tissue browning and aging: a review combined with bibliometrics and bioinformatics of 2527 documents published over the past four decades. *Ageing Res Rev.* (2026) 114:102959. doi: 10.1016/j.arr.2025.102959
14. Cypess AM, Cannon B, Nedergaard J, Kazak L, Chang DC, Krakoff J, et al. Emerging debates and resolutions in brown adipose tissue research. *Cell Metab.* (2025) 37:12–33. doi: 10.1016/j.cmet.2024.11.002
15. Alves JM. Two roads to thermogenesis: vascular and parenchymal sympathetic innervation of brown adipose tissue. *Nat Rev Endocrinol.* (2025) 22:198. doi: 10.1038/s41574-025-01224-7
16. Haley JA, Jang C, Guertin DA. A new era of understanding in vivo metabolic flux in thermogenic adipocytes. *Curr Opin Genet Dev.* (2023) 83:102112. doi: 10.1016/j.gde.2023.102112
17. Negriou CE, Tudora?cu I, Bezna CM, Godeanu S, Diaconu M, Danoiu R, et al. Beyond the cold: activating brown adipose tissue as an approach to combat obesity. *J Clin Med.* (2024) 13:1973. doi: 10.3390/jcm13071973
18. Yoneshiro T, Matsushita M, Sakai J, Saito M. Brown fat thermogenesis and cold adaptation in humans. *J Physiol Anthropol.* (2025) 44:11. doi: 10.1186/s40101-025-00391-w
19. Bombassaro B, Batitucci G, Reymond Simoes M, Araujo EP, Velloso LA. The impact of dietary factors on the function of brown and beige adipose tissues—implications on health and disease. *Front Nutr.* (2025) 12:1626068. doi: 10.3389/fnut.2025.1626068
20. Fedorenko A, Lishko PV, Kirichok Y. Mechanism of fatty-acid-dependent UCP1 uncoupling in brown fat mitochondria. *Cell.* (2012) 151:400–13. doi: 10.1016/j.cell.2012.09.010
21. Ragni M, Ruocco C, Nisoli E. Mitochondrial uncoupling, energy substrate utilization, and brown adipose tissue as therapeutic targets in cancer. *NPJ Metab Health Dis.* (2025) 3:37. doi: 10.1038/s44324-025-00080-3
22. Pfannenbergs C, Werner MK, Ripkens S, Stef I, Deckert A, Schmadl M, et al. Impact of age on the relationships of brown adipose tissue with sex and adiposity in humans. *Diabetes.* (2010) 59:1789–93. doi: 10.2337/db10-0004
23. Yoneshiro T, Aita S, Matsushita M, Okamatsu-Ogura Y, Kameya T, Kawai Y, et al. Age-related decrease in cold-activated brown adipose tissue and accumulation of body fat in healthy humans. *Obesity.* (2011) 19:1755–60. doi: 10.1038/oby.2011.125
24. Kindred JH, Tuulari JJ, Simon S, Luckasen GJ, Bell C, Rudroff T. Brown adipose and central nervous system glucose uptake is lower during cold exposure in older compared to young men: a preliminary PET study. *Ageing Clin Exp Res.* (2016) 28:557–60. doi: 10.1007/s40520-015-0521-2
25. Zoico E, Rubele S, De Caro A, Nori N, Mazzali G, Fantin F, et al. Brown and beige adipose tissue and aging. *Front Endocrinol.* (2019) 10:368. doi: 10.3389/fendo.2019.00368
26. Ma L, Xiong L, Huang G. Effects of mirabegron on brown adipose tissue and metabolism in humans: a systematic review and meta-analysis. *Eur J Clin Pharmacol.* (2024) 80:317–33. doi: 10.1007/s00228-023-03614-0
27. Pasha A, Tondo A, Favre C, Calvani M. Inside the biology of the  $\beta$ 3-adrenoceptor. *Biomolecules.* (2024) 14:159. doi: 10.3390/biom14020159
28. Cypess AM, Weiner LS, Roberts-Toler C, Franquet Elía E, Kessler SH, Kahn PA, et al. Activation of human brown adipose tissue by a  $\beta$ 3-adrenergic receptor agonist. *Cell Metab.* (2015) 21:33–8. doi: 10.1016/j.cmet.2014.12.009
29. Carpentier AC, Blondin DP. Human brown adipose tissue is not enough to combat cardiometabolic diseases. *J Clin Invest.* (2023) 133:e175288. doi: 10.1172/jci175288
30. Balligand JL, Brito D, Brosteau O, Casadei B, Depoix C, Edelmann F, et al. Repurposing the  $\beta$ 3-adrenergic receptor antagonist mirabegron in patients with structural cardiac disease: the Beta3-LVH Phase 2b randomized clinical trial. *JAMA Cardiol.* (2023) 8:1031–40. doi: 10.1001/jamacardio.2023.3003
31. Dumont L, Caron A, Richard G, Croteau E, Fortin M, Frisch F, et al. The effects of the  $\beta$ (1)-adrenergic receptor antagonist bisoprolol administration on mirabegron-stimulated human brown adipose tissue thermogenesis. *Acta Physiol.* (2024) 240:e14127. doi: 10.1111/apha.14127
32. Gorini Pereira F, Ryan CT, Miller S, Watters A, Mickleborough TD, Schlader ZJ, et al. The thermogenic effect of mirabegron ingestion during cool conditions. *Front Physiol.* (2025) 16:1645475. doi: 10.3389/fphys.2025.1645475
33. Kajimura S. An unexpected journey into brown fat research for metabolic health: the 2025 outstanding scientific achievement award lecture. *Diabetes.* (2025) 74:2216–22. doi: 10.2337/dbi25-0026
34. Bunk J, Hussain MF, Delgado-Martin M, Samborska B, Ersin M, Shaw A, et al. The futile creatine cycle powers UCP1-independent thermogenesis in classical BAT. *Nat Commun.* (2025) 16:3221. doi: 10.1038/s41467-025-58294-4
35. Rahbani JF, Bunk J, Lagarde D, Samborska B, Roesler A, Xiao H, et al. Parallel control of cold-triggered adipocyte thermogenesis by UCP1 and CKB. *Cell Metab.* (2024) 36:526–40.e7. doi: 10.1016/j.cmet.2024.01.001
36. Rahbani JF, Roesler A, Hussain MF, Samborska B, Dykstra CB, Tsai L, et al. Creatine kinase B controls futile creatine cycling in thermogenic fat. *Nature.* (2021) 590:480–5. doi: 10.1038/s41586-021-03221-y
37. Sun Y, Rahbani JF, Jedrychowski MP, Riley CL, Vidoni S, Bogoslavski D, et al. Mitochondrial TNAP controls thermogenesis by hydrolysis of phosphocreatine. *Nature.* (2021) 593:580–5. doi: 10.1038/s41586-021-03533-z
38. Su Y. Three-dimensional network of creatine metabolism: from intracellular energy shuttle to systemic metabolic regulatory switch. *Mol Metab.* (2025) 100:102228. doi: 10.1016/j.molmet.2025.102228
39. Poursharifi P, Madiraju SRM, Oppong A, Kajimura S, Nolan CJ, Blondin DP, et al. Glycerolipid cycling in thermogenesis, energy homeostasis, signaling, and diseases. *Physiol Rev.* (2025) 105:2449–99. doi: 10.1152/physrev.00024.2024
40. Sharma AK, Khandelwal R, Wolfrum C. Futile cycles: emerging utility from apparent futility. *Cell Metab.* (2024) 36:1184–203. doi: 10.1016/j.cmet.2024.03.008
41. Sharma AK, Khandelwal R, Wolfrum C. Futile lipid cycling: from biochemistry to physiology. *Nat Metab.* (2024) 6:808–24. doi: 10.1038/s42255-024-01003-0
42. Simoes MR, Gallo-Ferraz AL, Bombassaro B, Valdivieso-Rivera F, Nogueira GAS, Monfort-Pires M, et al. Bidirectional shifts in Pm20d1 expression impact thermogenesis and metabolism. *Mol Med.* (2025) 31:283. doi: 10.1186/s10020-025-01345-9
43. Ikeda K, Kang Q, Yoneshiro T, Camporez JP, Maki H, Homma M, et al. UCP1-independent signaling involving SERCA2b-mediated calcium cycling regulates beige fat thermogenesis and systemic glucose homeostasis. *Nat Med.* (2017) 23:1454–65. doi: 10.1038/nm.4429
44. Auger C, Li M, Fujimoto M, Ikeda K, Yook JS, O’Leary TR, et al. Identification of a molecular resistor that controls UCP1-independent Ca(2+) cycling thermogenesis in adipose tissue. *Cell Metab.* (2025) 37:1311–25.e9. doi: 10.1016/j.cmet.2025.03.009
45. Liu X, Zhang Z, Song Y, Xie H, Dong M. An update on brown adipose tissue and obesity intervention: function, regulation and therapeutic implications. *Front Endocrinol.* (2022) 13:1065263. doi: 10.3389/fendo.2022.1065263
46. Finlin BS, Memetimin H, Confides AL, Zhu B, Westgate PM, Dupont-Versteegden EE, et al. Macrophages expressing uncoupling protein 1 increase in adipose tissue in response to cold in humans. *Sci Rep.* (2021) 11:23598. doi: 10.1038/s41598-021-03014-3
47. Wu S, Qiu C, Ni J, Guo W, Song J, Yang X, et al. M2 macrophages independently promote beige adipogenesis via blocking adipocyte Ets1. *Nat Commun.* (2024) 15:1646. doi: 10.1038/s41467-024-45899-4
48. Sakers A, De Siqueira MK, Seale P, Villanueva CJ. Adipose-tissue plasticity in health and disease. *Cell.* (2022) 185:419–46. doi: 10.1016/j.cell.2021.12.016
49. Yang Y, Zhang G, Yi T, Yang S, Wu S, Zhang Y, et al. snRNA-seq reveals subcutaneous white adipose tissue remodeling upon return to thermoneutrality after cold stimulation. *Front Cell Dev Biol.* (2025) 13:1578180. doi: 10.3389/fcell.2025.1578180
50. Shamsi F, Wang CH, Tseng YH. The evolving view of thermogenic adipocytes - ontogeny, niche and function. *Nat Rev Endocrinol.* (2021) 17:726–44. doi: 10.1038/s41574-021-00562-6
51. Wu R, Park J, Qian Y, Shi Z, Hu R, Yuan Y, et al. Genetically prolonged beige fat in male mice confers long-lasting metabolic health. *Nat Commun.* (2023) 14:2731. doi: 10.1038/s41467-023-38471-z
52. Kyung DS, Lee E, Chae S, Son Y, Moon YJ, Hwang D, et al. Single-cell transcriptomic analysis reveals dynamic activation of cellular signaling pathways regulating beige adipogenesis. *Exp Mol Med.* (2024) 56:2309–22. doi: 10.1038/s12276-024-01252-9

53. Carpentier AC, Blondin DP, Haman F, Richard D. Brown Adipose Tissue-A Translational Perspective. *Endocr Rev.* (2023) 44:143–92. doi: 10.1210/edrv/bnac015
54. Jiang S, Bae JH, Wang Y, Song W. The potential roles of myokines in adipose tissue metabolism with exercise and cold exposure. *Int J Mol Sci.* (2022) 23:11523. doi: 10.3390/ijms231911523
55. Zhu H, Liu D, Sui M, Zhou M, Wang B, Qi Q, et al. CRISPRa-based activation of Fgf21 and Fndc5 ameliorates obesity by promoting adipocytes browning. *Clin Transl Med.* (2023) 13:e1326. doi: 10.1002/ctm2.1326
56. Holman CD, Sakers AP, Calhoun RP, Cheng L, Fein EC, Jacobs C, et al. Aging impairs cold-induced beige adipogenesis and adipocyte metabolic reprogramming. *Elife.* (2024) 12:R87756. doi: 10.7554/eLife.87756
57. Zhang H, Li Y, Ibáñez CF, Xie M. Perirenal adipose tissue contains a subpopulation of cold-inducible adipocytes derived from brown-to-white conversion. *Elife.* (2024) 13:R93151. doi: 10.7554/eLife.93151
58. Jespersen NZ, Larsen TJ, Pejts L, Dagaard S, Homøe P, Loft A, et al. A classical brown adipose tissue mRNA signature partly overlaps with white in the supraclavicular region of adult humans. *Cell Metab.* (2013) 17:798–805. doi: 10.1016/j.cmet.2013.04.011
59. Ogawa M, Oshiro H, Tamura Y, Ishido M, Okamoto T, Hata J. Characteristics of T<sup>2</sup>\* and anisotropy parameters in inguinal and epididymal adipose tissues after cold exposure in mice. *Sci Rep.* (2024) 14:29491. doi: 10.1038/s41598-024-78655-1
60. Wang H, Yu L, Wang J, Zhang Y, Xu M, Lv C, et al. SLC35D3 promotes white adipose tissue browning to ameliorate obesity by NOTCH signaling. *Nat Commun.* (2023) 14:7643. doi: 10.1038/s41467-023-4318-5
61. Miranda AMA, McAllan L, Mazzei G, Andrew I, Davies I, Ertugrul M, et al. Selective remodelling of the adipose niche in obesity and weight loss. *Nature.* (2025) 644:769–79. doi: 10.1038/s41586-025-09233-2
62. Kwok TC, Ramage LE, Kelman A, Suchacki KJ, Gray C, Boyle LD, et al. UCP1 expression in human brown adipose tissue is inversely associated with cardiometabolic risk factors. *Eur J Endocrinol.* (2024) 191:106–15. doi: 10.1093/aje/endo/lvae074
63. Cai Z, Zhong Q, Feng Y, Wang Q, Zhang Z, Wei C, et al. Non-invasive mapping of brown adipose tissue activity with magnetic resonance imaging. *Nat Metab.* (2024) 6:1367–79. doi: 10.1038/s42255-024-01082-z
64. Chen N, Yuan M, Zhang N, Chen M, Liu R, Wang J, et al. Ctnnb1/β-catenin inactivation in UCP1-positive adipocytes augments the browning of white adipose tissue. *iScience.* (2023) 26:106552. doi: 10.1016/j.isci.2023.106552
65. Nigro P, Vamvini M, Yang J, Caputo T, Ho LL, Carbone NP, et al. Exercise training remodels inguinal white adipose tissue through adaptations in innervation, vascularization, and the extracellular matrix. *Cell Rep.* (2023) 42:112392. doi: 10.1016/j.celrep.2023.112392
66. Vámos A, Shaw A, Varga K, Csomós I, Mocsár G, Balajthy Z, et al. Mitophagy mediates the beige to white transition of human primary subcutaneous adipocytes ex vivo. *Pharmaceuticals.* (2022) 15:363. doi: 10.3390/ph15030363
67. Benvie AM, Lee D, Steiner BM, Xue S, Jiang Y, Berry DC. Age-dependent Pdgfrβ signaling drives adipocyte progenitor dysfunction to alter the beige adipogenic niche in male mice. *Nat Commun.* (2023) 14:1806. doi: 10.1038/s41467-023-37386-z
68. Tian D, Zeng X, Gong Y, Zheng Y, Zhang J, Wu Z. HDAC1 inhibits beige adipocyte-mediated thermogenesis through histone crotonylation of Pgc1a/Ucp1. *Cell Signal.* (2023) 111:110875. doi: 10.1016/j.cellsig.2023.110875
69. Bódis K, Breuer S, Crepzia-Pevzner A, Zaharia OP, Schön M, Saatmann N, et al. Impact of physical fitness and exercise training on subcutaneous adipose tissue beiging markers in humans with and without diabetes and a high-fat diet-fed mouse model. *Diabetes Obes Metab.* (2024) 26:339–50. doi: 10.1111/dom.15322
70. Toyama Y, Kotani T, Tamaki N, Yamada S, Akiyama S, Nakai Y, et al. Distribution patterns of brown adipose tissue on FDG-PET/CT has age characteristics. *Ann Nucl Med.* (2025) 39:1291–6. doi: 10.1007/s12149-025-02087-8
71. Natarajan D, Plakkot B, Tiwari K, Ekambaram S, Wang W, Rudolph M, et al. Chronic β<sub>3</sub>-AR stimulation activates distinct thermogenic mechanisms in brown and white adipose tissue and improves systemic metabolism in aged mice. *Aging Cell.* (2024) 23:e14321. doi: 10.1111/acel.14321
72. Feng X, Wang L, Zhou R, Zhou R, Chen L, Peng H, et al. Senescent immune cells accumulation promotes brown adipose tissue dysfunction during aging. *Nat Commun.* (2023) 14:3208. doi: 10.1038/s41467-023-38842-6
73. Pellegrinelli V, Figueroa-Juárez E, Samuelson I, Niemi T, Rodriguez-Fdez S, Virtue S, et al. Defective extracellular matrix remodeling in brown adipose tissue is associated with fibro-inflammation and reduced diet-induced thermogenesis. *Cell Rep.* (2023) 42:112640. doi: 10.1016/j.celrep.2023.112640
74. Guo M, Yao J, Li J, Zhang J, Wang D, Zuo H, et al. Irisin ameliorates age-associated sarcopenia and metabolic dysfunction. *J Cachexia Sarcopenia Muscle.* (2023) 14:391–405. doi: 10.1002/jcsm.13141
75. Zhou R, Huang Y, Feng X, Zhou R, Wang L, Xie G, et al. Decreased YB-1 expression denervates brown adipose tissue and contributes to age-related metabolic dysfunction. *Cell Prolif.* (2024) 57:e13520. doi: 10.1111/cpr.13520
76. Sakamoto K, Butera MA, Zhou C, Maurizi G, Chen B, Ling L, et al. Overnutrition causes insulin resistance and metabolic disorder through increased sympathetic nervous system activity. *Cell Metab.* (2025) 37:121–37.e6. doi: 10.1016/j.cmet.2024.09.012.
77. Valentine JM, Ahmadian M, Keinan O, Abu-Odeh M, Zhao P, Zhou X, et al. β<sub>3</sub>-Adrenergic receptor downregulation leads to adipocyte catecholamine resistance in obesity. *J Clin Invest.* (2022) 132:e153357. doi: 10.1172/jci.153357
78. Ahmed F, Vranic M, Hetty S, Mathioudaki A, Patsoukaki V, Fanni G, et al. Increased OCT3 expression in adipose tissue with aging: implications for catecholamine and lipid turnover and insulin resistance in women. *Endocrinology.* (2023) 165:bqad172. doi: 10.1210/endo/bqad172
79. Straat ME, Hoekx CA, van Velden FHP, Pereira Arias-Bouda LM, Dumont L, Blondin DP, et al. Stimulation of the beta-2-adrenergic receptor with salbutamol activates human brown adipose tissue. *Cell Rep Med.* (2023) 4:100942. doi: 10.1016/j.xcrim.2023.100942
80. Ernande L, Stanford KI, Thoonen R, Zhang H, Clerte M, Hirshman MF, et al. Relationship of brown adipose tissue perfusion and function: a study through β<sub>2</sub>-adrenoreceptor stimulation. *J Appl Physiol.* (2016) 120:825–32. doi: 10.1152/jappphysiol.00634.2015
81. Monfort-Pires M, U-Din M, de Mello V, Saari T, Raiko J, Kerminen E, et al. Cold-induced serum short-chain fatty acids act as markers of brown adipose tissue activation in humans. *J Clin Endocrinol Metab.* (2025) 111:1377–88. doi: 10.1210/clinem/dgaf607
82. Gonzalez-Hurtado E, Leveau C, Li K, Mishra M, Qu R, Goldberg EL, et al. Nerve-associated macrophages control adipose homeostasis across lifespan and restrain age-related inflammation. *Nat Aging.* (2025) 5:1828–43. doi: 10.1038/s43587-025-00952-9
83. Carey A, Nguyen K, Kandikonda P, Kruglov V, Bradley C, Dahlquist KJV, et al. Age-associated accumulation of B cells promotes macrophage inflammation and inhibits lipolysis in adipose tissue during sepsis. *Cell Rep.* (2024) 43:113967. doi: 10.1016/j.celrep.2024.113967
84. Yadav MK, Ishida M, Gogoleva N, Liao CW, Salim FN, Kanai M, et al. MAFB in macrophages regulates cold-induced neuronal density in brown adipose tissue. *Cell Rep.* (2024) 43:113978. doi: 10.1016/j.celrep.2024.113978
85. Zhang X, Wang Q, Wang Y, Ma C, Zhao Q, Yin H, et al. Interleukin-6 promotes visceral adipose tissue accumulation during aging via inhibiting fat lipolysis. *Int Immunopharmacol.* (2024) 132:111906. doi: 10.1016/j.intimp.2024.111906
86. Kistner TM, Trinh B, Mfeketo K, van Hall G, Pedersen BK, Lieberman DE, et al. Myokine IL-6 activity enhances post-exercise fatty acid accumulation in skeletal muscle but does not affect glycogen resynthesis. *Mol Metab.* (2026) 103:102283. doi: 10.1016/j.molmet.2025.102283
87. Huang L, Guo Z, Huang M, Zeng X, Huang H. Triiodothyronine (T<sub>3</sub>) promotes browning of white adipose through inhibition of the PI3K/AKT signalling pathway. *Sci Rep.* (2024) 14:20370. doi: 10.1038/s41598-024-71591-0
88. Roth L, Hoffmann A, Hagemann T, Wagner L, Strehlau C, Sheikh B, et al. Thyroid hormones are required for thermogenesis of beige adipocytes induced by Zfp423 inactivation. *Cell Rep.* (2024) 43:114987. doi: 10.1016/j.celrep.2024.114987
89. Blondin DP, Haman F, Swibas TM, Hogan-Lamarre S, Dumont L, Guertin J, et al. Brown adipose tissue metabolism in women is dependent on ovarian status. *Am J Physiol Endocrinol Metab.* (2024) 326:E588–601. doi: 10.1152/ajpendo.00077.2024
90. Brown DE, Shreyer S, Witkowski S, Sievert LL. Brown adipose tissue, adiposity, and menopausal status in women at mid-life. *Am J Biol Anthropol.* (2025) 187:e70073. doi: 10.1002/ajpa.70073
91. Taniguchi H, Hashimoto Y, Dowaki N, Nirengi S. Association of brown adipose tissue activity with circulating sex hormones and fibroblast growth factor 21 in the follicular and luteal phases in young women. *J Physiol Anthropol.* (2024) 43:23. doi: 10.1186/s40101-024-00371-6
92. Stilgenbauer L, de Lima JBM, Debarba LK, Khan M, Koshko L, Kopchick JJ, et al. Growth hormone receptor (GHR) in AgRP neurons regulates thermogenesis in a sex-specific manner. *Geroscience.* (2023) 45:1745–59. doi: 10.1007/s11357-023-00726-4
93. Jurado-Fasoli L, Sanchez-Delgado G, Alcantara JMA, Acosta FM, Sanchez-Sanchez R, Labayen I, et al. Adults with metabolically healthy overweight or obesity present more brown adipose tissue and higher thermogenesis than their metabolically unhealthy counterparts. *EBioMedicine.* (2024) 100:104948. doi: 10.1016/j.ebiom.2023.104948
94. Shi H, Hao X, Sun Y, Zhao Y, Wang Y, Cao X, et al. Exercise-inducible circulating extracellular vesicle irisin promotes browning and the thermogenic program in white adipose tissue. *Acta Physiol.* (2024) 240:e14103. doi: 10.1111/apha.14103
95. Ma Y, Liu Y, Zheng J, Zheng Z, Li J. Fndc5/irisin mediates the benefits of aerobic exercise intervention on aging-associated sarcopenia in mice. *Eur Geriatr Med.* (2025) 16:1081–9. doi: 10.1007/s41999-025-01181-4
96. Jamrasi P, Bae JH, Song W. Effects of 12-week exercise on Meteorin-like levels, inflammation, and functional capacity in older adults: Korean national aging project randomized controlled study. *Eur Geriatr Med.* (2025) 16:2129–40. doi: 10.1007/s41999-025-01272-2
97. Jin L, Han S, Lv X, Li X, Zhang Z, Kuang H, et al. The muscle-enriched myokine Musclin impairs beige fat thermogenesis and systemic energy homeostasis via

- Tfrr1/PKA signaling in male mice. *Nat Commun.* (2023) 14:4257. doi: 10.1038/s41467-023-39710-z
98. Yu L, Wan Q, Liu Q, Fan Y, Zhou Q, Skowronski AA, et al. IgG is an aging factor that drives adipose tissue fibrosis and metabolic decline. *Cell Metab.* (2024) 36:793–807.e5. doi: 10.1016/j.cmet.2024.01.015.
99. Fu X, Zhao Y, Cui X, Huang S, Lv Y, Li C, et al. Cxcl9 modulates aging associated microvascular metabolic and angiogenic dysfunctions in subcutaneous adipose tissue. *Angiogenesis.* (2025) 28:17. doi: 10.1007/s10456-025-09970-y
100. Liu X, Zhao L, Chen Y, Gao Y, Tian Q, Son JS, et al. Obesity induces adipose fibrosis and collagen cross-linking through suppressing AMPK and enhancing lysyl oxidase expression. *Biochim Biophys Acta Mol Basis Dis.* (2022) 1868:166454. doi: 10.1016/j.bbdis.2022.166454
101. Gonzalez Porras MA, Stojkova K, Vaicik MK, Pelowe A, Goddi A, Carmona A, et al. Integrins and extracellular matrix proteins modulate adipocyte thermogenic capacity. *Sci Rep.* (2021) 11:5442. doi: 10.1038/s41598-021-84828-z
102. Furuuchi R, Shimizu I, Yoshida Y, Katsuumi G, Suda M, Kubota Y, et al. Endothelial SIRT-1 has a critical role in the maintenance of capillarization in brown adipose tissue. *iScience.* (2022) 25:105424. doi: 10.1016/j.isci.2022.105424
103. Shi Y, Huang X, Zeng Y, Zhai M, Yao H, Liu C, et al. Endothelial TET2 regulates the white adipose browning and metabolism via fatty acid oxidation in obesity. *Redox Biol.* (2024) 69:103013. doi: 10.1016/j.redox.2023.103013
104. Waters DL, Aguirre L, Gurney B, Sinacore DR, Fowler K, Gregori G, et al. Effect of aerobic or resistance exercise, or both, on intermuscular and visceral fat and physical and metabolic function in older adults with obesity while dieting. *J Gerontol A Biol Sci Med Sci.* (2022) 77:131–9. doi: 10.1093/geronl/ab111
105. Colletuori G, Viola V, Bathina S, Armamento-Villareal R, Qualls C, Giordano A, et al. Effect of aerobic or resistance exercise, or both on insulin secretion, ciliary neurotrophic factor, and insulin-like growth factor-1 in dieting older adults with obesity. *Clin Nutr.* (2025) 51:50–62. doi: 10.1016/j.clnu.2025.05.016
106. Bürki C, Tian C, Westerman K, Patel C. Objectively and subjectively measured physical activity and their associations with cardiometabolic risk in the UK biobank: retrospective cohort study. *JMIR Mhealth Uhealth.* (2025) 13:e54820. doi: 10.2196/54820
107. Martinez-Tellez B, Sanchez-Delgado G, Acosta FM, Alcantara JMA, Amaro-Gahete FJ, Martinez-Avila WD, et al. No evidence of brown adipose tissue activation after 24 weeks of supervised exercise training in young sedentary adults in the ACTIBATE randomized controlled trial. *Nat Commun.* (2022) 13:5259. doi: 10.1038/s41467-022-32502-x
108. Ahn C, Zhang T, Yang G, Rode T, Varshney P, Ghayur SJ, et al. Years of endurance exercise training remodel abdominal subcutaneous adipose tissue in adults with overweight or obesity. *Nat Metab.* (2024) 6:1819–36. doi: 10.1038/s42255-024-01103-x
109. Vamvini M, Nigro P, Caputo T, Stanford KI, Hirshman MF, Middelbeek RJW, et al. Exercise training and cold exposure trigger distinct molecular adaptations to inguinal white adipose tissue. *Cell Rep.* (2024) 43:114481. doi: 10.1016/j.celrep.2024.114481
110. Nirengi S, Stanford K. Brown adipose tissue and aging: a potential role for exercise. *Exp Gerontol.* (2023) 178:112218. doi: 10.1016/j.exger.2023.112218
111. Cui X, Liu H, Liu Y, Yu Z, Wang D, Wei W, et al. The role and mechanisms of myokines in sarcopenia: new intervention strategies for the challenges of aging. *Front Med.* (2025) 12:1665708. doi: 10.3389/fmed.2025.1665708
112. Ito R, Xie S, Tumenjargal M, Sugahara Y, Yang C, Takahashi H, et al. Mitochondrial biogenesis in white adipose tissue mediated by JMJD1A-PGC-1 axis limits age-related metabolic disease. *iScience.* (2024) 27:109398. doi: 10.1016/j.isci.2024.109398
113. Checa-Ros A, D'Marco L. Molecular mechanisms linking adipose tissue browning to reduced cardiovascular risk. *Atherosclerosis.* (2025) 411:120564. doi: 10.1016/j.atherosclerosis.2025.120564
114. Bonfante ILP, Monfort-Pires M, Duft RG, da Silva Mateus KC, de Lima Júnior JC, Dos Santos Trombeta JC, et al. Combined training increases thermogenic fat activity in patients with overweight and type 2 diabetes. *Int J Obes.* (2022) 46:1145–54. doi: 10.1038/s41366-022-01086-3
115. Ahn C, Ryan BJ, Schleh MW, Varshney P, Ludzki AC, Gillen JB, et al. Exercise training remodels subcutaneous adipose tissue in adults with obesity even without weight loss. *J Physiol.* (2022) 600:2127–46. doi: 10.1113/jp282371
116. Becher T, Palanisamy S, Kramer DJ, Eljalby M, Marx SJ, Wibmer AG, et al. Brown adipose tissue is associated with cardiometabolic health. *Nat Med.* (2021) 27:58–65. doi: 10.1038/s41591-020-1126-7
117. Bloch-Ibenfeldt M, Theil Gates A, Karlog K, Demnitz N, Kjaer M, Boraxbekk CJ. Heavy resistance training at retirement age induces 4-year lasting beneficial effects in muscle strength: a long-term follow-up of an RCT. *BMJ Open Sport Exerc Med.* (2024) 10:e001899. doi: 10.1136/bmjsem-2024-001899
118. Newman JE, King I, Flemming N, Broadhouse KM, Buhmann R, Rose GL, et al. The acute response of irisin to resistance and endurance exercise at both lower and higher intensities in healthy older adults. *Exp Gerontol.* (2025) 209:112850. doi: 10.1016/j.exger.2025.112850
119. Makiel K, Suder A, Targosz A, Maciejczyk M, Haim A. Effect of exercise interventions on Irisin and interleukin-6 concentrations and indicators of carbohydrate metabolism in males with metabolic syndrome. *J Clin Med.* (2023) 12:369. doi: 10.3390/jcm12010369
120. Goeder D, Kröppl JM, Angst T, Hanssen H, Hauser C, Infanger D, et al. VasuFit: aerobic exercise improves endothelial function independent of cardiovascular risk: a randomized-controlled trial. *Atherosclerosis.* (2024) 399:118631. doi: 10.1016/j.atherosclerosis.2024.118631
121. Monsegue AP, Betz MW, Aydeniz H, Sluifsmans WE, van Loon LJC, Snijders T, et al. Aerobic exercise training improves blood glucose homeostasis in healthy older adults. *Exp Gerontol.* (2025) 212:112940. doi: 10.1016/j.exger.2025.112940
122. Lino Rodrigues K, Vieira Dias Da Silva V, Nunes Goulart da Silva Pereira E, Rangel Silveiras R, Peres de Araujo B, Eduardo Ilaquita Flores E, et al. Aerobic exercise training improves microvascular function and oxidative stress parameters in diet-induced type 2 diabetic mice. *Diabetes Metab Syndr Obes.* (2022) 15:2991–3005. doi: 10.2147/dmso.S365496
123. Tsuzuki T, Yoshihara T, Ichinoseki-Sekine N, Kobayashi H, Negishi T, Yukawa K, et al. Exercise training improves obesity-induced inflammatory signaling in rat brown adipose tissue. *Biochem Biophys Res.* (2022) 32:101398. doi: 10.1016/j.bbrep.2022.101398
124. Montgomery MK, De Nardo W, Watt MJ. Exercise training induces depot-specific remodeling of protein secretion in skeletal muscle and adipose tissue of obese male mice. *Am J Physiol Endocrinol Metab.* (2023) 325:E227–38. doi: 10.1152/ajpendo.00178.2023
125. Boström P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, et al. A PGC1- $\alpha$ -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature.* (2012) 481:463–8. doi: 10.1038/nature10777
126. Zhang Y, Li R, Meng Y, Li S, Donelan W, Zhao Y, et al. Irisin stimulates browning of white adipocytes through mitogen-activated protein kinase p38 MAP kinase and ERK MAP kinase signaling. *Diabetes.* (2014) 63:514–25. doi: 10.2337/db13-1106
127. Dai J, Zhao Y, Chen Y, Jiang Y, Sun R, Tang X, et al. Irisin reverses high-fat diet-induced metabolic dysfunction via activation of brown adipose tissue in mice. *Int J Obes.* (2025) 49:1066–75. doi: 10.1038/s41366-025-01739-z
128. Rioux B, Paudel Y, Thomson A, Peskett L, Sénéchal M. An examination of exercise intensity and its impact on the acute release of irisin across obesity status: a randomized controlled crossover trial. *Appl Physiol Nutr Metab.* (2024) 49:1712–28. doi: 10.1139/apnm-2024-0091
129. De Menech F, de Souza LV, Brioschi ML, Franco MDC. Emerging evidence for the opposite role of circulating irisin levels and brown adipose tissue activity measured by infrared thermography in anthropometric and metabolic profile during childhood. *J Therm Biol.* (2021) 99:103010. doi: 10.1016/j.jtherbio.2021.103010
130. Ma Y, Zheng J, Yu M, Zheng Z, Yang F, Liu Y. Effects of combined aerobic and resistance exercise on sarcopenia in elderly patients with type 2 diabetes mellitus. *J Endocrinol.* (2025) 267:e250275. doi: 10.1530/joe-25-0275
131. Trinh B, Peletier M, Simonsen C, Plomgaard P, Karstoft K, Klarlund Pedersen B, et al. Blocking endogenous IL-6 impairs mobilization of free fatty acids during rest and exercise in lean and obese men. *Cell Rep Med.* (2021) 2:100396. doi: 10.1016/j.xcrm.2021.100396
132. Izumi-Mishima Y, Tsutsumi R, Shiuchi T, Fujimoto S, Taniguchi M, Sugiuchi M, et al. Brown adipose tissue and skeletal muscle coordinately contribute to thermogenesis in mice. *Elife.* (2025) 13:R99982. doi: 10.7554/eLife.99982
133. Stanford KI, Middelbeek RJ, Townsend KL, An D, Nygaard EB, Hitchcox KM, et al. Brown adipose tissue regulates glucose homeostasis and insulin sensitivity. *J Clin Invest.* (2013) 123:215–23. doi: 10.1172/jci62308
134. Rao RR, Long JZ, White JB, Svensson KJ, Lou J, Lokurkar I, et al. Meteorin-like is a hormone that regulates immune-adipose interactions to increase beige fat thermogenesis. *Cell.* (2014) 157:1279–91. doi: 10.1016/j.cell.2014.03.065
135. Javaid HMA, Sahar NE, Zhuge DL, Huh JY. Exercise inhibits NLRP3 inflammasome activation in obese mice via the anti-inflammatory effect of meteorin-like. *Cells.* (2021) 10:3480. doi: 10.3390/cells10123480
136. Garcia-Beltran C, Navarro-Gascon A, López-Bermejo A, Quesada-López T, de Zegher F, Ibáñez L, et al. Meteorin-like levels are associated with active brown adipose tissue in early infancy. *Front Endocrinol.* (2023) 14:1136245. doi: 10.3389/fendo.2023.1136245
137. Wang Y, Yuan J, Liu H, Chen J, Zou J, Zeng X, et al. Elevated meteorin-like protein from high-intensity interval training improves heart function via AMPK/HDAC4 pathway. *Genes Dis.* (2024) 11:101100. doi: 10.1016/j.gendis.2023.101100
138. Hoekx CA, Martinez-Tellez B, Straat ME, Verkleij MMA, Kemmeren M, Kooijman S, et al. Cold exposure increases circulating fibroblast growth factor 21 in the evening in males and females. *Endocr Connect.* (2024) 13:e240074. doi: 10.1530/ec-24-0074
139. Peterson M, Richardson KA, Funderburk L. Effect of exercise on fibroblast growth factor 21 levels in healthy males and females. *PLoS One.* (2025) 20:e0321738. doi: 10.1371/journal.pone.0321738
140. Abu-Odeh M, Zhang Y, Reilly SM, Ebadat N, Keinan O, Valentine JM, et al. FGF21 promotes thermogenic gene expression as an autocrine factor in adipocytes. *Cell Rep.* (2021) 35:109331. doi: 10.1016/j.celrep.2021.109331
141. Jurado-Fasoli L, Sanchez-Delgado G, Di X, Yang W, Kohler I, Villarroya F, et al. Cold-induced changes in plasma signaling lipids are associated with a healthier

- cardiometabolic profile independently of brown adipose tissue. *Cell Rep Med.* (2024) 5:101387. doi: 10.1016/j.xcrm.2023.101387
142. Hollstein T, Vinales K, Chen KY, Cypess AM, Basolo A, Schlögl M, et al. Reduced brown adipose tissue activity during cold exposure is a metabolic feature of the human thrifty phenotype. *Metabolism.* (2021) 117:154709. doi: 10.1016/j.metabol.2021.154709
143. Miniewska K, Maliszewska K, Pietrowska K, Godzienie J, Labieniec Ł, Mojsak M, et al. PET/MRI-evaluated activation of brown adipose tissue via cold exposure impacts lipid metabolism. *Metabolites.* (2022) 12:456. doi: 10.3390/metabo12050456
144. Wang GX, Zhao XY, Meng ZX, Kern M, Dietrich A, Chen Z, et al. The brown fat-enriched secreted factor Nrg4 preserves metabolic homeostasis through attenuation of hepatic lipogenesis. *Nat Med.* (2014) 20:1436–43. doi: 10.1038/nm.3173
145. Chen Z, Wang GX, Ma SL, Jung DY, Ha H, Altamimi T, et al. Nrg4 promotes fuel oxidation and a healthy adipokine profile to ameliorate diet-induced metabolic disorders. *Mol Metab.* (2017) 6:863–72. doi: 10.1016/j.molmet.2017.03.016
146. Liu Y, Chen M. Neuregulin 4 as a novel adipokine in energy metabolism. *Front Physiol.* (2022) 13:1106380. doi: 10.3389/fphys.2022.1106380
147. Saeidi A, Shishvan SR, Soltani M, Tarazi F, Doyle-Baker PK, Shahrbanian S, et al. Differential effects of exercise programs on neuregulin 4, body composition and cardiometabolic risk factors in men with obesity. *Front Physiol.* (2021) 12:797574. doi: 10.3389/fphys.2021.797574
148. Fang P, She Y, Yu M, Min W, Shang W, Zhang Z. Adipose-Muscle crosstalk in age-related metabolic disorders: the emerging roles of adipo-myokines. *Ageing Res Rev.* (2023) 84:101829. doi: 10.1016/j.arr.2022.101829
149. Chou TJ, Lin LY, Lu CW, Hsu YJ, Huang CC, Huang KC. Effects of aerobic, resistance, and high-intensity interval training on thermogenic gene expression in white adipose tissue in high fat diet induced obese mice. *Obes Res Clin Pract.* (2024) 18:64–72. doi: 10.1016/j.orcp.2024.01.003
150. Chen X, Li Y, Zhang J, Huang W, Su J, Zhang J. Lactate coordinated with exercise promoted the browning of inguinal white adipose tissue. *J Physiol Biochem.* (2024) 80:303–15. doi: 10.1007/s13105-023-01004-9
151. Chen W, Liu Y, Liu J, Chen Y, Wang X. Acute exercise promotes WAT browning by remodeling mRNA m(6)A methylation. *Life Sci.* (2025) 361:123269. doi: 10.1016/j.lfs.2024.123269
152. Fu X, Murakami M, Hashimoto O, Matsui T, Funaba M. Regulatory mechanisms underlying interleukin-6 expression in murine brown adipocytes. *Cell Biochem Funct.* (2024) 42:e3915. doi: 10.1002/cbf.3915
153. Mendez-Gutierrez A, Aguilera CM, Cereijo R, Osuna-Prieto FJ, Martinez-Tellez B, Rico MC, et al. Cold exposure modulates potential brown adipokines in humans, but only FGF21 is associated with brown adipose tissue volume. *Obesity.* (2024) 32:560–70. doi: 10.1002/oby.23970
154. Chamorro R, Bertozzi B, Backhaus J, Iwen KA, Rademacher L, Meyhoefer S, et al. Acute cold exposure improves glucose tolerance and induces beta-cell secretion response linked to lipid utilization in young male with obesity. *Exp Clin Endocrinol Diabetes.* (2025) 133:532–40. doi: 10.1055/a-2730-1689
155. Pinckard KM, Shettigar VK, Wright KR, Abay E, Baer LA, Vidal P, et al. A novel endocrine role for the BAT-released lipokine 12,13-diHOME to mediate cardiac function. *Circulation.* (2021) 143:145–59. doi: 10.1161/circulationaha.120.049813
156. Sellami M, Almuraikhy S, Naja K, Anwardeen N, Al-Amri HS, Prince MS, et al. Eight weeks of aerobic exercise, but not four, improves insulin sensitivity and cardiovascular performance in young women. *Sci Rep.* (2025) 15:1991. doi: 10.1038/s41598-025-86306-2
157. Xu X, Ying Z, Cai M, Xu Z, Li Y, Jiang SY, et al. Exercise ameliorates high-fat diet-induced metabolic and vascular dysfunction, and increases adipocyte progenitor cell population in brown adipose tissue. *Am J Physiol Regul Integr Comp Physiol.* (2011) 300:R1115–25. doi: 10.1152/ajpregu.00806.2010
158. Rao Z, Geng X, Huang P, Wei Q, Liu S, Qu C, et al. Housing temperature influences exercise-induced glucose regulation and expression of exerkines in mice. *Exp Physiol.* (2025) 110:1099–113. doi: 10.1113/ep092319
159. Tanimura R, Kobayashi L, Shirai T, Takemasa T. Effects of exercise intensity on white adipose tissue browning and its regulatory signals in mice. *Physiol Rep.* (2022) 10:e15205. doi: 10.14814/phy2.15205
160. Motiani P, Teuho J, Saari T, Virtanen KA, Honkala SM, Middelbeek RJ, et al. Exercise training alters lipoprotein particles independent of brown adipose tissue metabolic activity. *Obes Sci Pract.* (2019) 5:258–72. doi: 10.1002/osp4.330
161. Jurado-Fasoli L, Di X, Sanchez-Delgado G, Yang W, Osuna-Prieto FJ, Ortiz-Alvarez L, et al. Acute and long-term exercise differently modulate plasma levels of oxylipins, endocannabinoids, and their analogues in young sedentary adults: a sub-study and secondary analyses from the ACTIBATE randomized controlled-trial. *EBioMedicine.* (2022) 85:104313. doi: 10.1016/j.ebiom.2022.104313
162. Richard G, Blondin DP, Syed SA, Rossi L, Fontes ME, Fortin M, et al. High-fructose feeding suppresses cold-stimulated brown adipose tissue glucose uptake independently of changes in thermogenesis and the gut microbiome. *Cell Rep Med.* (2022) 3:100742. doi: 10.1016/j.xcrm.2022.100742
163. Cordingley DM, Anderson JE, Cornish SM. Resting systemic Irisin concentrations are lower in older versus younger males after 12 weeks of resistance-exercise training while Apelin and IL-15 concentrations were increased in the whole cohort. *Muscles.* (2024) 3:202–11. doi: 10.3390/muscles3030018
164. Ataeinosrat A, Saeidi A, Abednatanzi H, Rahmani H, Dalooi AA, Pashaei Z, et al. Intensity dependent effects of interval resistance training on myokines and cardiovascular risk factors in males with obesity. *Front Endocrinol.* (2022) 13:895512. doi: 10.3389/fendo.2022.895512
165. Jiang G, Qin S, Yan B, Girard O. Metabolic and hormonal responses to acute high-load resistance exercise in normobaric hypoxia using a saturation clamp. *Front Physiol.* (2024) 15:1445229. doi: 10.3389/fphys.2024.1445229
166. Tsai SH, Cheng HC, Liu HW. Supervised high-load resistance training for improving muscle strength and quality in prediabetic older adults: a pilot randomized controlled trial. *Geriatr Nurs.* (2022) 48:350–5. doi: 10.1016/j.gerinurse.2022.11.001
167. Hu Y, Wu Y, Wang C, Jin Q, Chen X. Synergistic impact of aerobic exercise and resveratrol on white adipose tissue browning in obese rats: mechanistic exploration and biological insights. *Metabolites.* (2025) 15:331. doi: 10.3390/metabo15050331
168. Tanahashi K, Kato D, Kojima T, Tsuda T. Low dose of curcumin combined with exercise synergistically induces beige adipocyte formation in mice. *J Nutr Sci Vitaminol.* (2023) 69:299–304. doi: 10.3177/jnsv.69.299
169. Rodak K, Kokot I, Kratz EM. Caffeine as a factor influencing the functioning of the human body-friend or foe? *Nutrients.* (2021) 13:3088. doi: 10.3390/nu13093088
170. Hamad AKS. Caffeine and arrhythmias: a critical analysis of cardiovascular responses and arrhythmia susceptibility. *J Saudi Heart Assoc.* (2024) 36:335–48. doi: 10.37616/2212-5043.1402
171. Straat ME, Martinez-Tellez B, Sardjoe Mishre A, Verkleij MMA, Kemmeren M, Pelsma ICM, et al. Cold-induced thermogenesis shows a diurnal variation that unfolds differently in males and females. *J Clin Endocrinol Metab.* (2022) 107:1626–35. doi: 10.1210/clinem/dgac094
172. Demirli A, Ulupınar S, Terzi M, Özbay S, Özkara AB, Gençoğlu C, et al. Synergistic effects of green tea extract and ginger supplementation on endurance performance and thermal perception in normothermic and cold environments: a randomized, placebo-controlled, double-blind crossover trial. *Nutrients.* (2025) 17:2949. doi: 10.3390/nu17182949
173. Petran EM, Periferakis A, Troumpala L, Periferakis AT, Scheau AE, Badarau IA, et al. Capsaicin: emerging pharmacological and therapeutic insights. *Curr Issues Mol Biol.* (2024) 46:7895–943. doi: 10.3390/cimb46080468
174. Abdillah AM, Yun JW. Capsaicin induces ATP-dependent thermogenesis via the activation of TRPV1/β3-AR/α1-AR in 3T3-L1 adipocytes and mouse model. *Arch Biochem Biophys.* (2024) 755:109975. doi: 10.1016/j.abb.2024.109975
175. Irandoost P, Lotfi Yagin N, Namazi N, Keshkar A, Farsi F, Mesri Alamdari N, et al. The effect of Capsaicinoids or Capsinoids in red pepper on thermogenesis in healthy adults: a systematic review and meta-analysis. *Phytother Res.* (2021) 35:1358–77. doi: 10.1002/ptr.6897
176. Zhang W, Zhang Q, Wang L, Zhou Q, Wang P, Qing Y, et al. The effects of capsaicin intake on weight loss among overweight and obese subjects: a systematic review and meta-analysis of randomised controlled trials. *Br J Nutr.* (2023) 130:1645–56. doi: 10.1017/s0007114523000697
177. Fagundes GBP, Rodrigues A, Martins LB, Monteze NM, Correia M, Teixeira AL, et al. Acute effects of dry extract of ginger on energy expenditure in eutrophic women: a randomized clinical trial. *Clin Nutr ESPEN.* (2021) 41:168–74. doi: 10.1016/j.clnesp.2020.10.001
178. Evans C, Kalman D, Jiannine L, Ricci T, Byers P, Pereira F, et al. The acute metabolic impacts of kamferia parviflora extract in healthy men: a randomized, double-blind, proof-of-concept study. *Cureus.* (2025) 17:e81561. doi: 10.7759/cureus.81561
179. Bolat E, Sarıtaş S, Duman H, Eker F, Akdaşçı E, Karav S, et al. Polyphenols: secondary metabolites with a biological impression. *Nutrients.* (2024) 16:2550. doi: 10.3390/nu16152550
180. Vannuchi N, Pisani L. PGC-1α activation by polyphenols: a pathway to thermogenesis. *Mol Nutr Food Res.* (2025) 69:e70072. doi: 10.1002/mnfr.70072
181. Gómez-García I, Fernández-Quintela A, Portillo MP, Trepiana J. Changes in brown adipose tissue induced by resveratrol and its analogue pterostilbene in rats fed with a high-fat high-fructose diet. *J Physiol Biochem.* (2024) 80:627–37. doi: 10.1007/s13105-023-00985-x
182. Santos ACC, Amaro LBR, Batista Jorge AH, Lelis SF, Lelis DF, Guimarães ALS, et al. Curcumin improves metabolic response and increases expression of thermogenesis-associated markers in adipose tissue of male offspring from obese dams. *Mol Cell Endocrinol.* (2023) 563:111840. doi: 10.1016/j.mce.2022.111840
183. Hui S, Liu Y, Huang L, Zheng L, Zhou M, Lang H, et al. Resveratrol enhances brown adipose tissue activity and white adipose tissue browning in part by regulating bile acid metabolism via gut microbiota remodeling. *Int J Obes.* (2020) 44:1678–90. doi: 10.1038/s41366-020-0566-y
184. Yan H, Shao M, Lin X, Peng T, Chen C, Yang M, et al. Resveratrol stimulates brown of white adipose via regulating ERK/DRP1-mediated mitochondrial fission and improves systemic glucose homeostasis. *Endocrine.* (2025) 87:144–58. doi: 10.1007/s12020-024-04008-7
185. Khamis MM, Moselhy SS, Rihan S. Role of trans-resveratrol in ameliorating biochemical and molecular alterations in obese rats induced by a high fructose/fat diet. *Sci Rep.* (2025) 15:7879. doi: 10.1038/s41598-025-91027-7

186. Sikur N, Böröczky C, Paszternák A, Gyöngyössi R, Szökő É, Varga K, et al. Resveratrol and its derivatives diminish lipid accumulation in adipocytes in vitro-mechanism of action and structure-activity relationship. *Nutrients*. (2024) 16:3869. doi: 10.3390/nu16223869
187. Kononova YA, Tuchina TP, Babenko AY. Brown and beige adipose tissue: one or different targets for treatment of obesity and obesity-related metabolic disorders? *Int J Mol Sci*. (2024) 25:13295. doi: 10.3390/ijms252413295
188. Pang J, Raka F, Heirali AA, Shao W, Liu D, Gu J, et al. Resveratrol intervention attenuates chylomicron secretion via repressing intestinal FXR-induced expression of scavenger receptor SR-B1. *Nat Commun*. (2023) 14:2656. doi: 10.1038/s41467-023-38259-1
189. Rondanelli M, Riva A, Petrangolini G, Allegrini P, Perna S, Faliva MA, et al. Effect of acute and chronic dietary supplementation with green tea catechins on resting metabolic rate, energy expenditure and respiratory quotient: a systematic review. *Nutrients*. (2021) 13:644. doi: 10.3390/nu13020644
190. Van Schaik L, Kettle C, Green R, Irving HR, Rathner JA. Effects of caffeine on brown adipose tissue thermogenesis and metabolic homeostasis: a review. *Front Neurosci*. (2021) 15:621356. doi: 10.3389/fnins.2021.621356
191. Wilasrusmee KT, Sitticharoon C, Keadkraichaiwat I, Maikaew P, Pongwattanapakin K, Chatree S, et al. Epigallocatechin gallate enhances sympathetic heart rate variability and decreases blood pressure in obese subjects: a randomized control trial. *Sci Rep*. (2024) 14:21628. doi: 10.1038/s41598-024-72269-3
192. Zou T, Li S, Wang B, Wang Z, Liu Y, You J. Curcumin improves insulin sensitivity and increases energy expenditure in high-fat-diet-induced obese mice associated with activation of FNDC5/irisin. *Nutrition*. (2021) 90:111263. doi: 10.1016/j.nut.2021.111263
193. Chen X, Zou J, Cao Z, Hong T, Zhang H, Yang J, et al. Curcumin inhibits bisphenol A-induced fat mass gain by enhancing white adipose tissue browning via modulating gut microbiota-dependent bile acid metabolism in CD-1 Mice. *J Med Food*. (2025) 28:1244–55. doi: 10.1177/1096620x251383432
194. Lamichhane G, Godsey TJ, Liu J, Franks R, Zhang G, Emerson SR, et al. Twelve-week curcumin supplementation improves glucose homeostasis and gut health in prediabetic older adults: a pilot, double-blind, placebo-controlled trial. *Nutrients*. (2025) 17:2164. doi: 10.3390/nu17132164
195. Yaikawong M, Jansarikit L, Jirawatnotai S, Chuengsamarn S. Curcumin extract improves beta cell functions in obese patients with type 2 diabetes: a randomized controlled trial. *Nutr J*. (2024) 23:119. doi: 10.1186/s12937-024-01022-3
196. Bertoncini-Silva C, Vlad A, Ricciarelli R, Giacomo Fassini P, Suen VMM, Zingg JM. Enhancing the bioavailability and bioactivity of curcumin for disease prevention and treatment. *Antioxidants*. (2024) 13:331. doi: 10.3390/antiox13030331
197. Jia M, Xu T, Xu YJ, Liu Y. Dietary fatty acids activate or deactivate brown and beige fat. *Life Sci*. (2023) 330:121978. doi: 10.1016/j.lfs.2023.121978
198. Trouwborst I, Gijbels A, Jardon KM, Siebelink E, Hul GB, Wanders L, et al. Cardiometabolic health improvements upon dietary intervention are driven by tissue-specific insulin resistance phenotype: a precision nutrition trial. *Cell Metab*. (2023) 35:71–83.e5. doi: 10.1016/j.cmet.2022.12.002
199. Zu Y, Pahlavani M, Ramalingam L, Jayarathne S, Andrade J, Scoggin S, et al. Temperature-Dependent Effects of Eicosapentaenoic Acid (EPA) on browning of subcutaneous adipose tissue in UCP1 knockout male mice. *Int J Mol Sci*. (2023) 24:8708. doi: 10.3390/ijms24108708
200. Yang X, Li X, Hu M, Huang J, Yu S, Zeng H, et al. EPA and DHA differentially improve insulin resistance by reducing adipose tissue inflammation-targeting GPR120/PPAR $\gamma$  pathway. *J Nutr Biochem*. (2024) 130:109648. doi: 10.1016/j.jnutbio.2024.109648
201. Yamazaki T, Li D, Ikaga R. Fish oil increases diet-induced thermogenesis in mice. *Mar Drugs*. (2021) 19:278. doi: 10.3390/md19050278
202. Rust BM, Raatz SK, Casperman SL, Duke SE, Picklo MJ. Dietary fat chain length, saturation, and PUFA source acutely affect diet-induced thermogenesis but not satiety in adults in a randomized, crossover trial. *Nutrients*. (2021) 13:2615. doi: 10.3390/nu13082615
203. Rodríguez-García C, Sánchez-Quesada C, Algarra I, Gaforio JJ. Differential immunometabolic effects of high-fat diets containing coconut, sunflower, and extra virgin olive oils in female mice. *Mol Nutr Food Res*. (2022) 66:e2200082. doi: 10.1002/mnfr.202200082
204. Virani SS, Morris PB, Agarwala A, Ballantyne CM, Birtcher KK, Kris-Etherton PM, et al. 2021 ACC expert consensus decision pathway on the management of ASCVD risk reduction in patients with persistent hypertriglyceridemia: a report of the American College of cardiology solution set oversight committee. *J Am Coll Cardiol*. (2021) 78:960–93. doi: 10.1016/j.jacc.2021.06.011
205. Stanford KI, Lynes MD, Takahashi H, Baer LA, Arts PJ, May FJ, et al. 12,13-diHOME: an exercise-induced lipokine that increases skeletal muscle fatty acid uptake. *Cell Metab*. (2018) 27:1111–20.e3. doi: 10.1016/j.cmet.2018.03.020
206. Wang B, Fu X, Liang X, Deavila JM, Wang Z, Zhao L, et al. Retinoic acid induces white adipose tissue browning by increasing adipose vasculature and inducing beige adipogenesis of PDGFR $\alpha$ (+) adipose progenitors. *Cell Discov*. (2017) 3:17036. doi: 10.1038/celldisc.2017.36
207. Rubinow KB, Zhong G, Czuba LC, Chen JY, Williams E, Parr Z, et al. Evidence of depot-specific regulation of all-trans-retinoic acid biosynthesis in human adipose tissue. *Clin Transl Sci*. (2022) 15:1460–71. doi: 10.1111/cts.13259
208. Cassim Bawa FN, Hu S, Gopaju R, Shiyab A, Mongan K, Xu Y, et al. Adipocyte retinoic acid receptor  $\alpha$  prevents obesity and steatohepatitis by regulating energy expenditure and lipogenesis. *Obesity*. (2024) 32:120–30. doi: 10.1002/oby.23929
209. Park JY, Ha ES, Lee J, Brun PJ, Kim Y, Chung SS, et al. The brown fat-specific overexpression of RBP4 improves thermoregulation and systemic metabolism by activating the canonical adrenergic signaling pathway. *Exp Mol Med*. (2025) 57:554–66. doi: 10.1038/s12276-025-01411-6
210. Han H, Zhang S, Wang M, Yi B, Zhao Y, Schroyen M, et al. Retinol metabolism signaling participates in microbiota-regulated fat deposition in obese mice. *J Nutr Biochem*. (2025) 136:109787. doi: 10.1016/j.jnutbio.2024.109787
211. Berry DC, Noy N. All-trans-retinoic acid represses obesity and insulin resistance by activating both peroxisome proliferation-activated receptor beta/delta and retinoic acid receptor. *Mol Cell Biol*. (2009) 29:3286–96. doi: 10.1128/mcb.01742-08
212. Lu S, Cao ZB. Interplay between Vitamin D and adipose tissue: implications for adipogenesis and adipose tissue function. *Nutrients*. (2023) 15:4832. doi: 10.3390/nu15224832
213. Ji L, Gupta M, Feldman BJ. Vitamin D regulates fatty acid composition in subcutaneous adipose tissue through Elovl3. *Endocrinology*. (2016) 157:91–7. doi: 10.1210/en.2015-1674
214. Amaro-Gahete FJ, Vázquez-Lorente H, Sanchez-Delgado G, Ruiz JR. Soluble alpha-klotho and 25-hydroxyvitamin D are not associated with brown adipose tissue metabolism in young healthy adults. *J Physiol Biochem*. (2025) 81:291–8. doi: 10.1007/s13105-025-01072-z
215. Cheung WW, Ding W, Hoffman HM, Wang Z, Hao S, Zheng R, et al. Vitamin D ameliorates adipose browning in chronic kidney disease cachexia. *Sci Rep*. (2020) 10:14175. doi: 10.1038/s41598-020-70190-z
216. Moro ML, Reis NG, Schavinski AZ, Neto JBC, Assis AP, Santos JR, et al. Perinatal vitamin D deficiency enhances brown adipose tissue thermogenesis in weanling rats. *Int J Mol Sci*. (2025) 26:4534. doi: 10.3390/ijms26104534
217. Zhou Y, Shu J, Zhao Y, Wu X, He Z, Lyu X, et al. Vitamin D(3) promotes white fat beige through IL-27/P38MAPK/PGC-1 $\alpha$  pathway. *Front Nutr*. (2025) 12:1661072. doi: 10.3389/fnut.2025.1661072
218. Choi SM, Lim SH, Lee HS, Choi G, Kim MJ, Kim H, et al. Coixol and Sinigrin from Coix lacryma-jobi L. and *Raphanus sativus* L. Promote fat browning in 3T3-L1 adipocytes. *Pharmaceuticals*. (2025) 18:1843. doi: 10.3390/ph18121843
219. Arianti R, Vinnai B, Györy F, Guba A, Csösz É, Kristóf E, et al. Availability of abundant thiamine determines efficiency of thermogenic activation in human neck area derived adipocytes. *J Nutr Biochem*. (2023) 119:109385. doi: 10.1016/j.jnutbio.2023.109385
220. Carrodeguas L, Kaidar-Person O, Szomstein S, Antozzi P, Rosenthal R. Preoperative thiamine deficiency in obese population undergoing laparoscopic bariatric surgery. *Surg Obes Relat Dis*. (2005) 1:517–22; discussion 22. doi: 10.1016/j.soard.2005.08.003
221. Flancbaum L, Belsley S, Drake V, Colarusso T, Tayler E. Preoperative nutritional status of patients undergoing Roux-en-Y gastric bypass for morbid obesity. *J Gastrointest Surg*. (2006) 10:1033–7. doi: 10.1016/j.gassur.2006.03.004
222. Nath A, Tran T, Shope TR, Koch TR. Prevalence of clinical thiamine deficiency in individuals with medically complicated obesity. *Nutr Res*. (2017) 37:29–36. doi: 10.1016/j.nutres.2016.11.012
223. Peterson LA, Cheskin LJ, Furtado M, Papas K, Schweitzer MA, Magnuson TH, et al. Malnutrition in bariatric surgery candidates: multiple micronutrient deficiencies prior to surgery. *Obes Surg*. (2016) 26:833–8. doi: 10.1007/s11695-015-1844-y
224. Pereira MJ, Andersson-Assarsson JC, Jacobson P, Kamble P, Taube M, Sjöholm K, et al. Human adipose tissue gene expression of solute carrier family 19 member 3 (SLC19A3); relation to obesity and weight-loss. *Obes Sci Pract*. (2022) 8:21–31. doi: 10.1002/osp4.541
225. Thornalley PJ, Babaei-Jadidi R, Al Ali H, Rabbani N, Antonysunil A, Larkin J, et al. High prevalence of low plasma thiamine concentration in diabetes linked to a marker of vascular disease. *Diabetologia*. (2007) 50:2164–70. doi: 10.1007/s00125-007-0771-4
226. Alaei Shahmiri F, Soares MJ, Zhao Y, Sherriff J. High-dose thiamine supplementation improves glucose tolerance in hyperglycemic individuals: a randomized, double-blind cross-over trial. *Eur J Nutr*. (2013) 52:1821–4. doi: 10.1007/s00394-013-0534-6
227. Hansen B, Larsson C, Wirén J, Hallgren J. Hypothermia and infection in Wernicke's encephalopathy. *Acta Med Scand*. (1984) 215:185–7. doi: 10.1111/j.0954-6820.1984.tb04991.x
228. Teh MR, Armitage AE, Drakesmith H. Why cells need iron: a compendium of iron utilisation. *Trends Endocrinol Metab*. (2024) 35:1026–49. doi: 10.1016/j.tem.2024.04.015
229. Ding X, Bian N, Wang J, Chang X, An Y, Wang G, et al. Serum ferritin levels are associated with adipose tissue dysfunction-related indices in obese adults. *Biol Trace Elem Res*. (2023) 201:636–43. doi: 10.1007/s12011-022-03198-3

230. Hinojosa-Moscoso A, Motger-Albertí A, De la Calle-Vargas E, Martí-Navas M, Biarnés C, Arrioriaga-Rodríguez M, et al. The longitudinal changes in subcutaneous abdominal tissue and visceral adipose tissue volumetries are associated with iron status. *Int J Mol Sci.* (2023) 24:4750. doi: 10.3390/ijms24054750
231. Yook JS, Thomas SS, Toney AM, You M, Kim YC, Liu Z, et al. Dietary iron deficiency modulates adipocyte iron homeostasis, adaptive thermogenesis, and obesity in C57BL/6 Mice. *J Nutr.* (2021) 151:2967–75. doi: 10.1093/jn/nxab222
232. Mai X, Liu Y, Fan J, Xiao L, Liao M, Huang Z, et al. Iron supplementation and iron accumulation promote adipocyte thermogenesis through PGC1 $\alpha$ -ATGL-mediated lipolysis. *J Biol Chem.* (2024) 300:107690. doi: 10.1016/j.jbc.2024.107690
233. Lu B, Guo S, Zhao J, Wang X, Zhou B. Adipose knockout of H-ferritin improves energy metabolism in mice. *Mol Metab.* (2024) 80:101871. doi: 10.1016/j.molmet.2024.101871
234. Wang X, Wu Q, Zhong M, Chen Y, Wang Y, Li X, et al. Adipocyte-derived ferroptotic signaling mitigates obesity. *Cell Metab.* (2025) 37:673–91.e7. doi: 10.1016/j.cmet.2024.11.010.
235. Qiu J, Zhang Z, Hu Y, Guo Y, Liu C, Chen Y, et al. Transferrin receptor levels and its rare variant are associated with human obesity. *J Diabetes.* (2024) 16:e13467. doi: 10.1111/1753-0407.13467
236. Alrifai R, Seo M, Karadsheh G, Mahendra FR, Demény M, Gyóry E, et al. Transferrin receptor 1-mediated iron uptake supports thermogenic activation in human cervical-derived adipocytes. *FEBS Lett.* (2026) [Online ahead of print]. doi: 10.1002/1873-3468.70312.
237. Lowe NM, Hall AG, Broadley MR, Foley J, Boy E, Bhutta ZA. Preventing and controlling zinc deficiency across the life course: a call to action. *Adv Nutr.* (2024) 15:100181. doi: 10.1016/j.advnut.2024.100181
238. Yokokawa H, Morita Y, Hamada I, Ohta Y, Fukui N, Makino N, et al. Demographic and clinical characteristics of patients with zinc deficiency: analysis of a nationwide Japanese medical claims database. *Sci Rep.* (2024) 14:2791. doi: 10.1038/s41598-024-53202-0
239. Gupta N, Carmichael MF. Zinc-induced copper deficiency as a rare cause of neurological deficit and anemia. *Cureus.* (2023) 15:e43856. doi: 10.7759/cureus.43856